

*Phenix workshop at the University of Nebraska Lincoln,  
April 30, 2026*

# Using Predicted models in Phenix (cryo-EM)



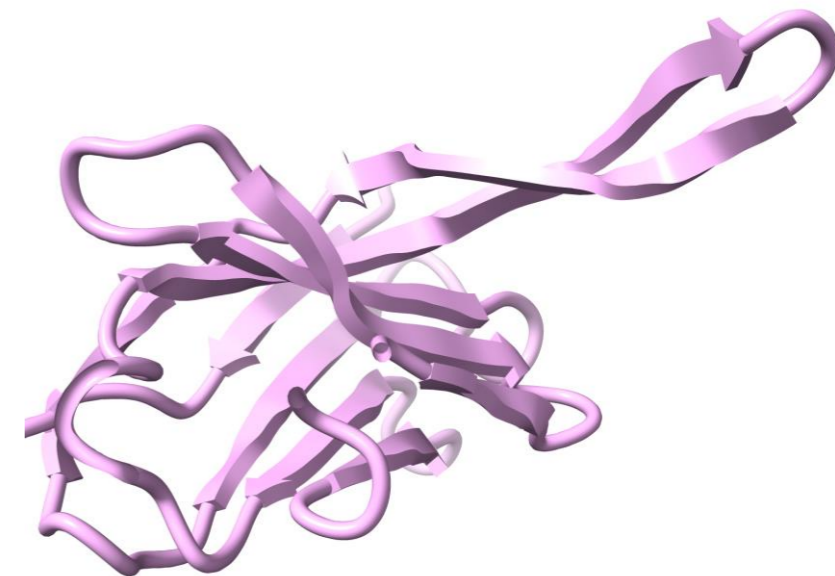
Dorothee Liebschner  
Lawrence Berkeley Laboratory

# Predicting models with AlphaFold



```
EVQLVESGGGLVQPGGSLRLSCAASGFTNIYSSSIHWVRQAPGKGLEWVAYI
.....F.....M.....Q.....
.....K.....Y.....L.....A.....
.....A.....V.....
.....A.....
.....L.....V.....E.....
.....A.....Q.....
```

Sequence  
Multiple sequence alignment



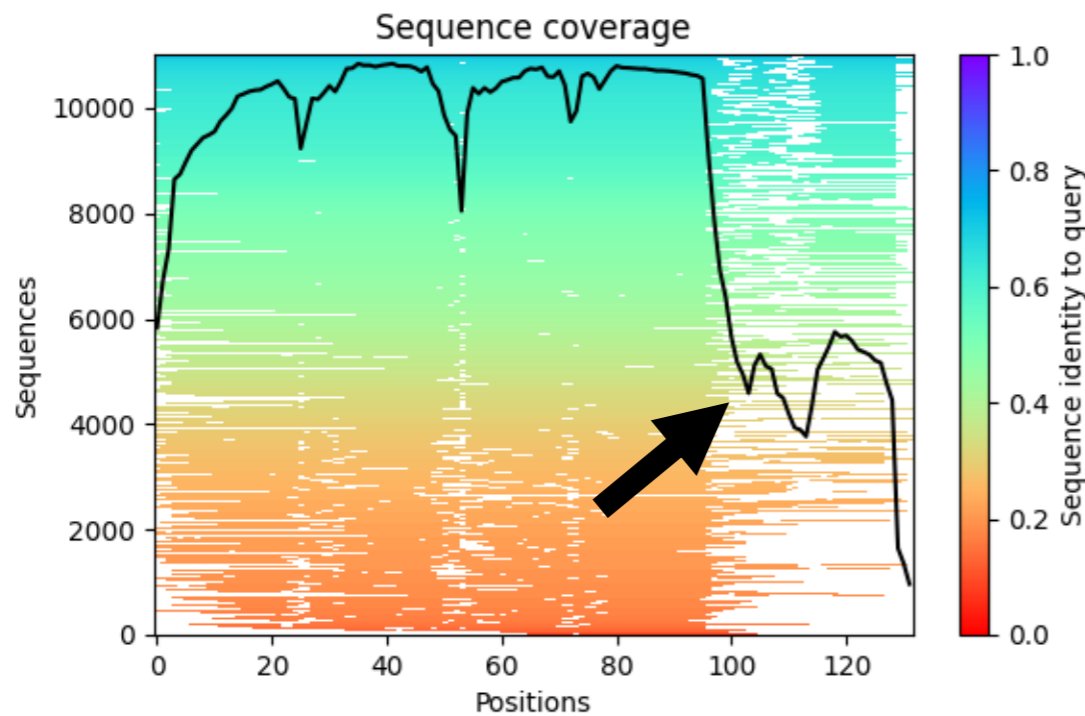
3D prediction

# Predictions come with confidence measures

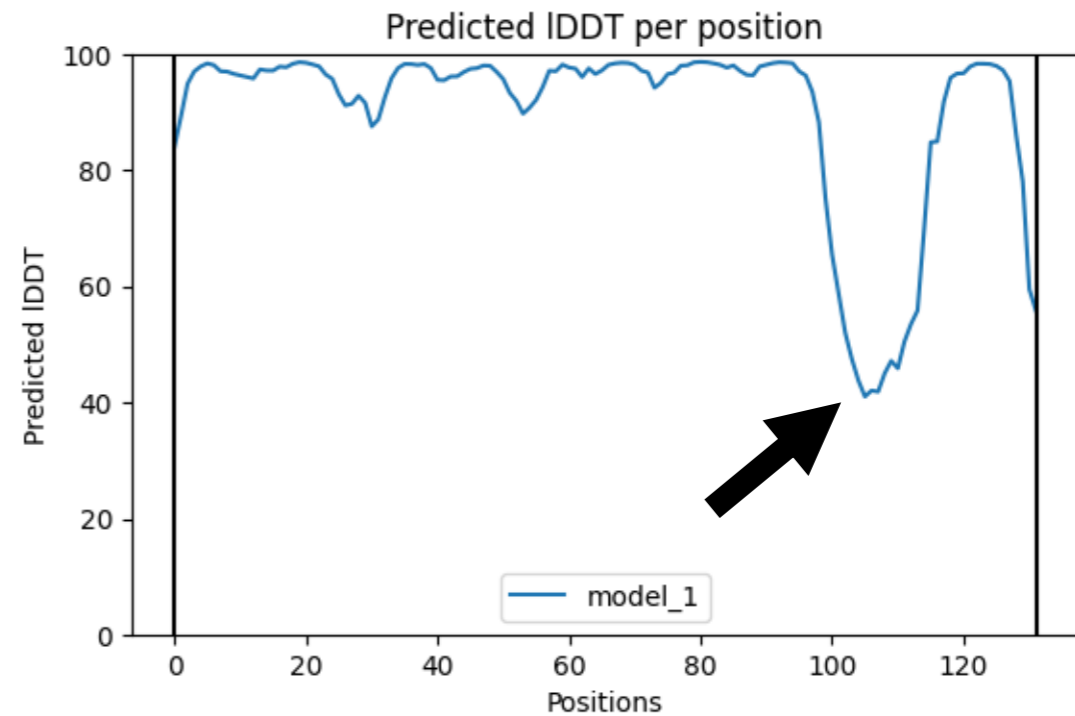
## 1. pLDDt (predicted Local Distance Difference Test)

- pLDDt identifies where errors are more likely.
- Per-residue confidence measure.
- Scales from 0 – 100 (pLDDt > 90: predicted with high accuracy).

Sequence coverage

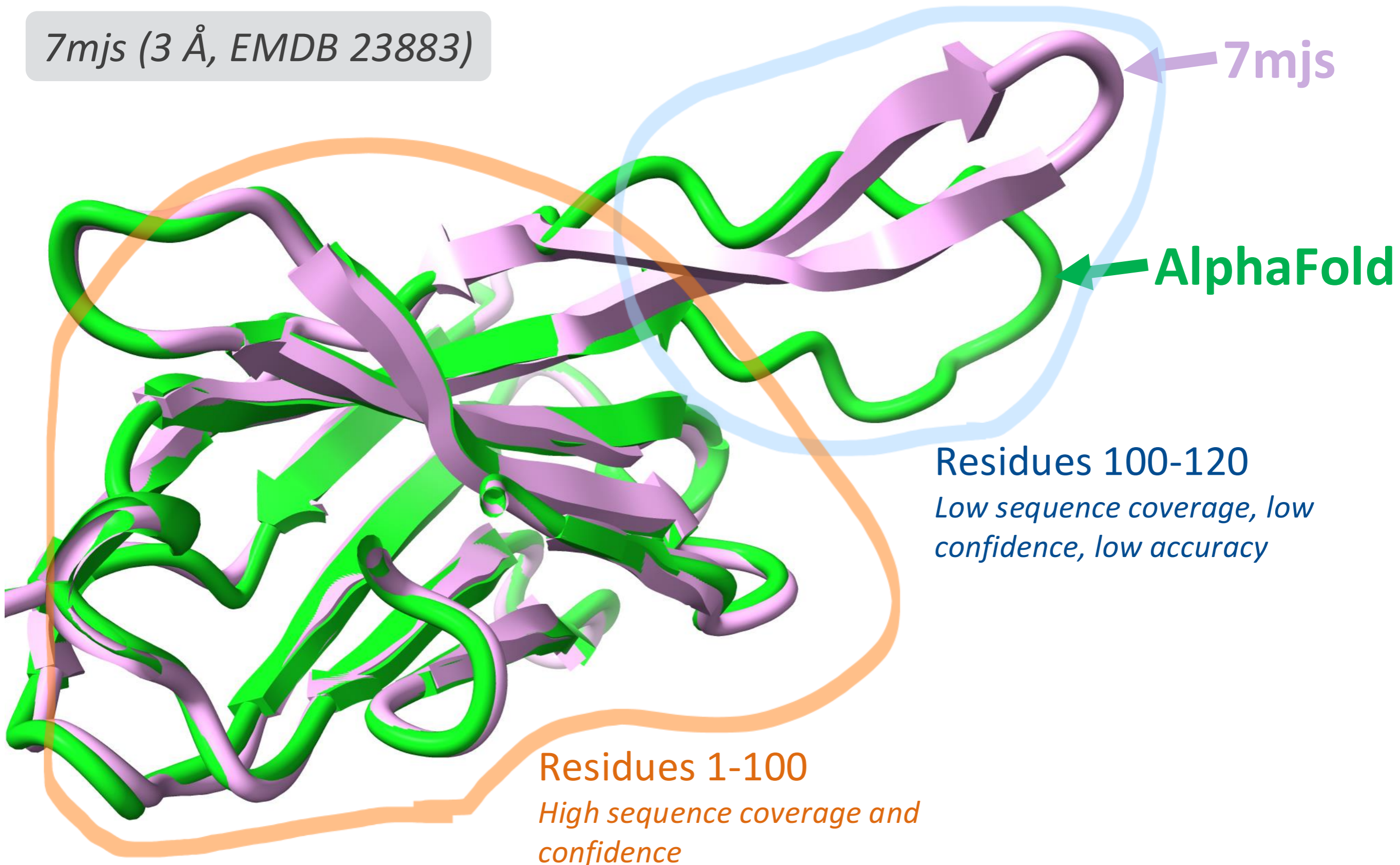


Confidence



# Predictions come with confidence measures

7mjs (3 Å, EMDB 23883)



# Predictions come with confidence measures

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## 1. pLDDt (predicted Local Distance Difference Test)

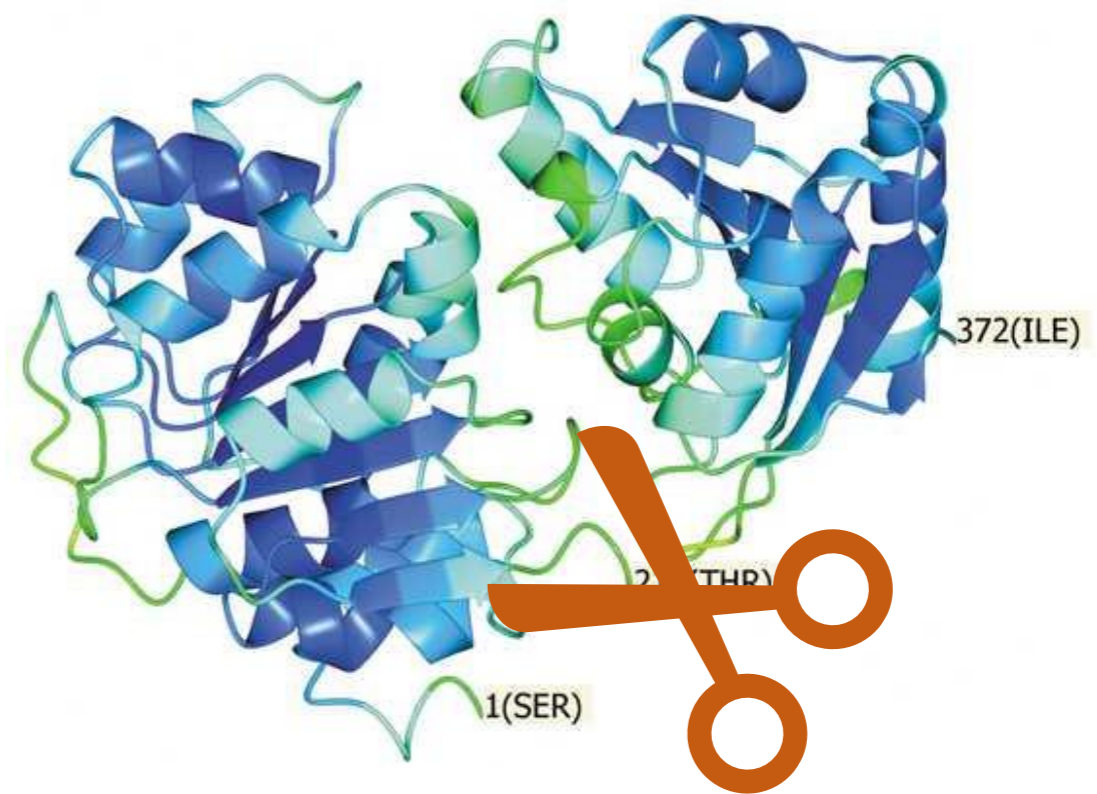
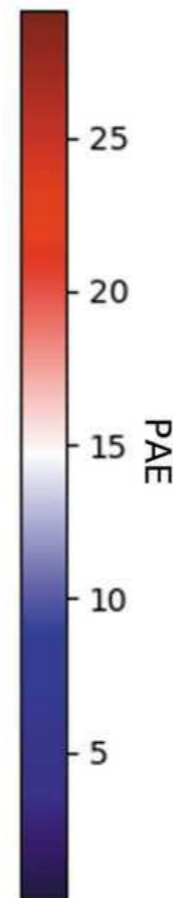
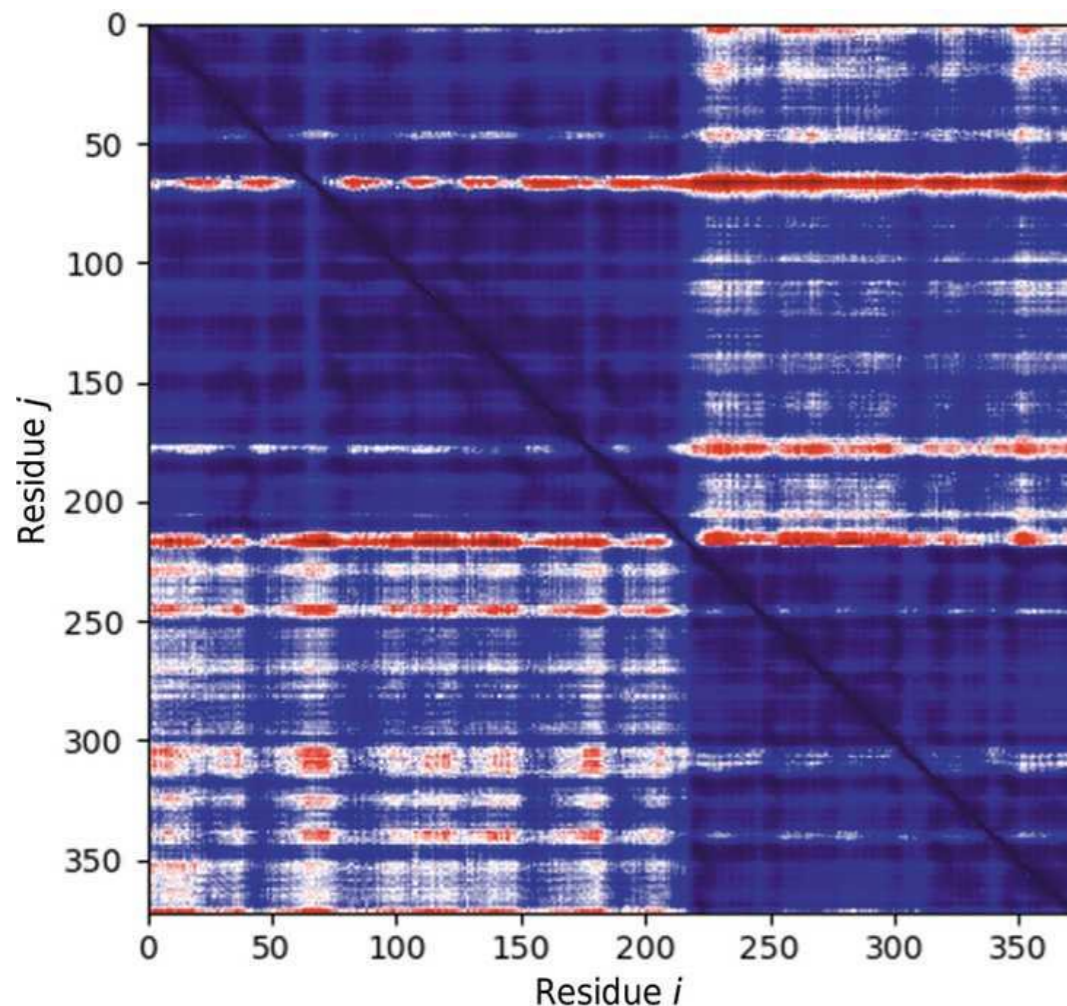
- pLDDt identifies where errors are more likely.
- Per-residue confidence measure.
- Scales from 0 – 100 (pLDDt > 90: predicted with high accuracy).

AlphaFold confidence (pLDDT)	Median prediction error (Å)	Percentage with error over 2 Å
>90	0.6	10
80 - 90	1.1	22
70 - 80	1.5	33
<70	3.5	77

# Predictions come with confidence measures

## 2. Predicted aligned error (PAE)

- Certainty of relative positions between two residues.
- Identifies accurately-predicted domains.
- Dark blue: uncertainty in relative positions  $< 5 \text{ \AA}$ .

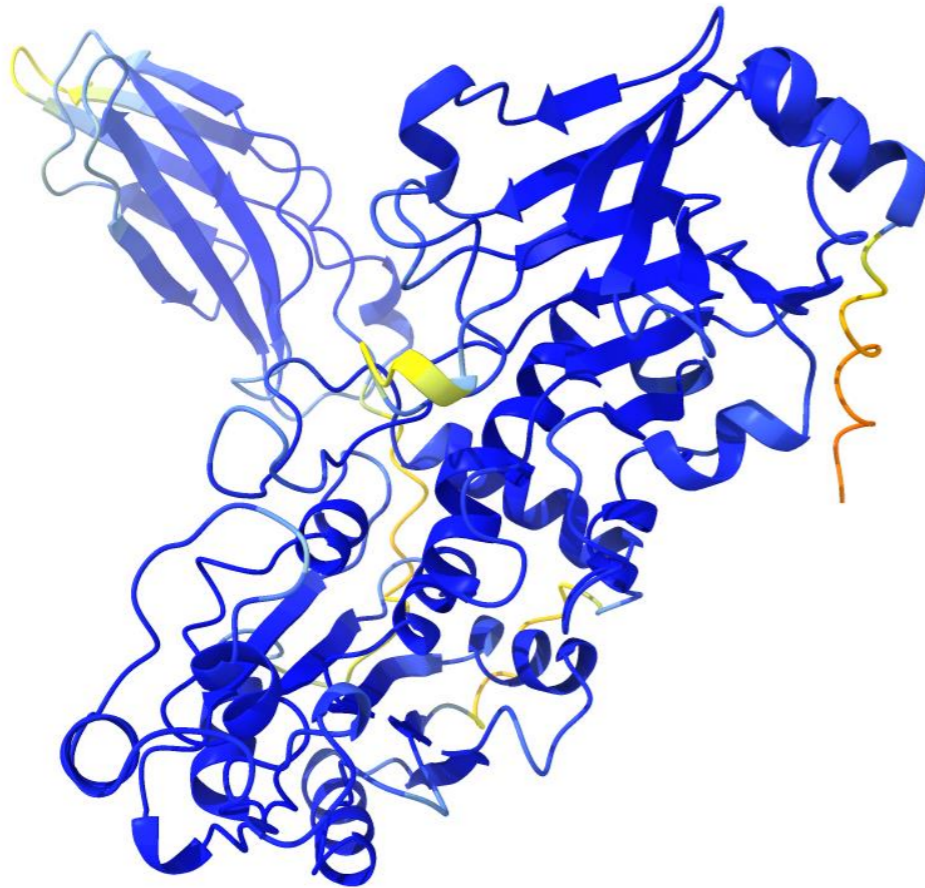


- Suggests 2 domains

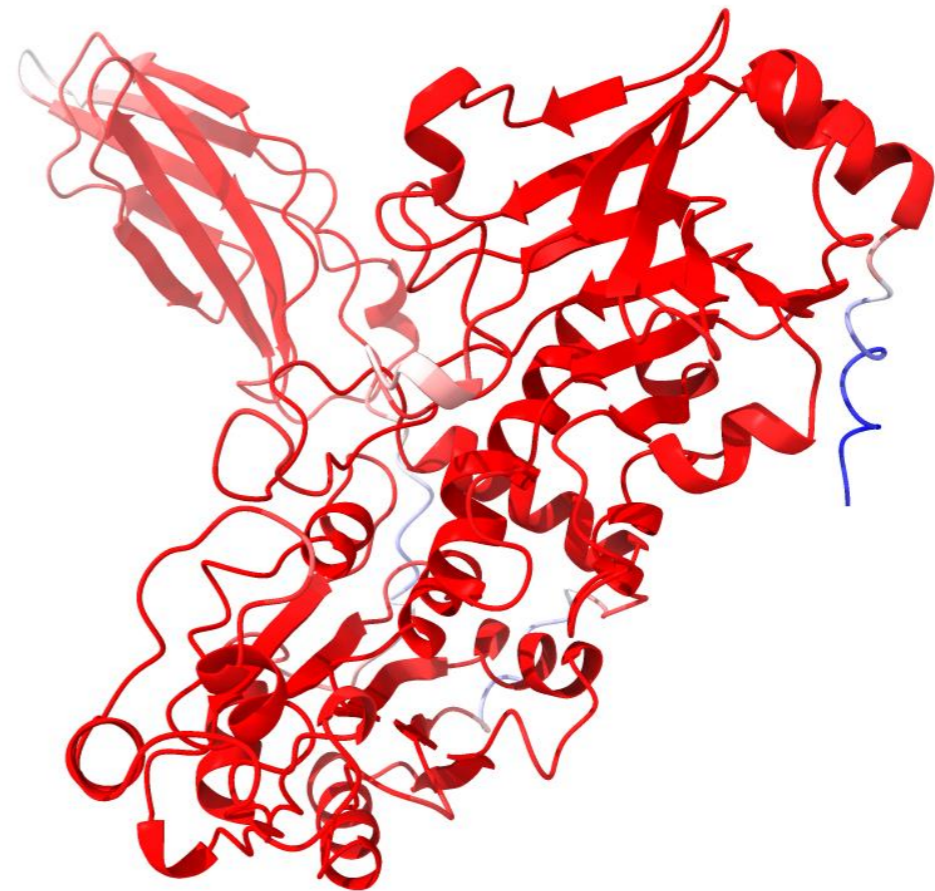
# Using predicted models: B-factors

---

Color by pLDDT



Color by B-factor



high pLDDT (high confidence)

low pLDDT (low confidence, uncertain)



high B-factor (disordered, uncertain)

low B-factor (ordered)

# Using predicted models: B-factors

---

high pLDDT (high confidence)

low pLDDT (low confidence, uncertain)



high B-factor (disordered, uncertain)

low B-factor (ordered)

B-factor may be used in downstream calculations, e.g. to calculate weights for docking. Residues with high B-factors are downweighed.

→ Convert pLDDT to pseudo B-factors.

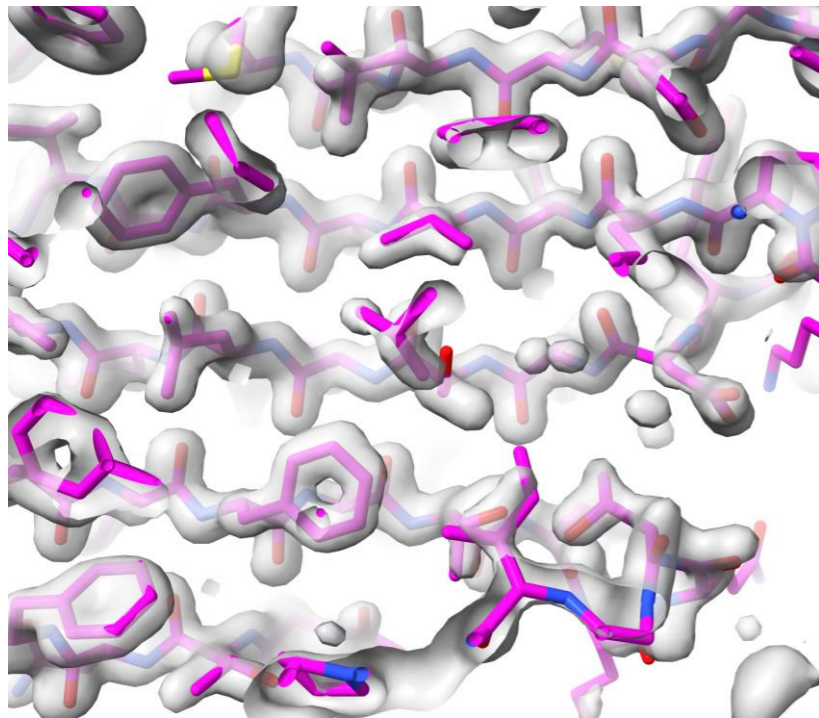
$$\Delta = 1.5 \exp[4(0.7 - \text{pLDDT})]$$

$$B = \frac{8\pi^2 \Delta^2}{3}$$

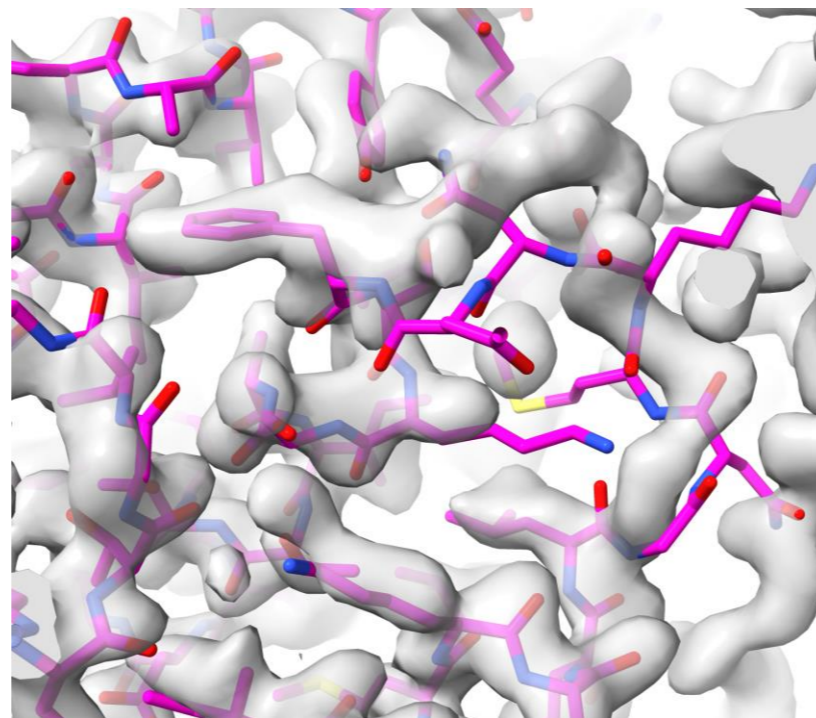
# AlphaFold predictions are great hypotheses

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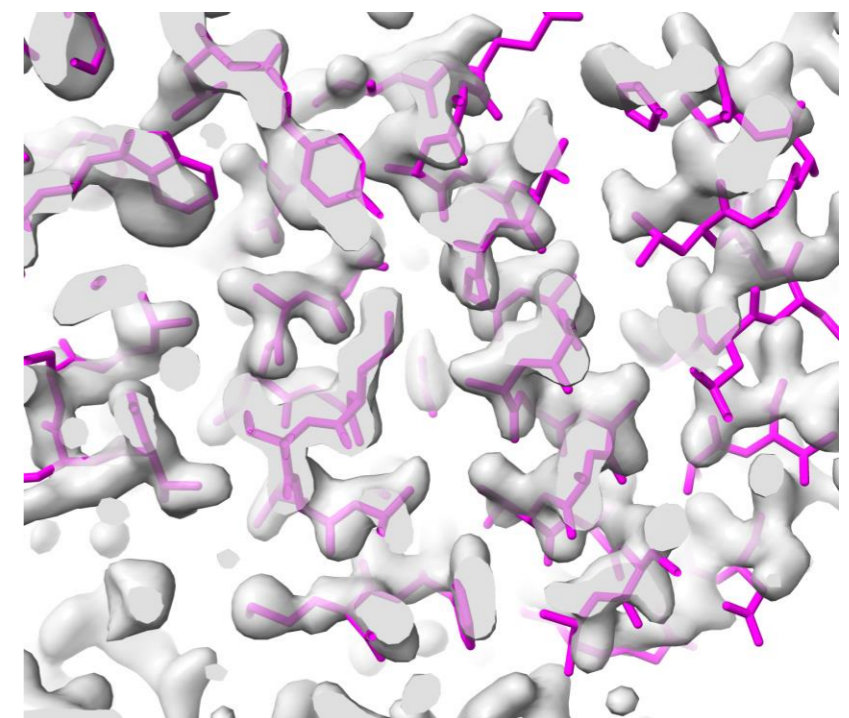
AlphaFold models  
can be....



**Awesome**



**Wrong**

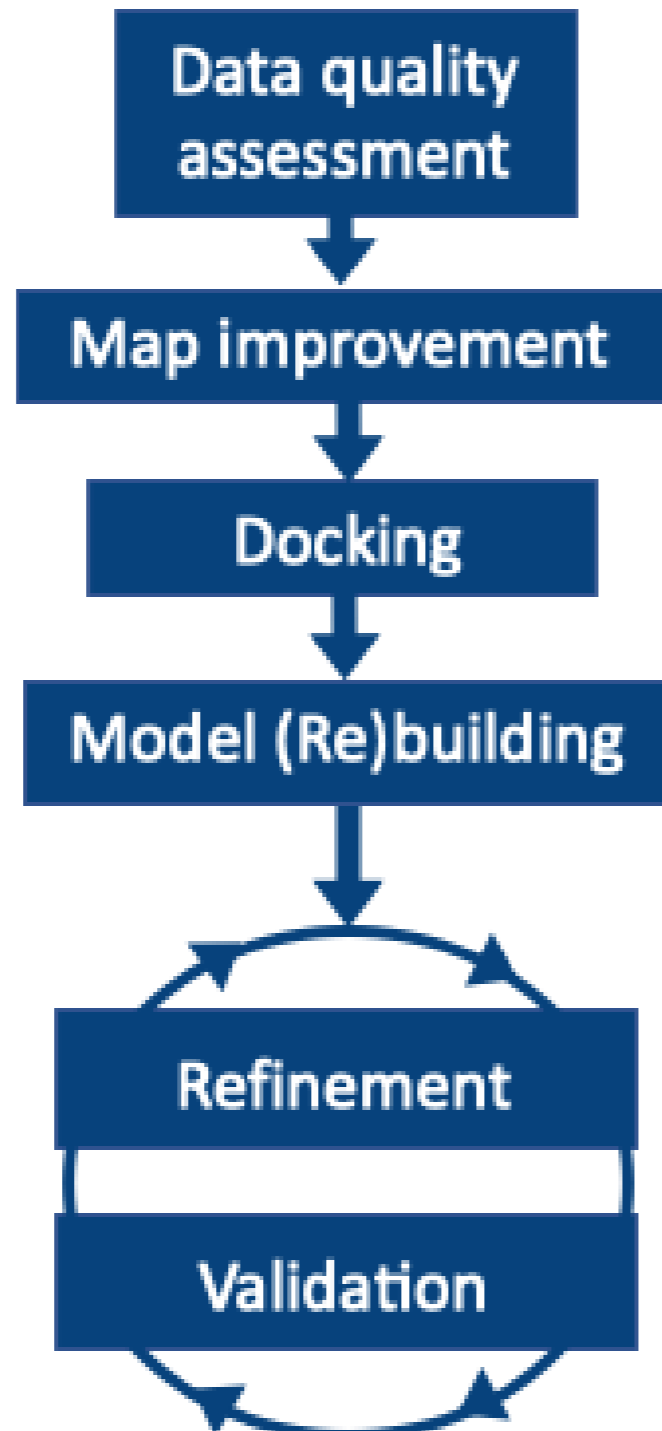


**Distorted**

# What can predicted models be used for?

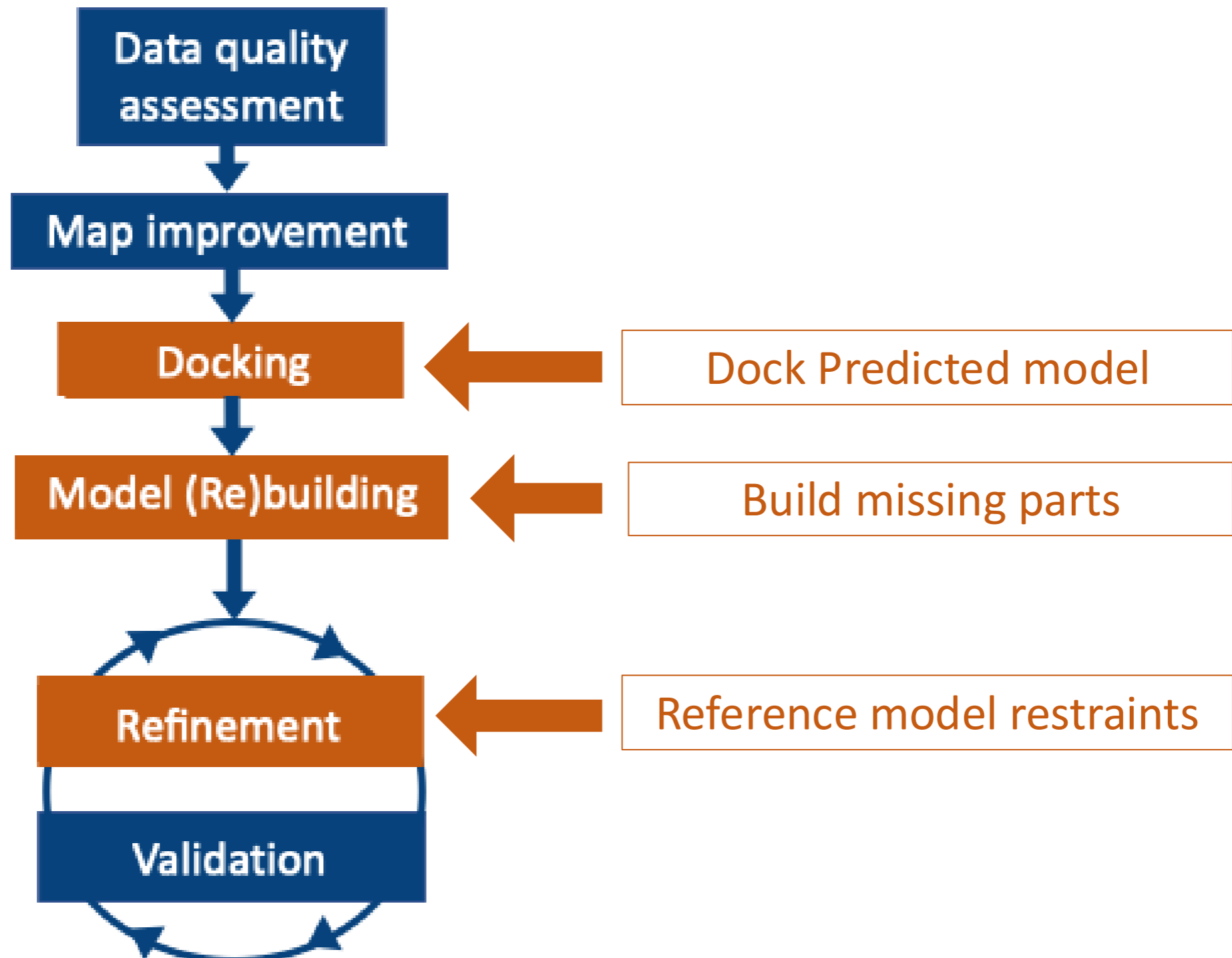
---

Incorporate predictions into the typical cryo-EM workflow.



# What can predicted models be used for?

Incorporate predictions into the typical cryo-EM workflow.



# 1. Use a prediction for cryo-EM docking

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Cryo-EM maps are often insufficiently resolved for reliable ab initio model building.

→ Dock a pre-existing model into the map.

# 1. Use a prediction for cryo-EM docking

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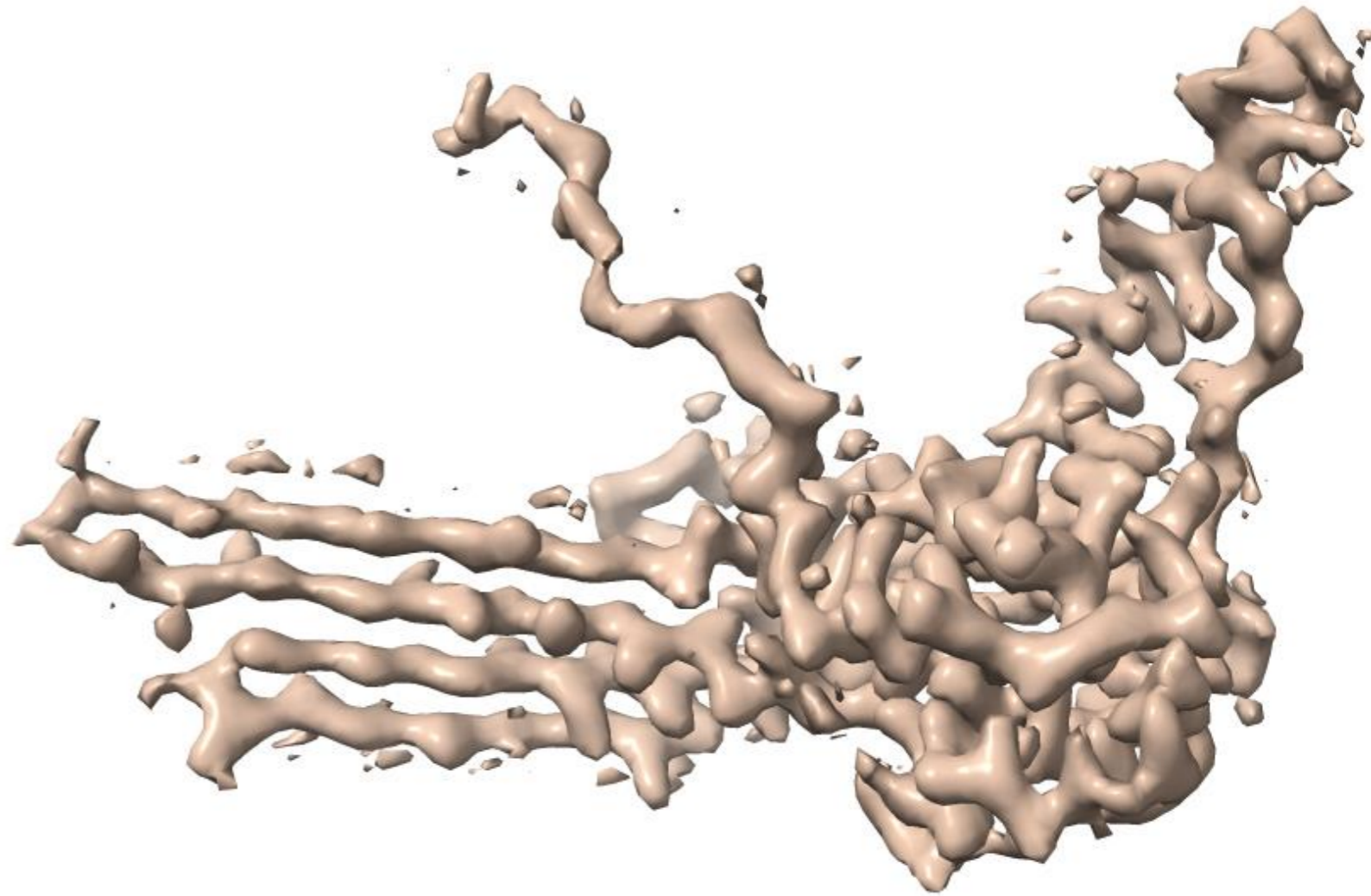
## Example:

Cryo-EM map (30160 – 7brm)

3.6 Å

```
>chain ' A'  
XXXXXXXXXXXXXXXXCLTAPPKEAARPTLMPRAQSYKDLTHLPAP  
TGKIFVSVYNIQDETGQFKPYPASNFSSTAVPQSATAMLVTALKDS  
RWFIPLERQGLQNLNLERKIIRAAQENGTVAINNRIPLQSLTAAN  
IMVEGSIIGYESNVKSGGVGARYFGIGADTQYQLDQIAVNLRVVN  
VSTGEILSSVNTSKTILSYEVQAGVFRFIDYQRLLEGEVGYTSNE  
PVMLCLMSAIETGVIFLINDGIDRGLWDLQNKAERQNDILVKYRH  
MS
```

sequence

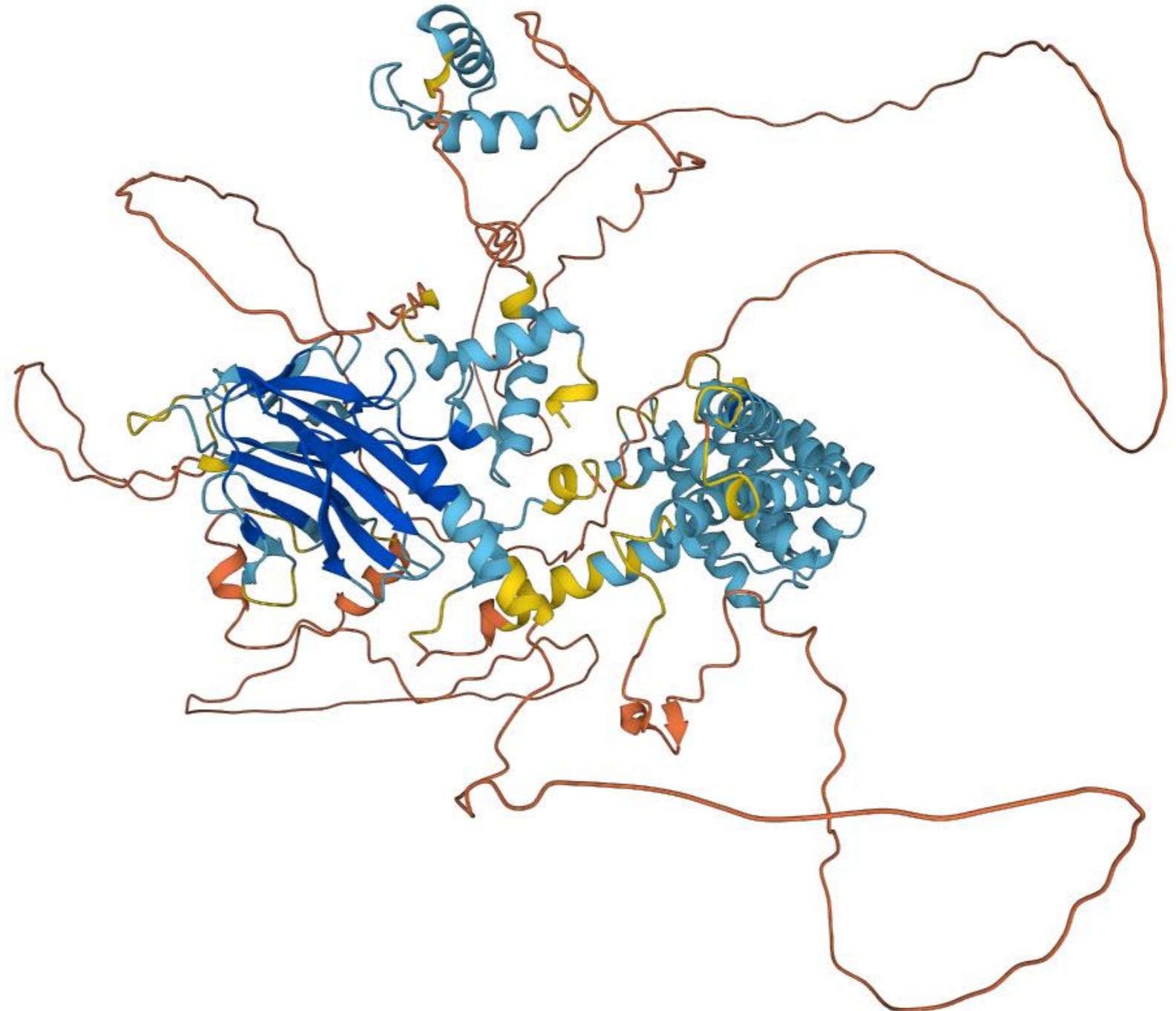


cryo-EM map

# Get a prediction

sequence

```
>chain ' A'  
XXXXXXXXXXXXXXXXXCLTAPPKEAARPTLMPRAQSYKDLTHLPAP  
TGKIFVLSLNNITADNETGAEKDYBACNEETAUDAGATAMLLTALVDE  
RWFIP  
IMVEG  
VSTGE  
PVMLC  
MS
```



AlphaFold  
model



# Process prediction

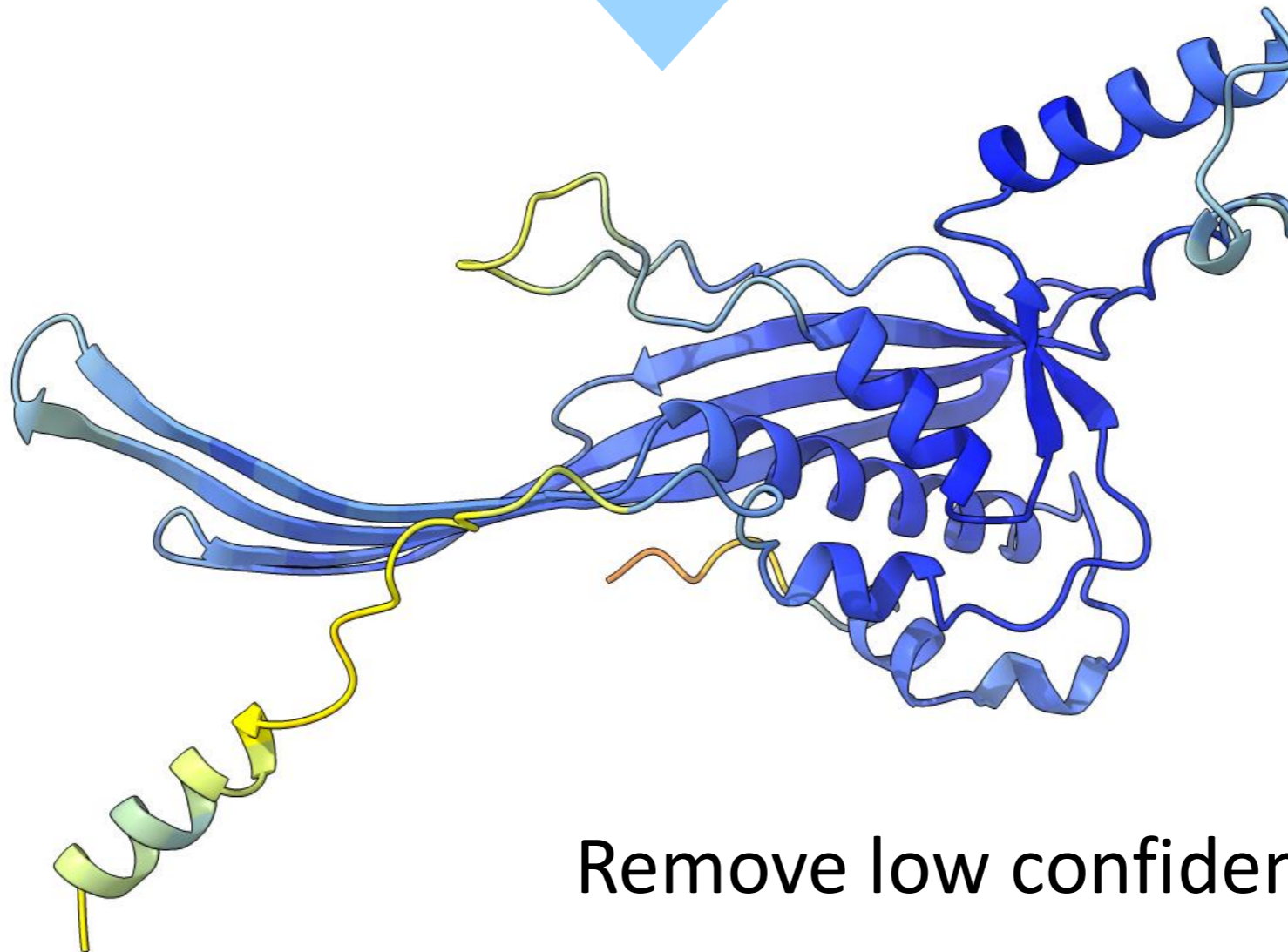
---

sequence

```
>chain ' A'  
XXXXXXXXXXXXXXXXXCLTAPPKEAARPTLMPRAQSYKDLTHLPAP  
TGKIFVSVYNIQDETGQFKYPASNFSSTAVPQSATAMLVTALKDS  
RWFIPLERQGLQNLNLERKIIRAAQENGTVAINNRIPLQSLTAAN  
IMVEGSIIGYESNVKSGGVGARYFGIGADTQYQLDQIAVNLRVVN  
VSTGEILSSVNTSKTILSYEVQAGVFRFIDYQRLLEGEVGYTSNE  
PVMLCLMSAIETGVIFLINDGIDRGLWDLQNKAEQNDILVKYRH  
MS
```



AlphaFold  
model



Remove low confidence parts

# Process prediction

## AlphaFold: Predicted models with Crystals or Cryo-EM



### How to use AlphaFold in Phenix

Documentation for using AlphaFold2 models in Phenix



### AlphaFold model prediction

Predict the chains in a sequence file with AlphaFold



### Process Predicted Model

Replace B-factor field in model and optionally split into domains



### Predict and Build: Crystallography

Iterative AlphaFold prediction, MR/docking, and rebuilding

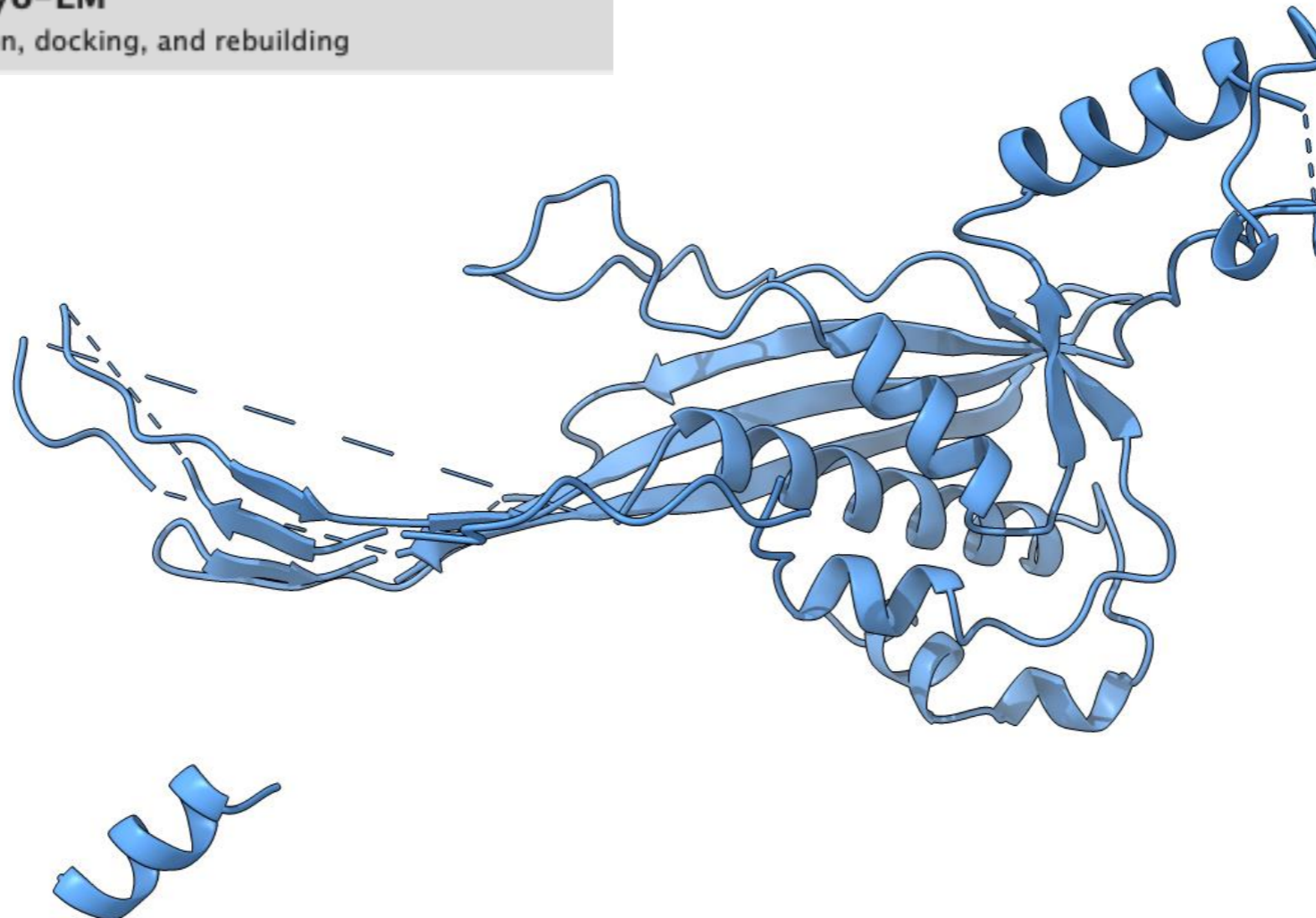


### Predict and Build: Cryo-EM

Iterative AlphaFold prediction, docking, and rebuilding

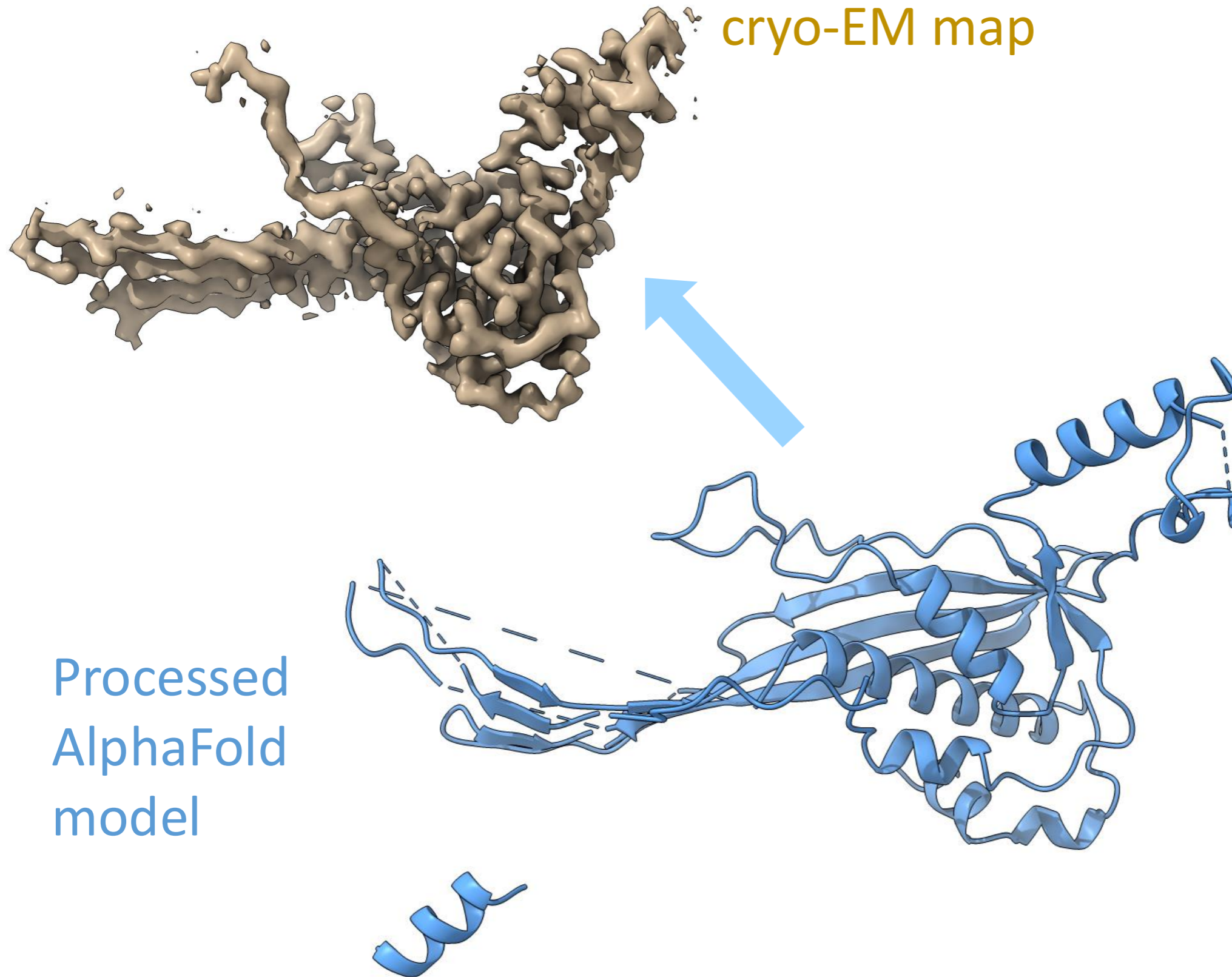
`phenix.process_predicted_model`

Processed  
AlphaFold  
model



# Dock processed model

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# Docking in Phenix

## Dock in map (T. Terwilliger)

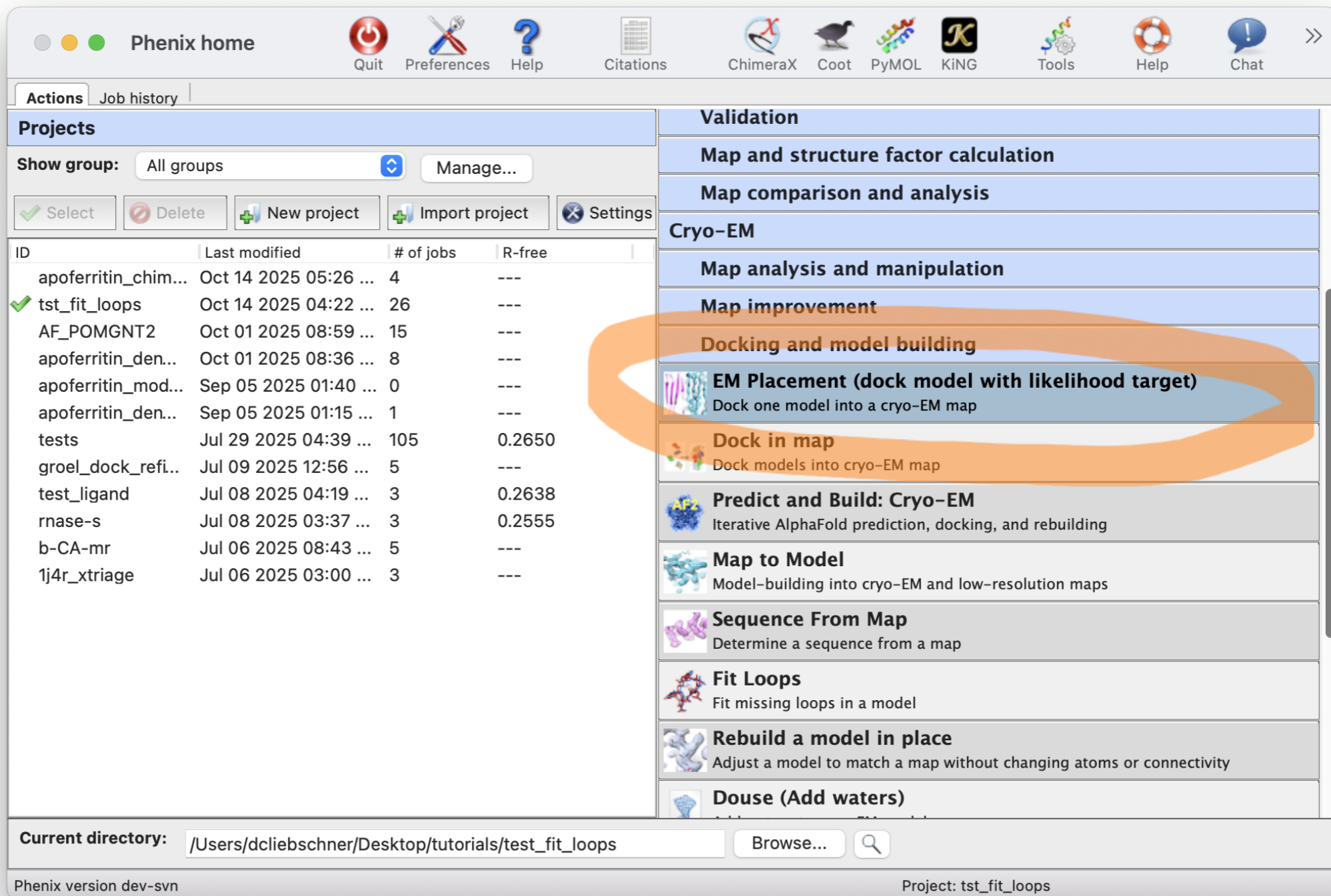
The screenshot displays the Phenix web interface. The top navigation bar includes 'Phenix home' and various utility icons like 'Quit', 'Preferences', 'Help', 'Citations', 'ChimeraX', 'Coot', 'PyMOL', 'KING', 'Tools', 'Help', and 'Chat'. Below this, there are tabs for 'Actions' and 'Job history'. The main content area is divided into two columns. The left column, titled 'Projects', shows a table of project entries with columns for ID, Last modified, # of jobs, and R-free. The right column, titled 'Validation', lists various actions such as 'Map and structure factor calculation', 'Map comparison and analysis', 'Cryo-EM', 'Map analysis and manipulation', 'Map improvement', 'Docking and model building', 'EM Placement (dock model with likelihood target)', 'Dock in map', 'Predict and Build: Cryo-EM', 'Map to Model', 'Sequence From Map', 'Fit Loops', 'Rebuild a model in place', and 'Douse (Add waters)'. The 'Dock in map' option is highlighted with an orange oval. At the bottom, the 'Current directory' is shown as '/Users/dcliebschner/Desktop/tutorials/test\_fit\_loops' and the 'Project' is 'tst\_fit\_loops'.

ID	Last modified	# of jobs	R-free
apoferritin_chim...	Oct 14 2025 05:26 ...	4	---
✓ tst_fit_loops	Oct 14 2025 04:22 ...	26	---
AF_POMGNT2	Oct 01 2025 08:59 ...	15	---
apoferritin_den...	Oct 01 2025 08:36 ...	8	---
apoferritin_mod...	Sep 05 2025 01:40 ...	0	---
apoferritin_den...	Sep 05 2025 01:15 ...	1	---
tests	Jul 29 2025 04:39 ...	105	0.2650
groel_dock_refi...	Jul 09 2025 12:56 ...	5	---
test_ligand	Jul 08 2025 04:19 ...	3	0.2638
rnase-s	Jul 08 2025 03:37 ...	3	0.2555
b-CA-mr	Jul 06 2025 08:43 ...	5	---
1j4r_xtriage	Jul 06 2025 03:00 ...	3	---

# Docking in Phenix

## Likelihood-based EM docking (R. Read)

- Use likelihood scores to dock a model into a map
- Works at low resolution (8.5 Å)

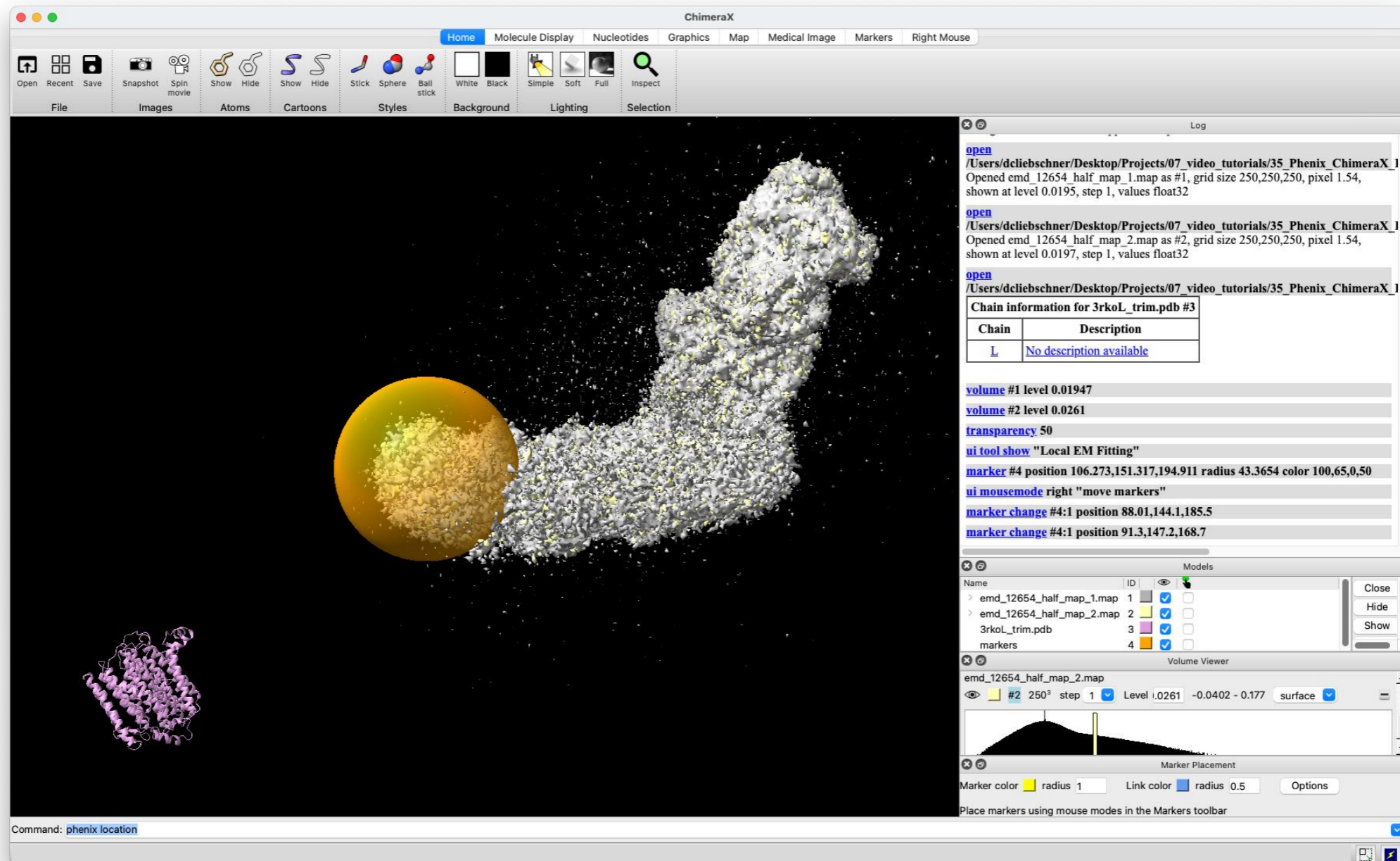


The screenshot displays the Phenix software interface. The top menu bar includes options like Quit, Preferences, Help, Citations, ChimeraX, Coot, PyMOL, KING, Tools, Help, and Chat. The main window is divided into several sections:

- Projects:** A table listing various projects with columns for ID, Last modified, # of jobs, and R-free. The project 'tst\_fit\_loops' is selected.
- Validation:** A list of validation tasks including 'Map and structure factor calculation', 'Map comparison and analysis', 'Cryo-EM', 'Map analysis and manipulation', 'Map improvement', 'Docking and model building', 'EM Placement (dock model with likelihood target)', 'Dock in map', 'Predict and Build: Cryo-EM', 'Map to Model', 'Sequence From Map', 'Fit Loops', 'Rebuild a model in place', and 'Douse (Add waters)'. The 'Docking and model building' section is highlighted with an orange oval, and the 'EM Placement (dock model with likelihood target)' option is specifically highlighted with a blue arrow.
- Current directory:** /Users/dcliebschner/Desktop/tutorials/test\_fit\_loops
- Phenix version:** dev-svn
- Project:** tst\_fit\_loops

# Docking in Phenix/ChimeraX

- Likelihood-based docking can be done via ChimeraX.
- Can select the region into which the model should be docked.



## 2. Use a prediction as reference model

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### Concept

- Use a related model to generate a set of **torsion restraints**.
- Restrain **each torsion angle** in the working model to the corresponding torsion angle in the reference model.
- Allows for structural differences.

### When to use

Low resolution (worse than 3Å).

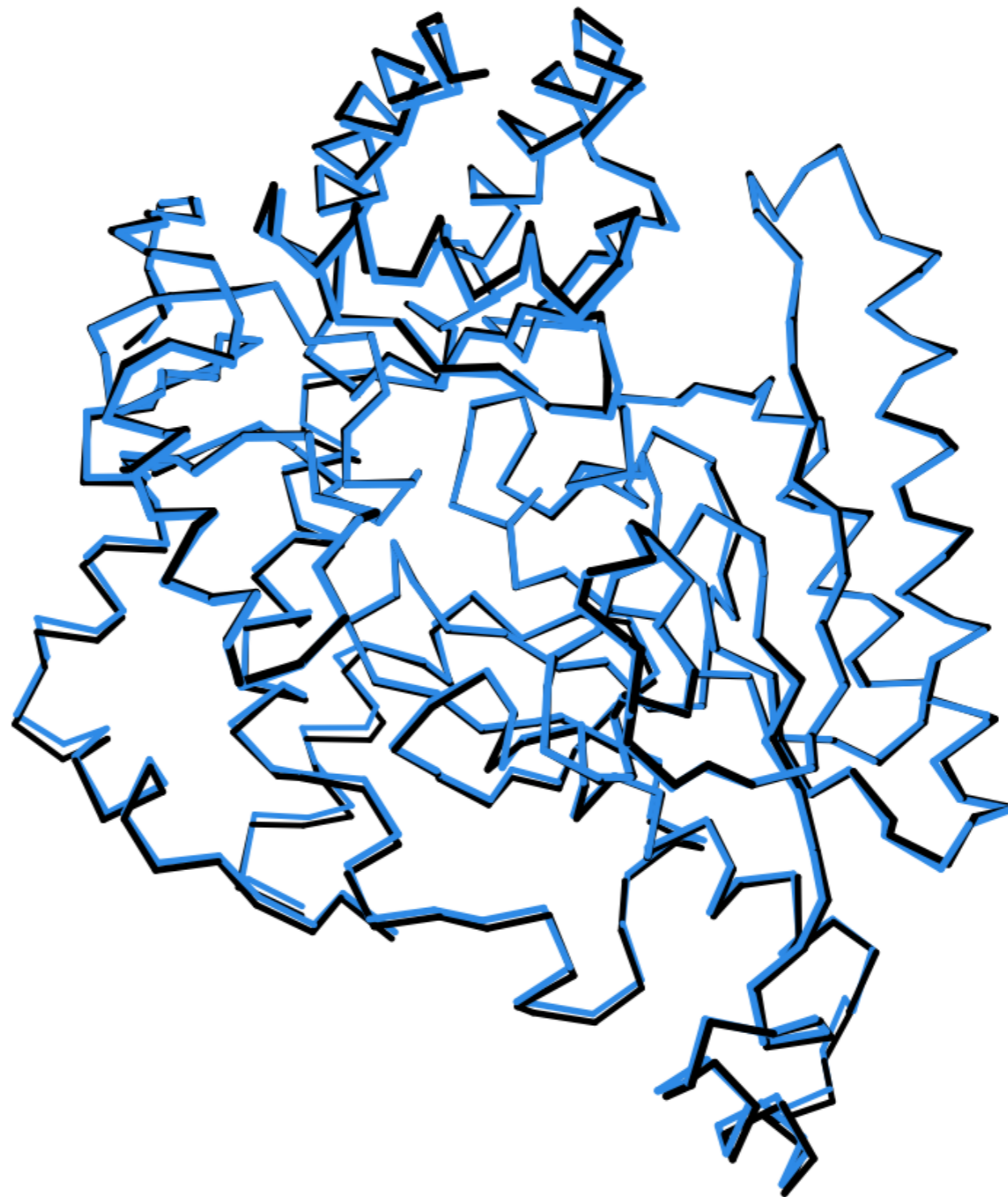
If no high resolution homologue available, can use AF model for reference model (AF models have good geometry).

# Reference model Restraints: example

---

**1GTX:** 3.0 Å

**1OHV:** 2.3 Å



4-aminobutyrate-aminotransferase

# Reference model Restraints: example

---



**1OHV:** 2.3 Å



**1GTX:** 3.0 Å

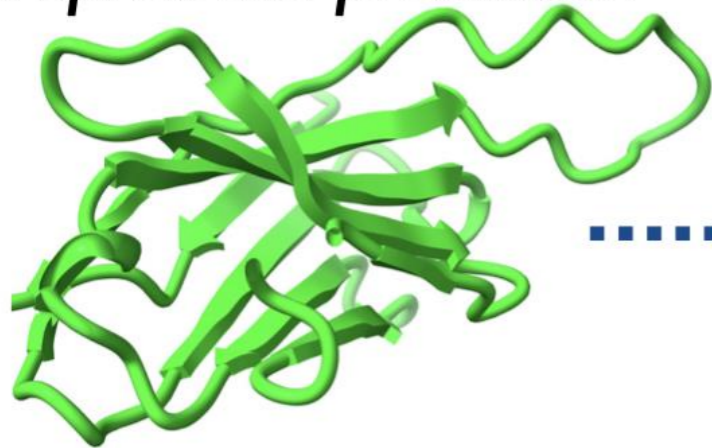
4-aminobutyrate-aminotransferase

# Use your working model to get a new AlphaFold prediction

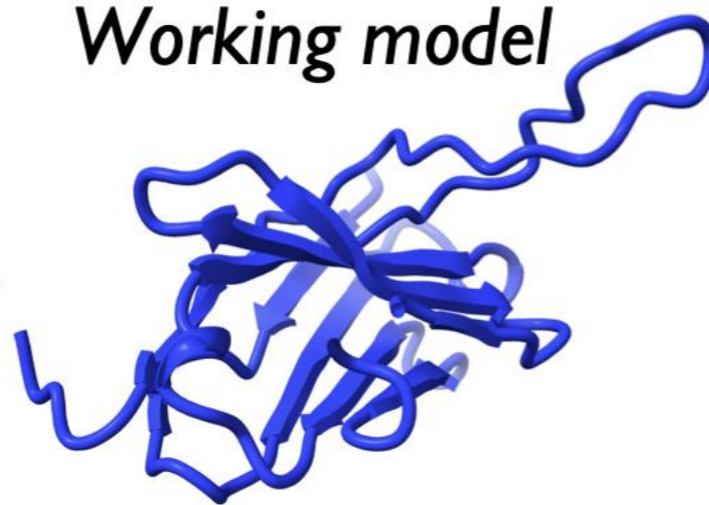
*Why?*

Because your new prediction might be better than your model

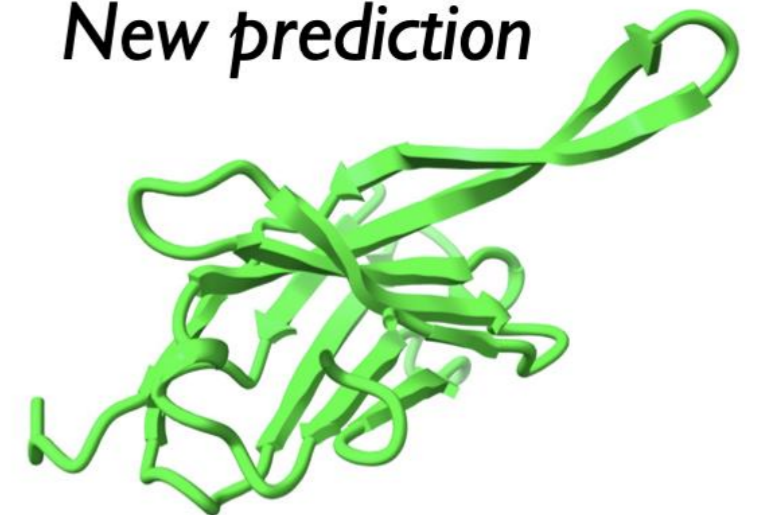
*AlphaFold prediction*



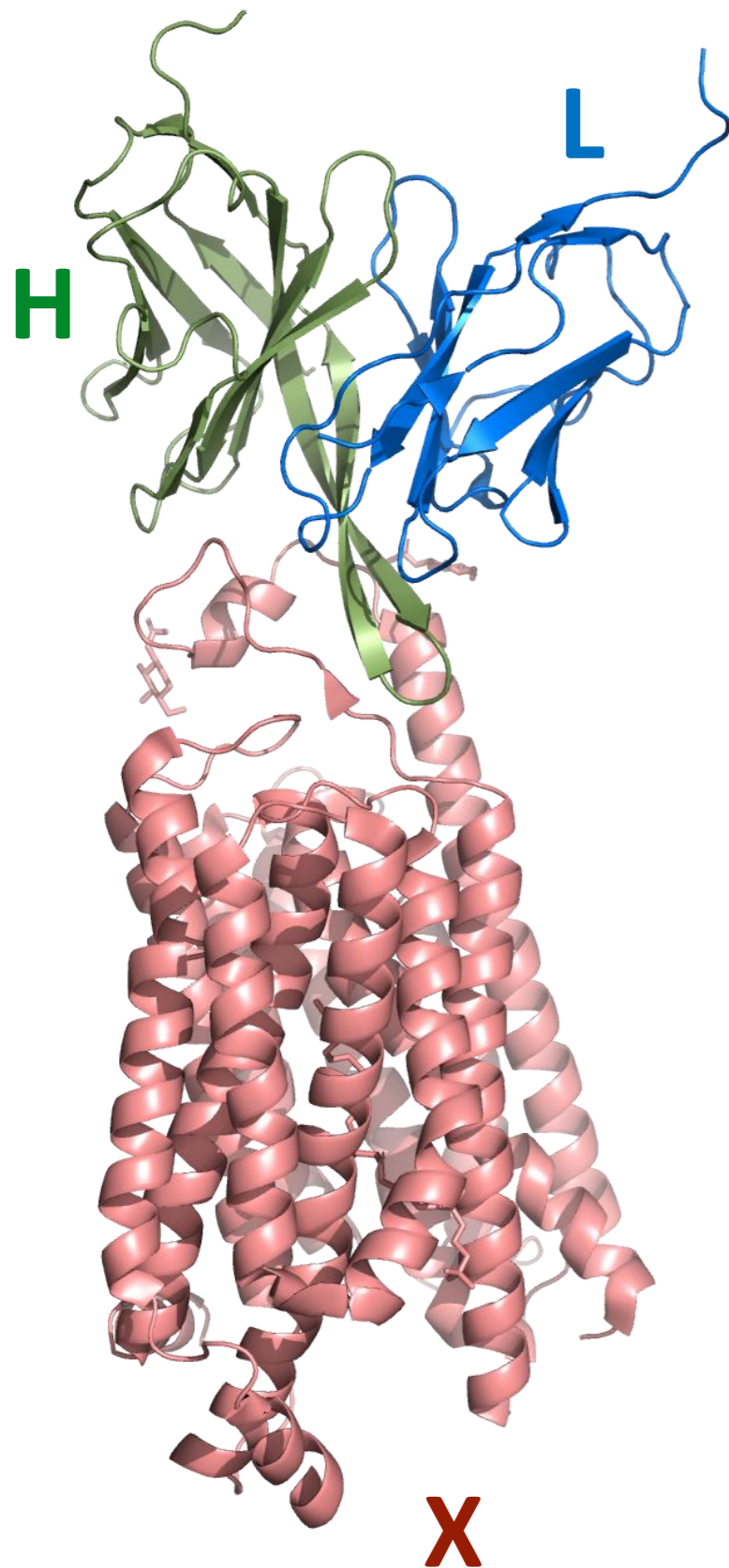
*Working model*



*New prediction*



# Example: Fab heavy chain



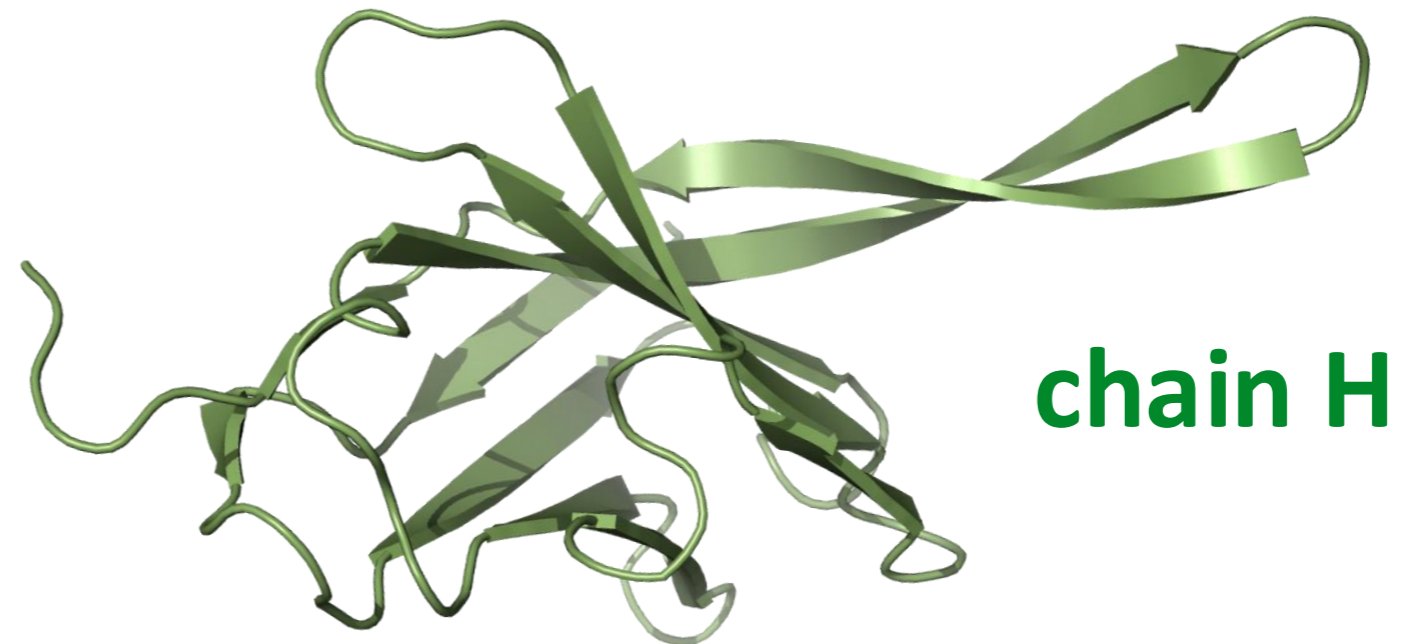
## 7MJS

Single-Particle Cryo-EM Structure of Major Facilitator Superfamily Domain containing 2A in complex with LPC-18:3

PDB DOI: <https://doi.org/10.2210/pdb7MJS/pdb>

EM Map EMD-23883: [EMDB EMDataResource](https://emdb.emdataresource.org/EMD-23883)

3.03 Å resolution



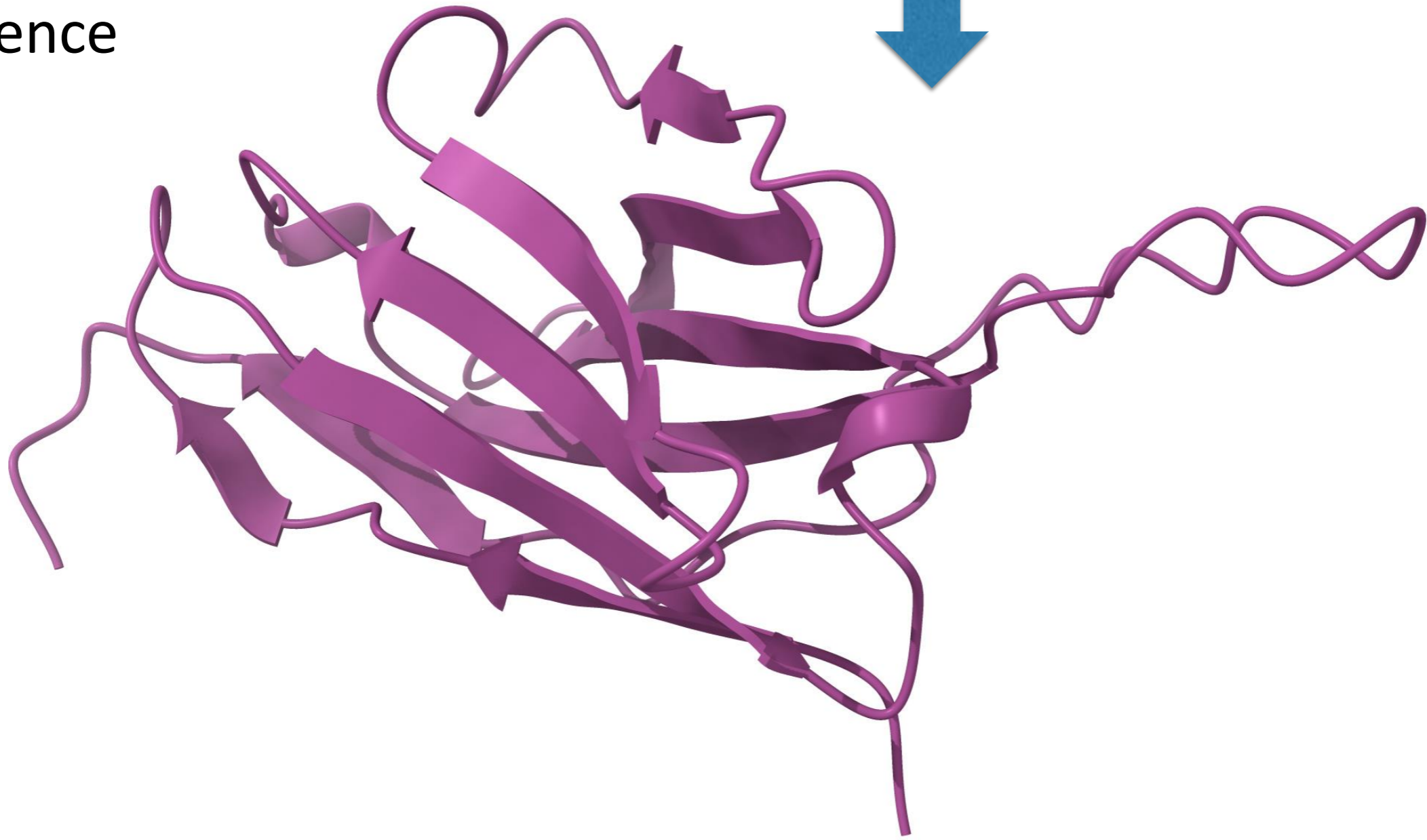
A loop that interacts with other chains is not correctly predicted.

# AF2 prediction of chain H

---

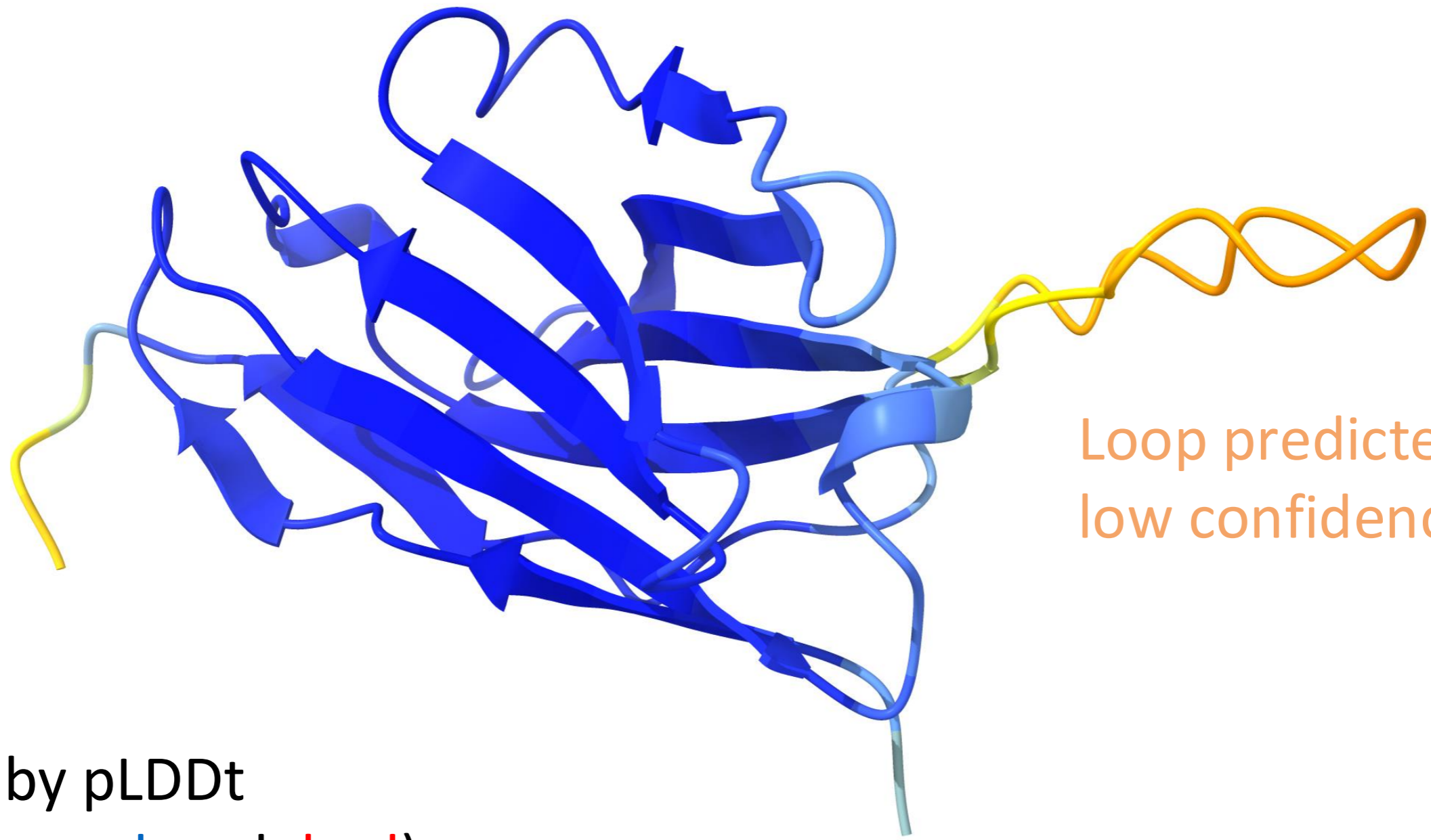
EVQLVESGGGLVQPGGSLRLSCAASGFNIYSSSIHWVRQ  
APGKGLEWVAYIYSYSGYTSYADSVKGRFTISADTSKNTA  
YLQMNSLRAEDTAVYYCARSLEYLYSSGYQYKWATGLDY  
WGQGTLVTVSSAST

Sequence



# AF2 prediction of chain H

---

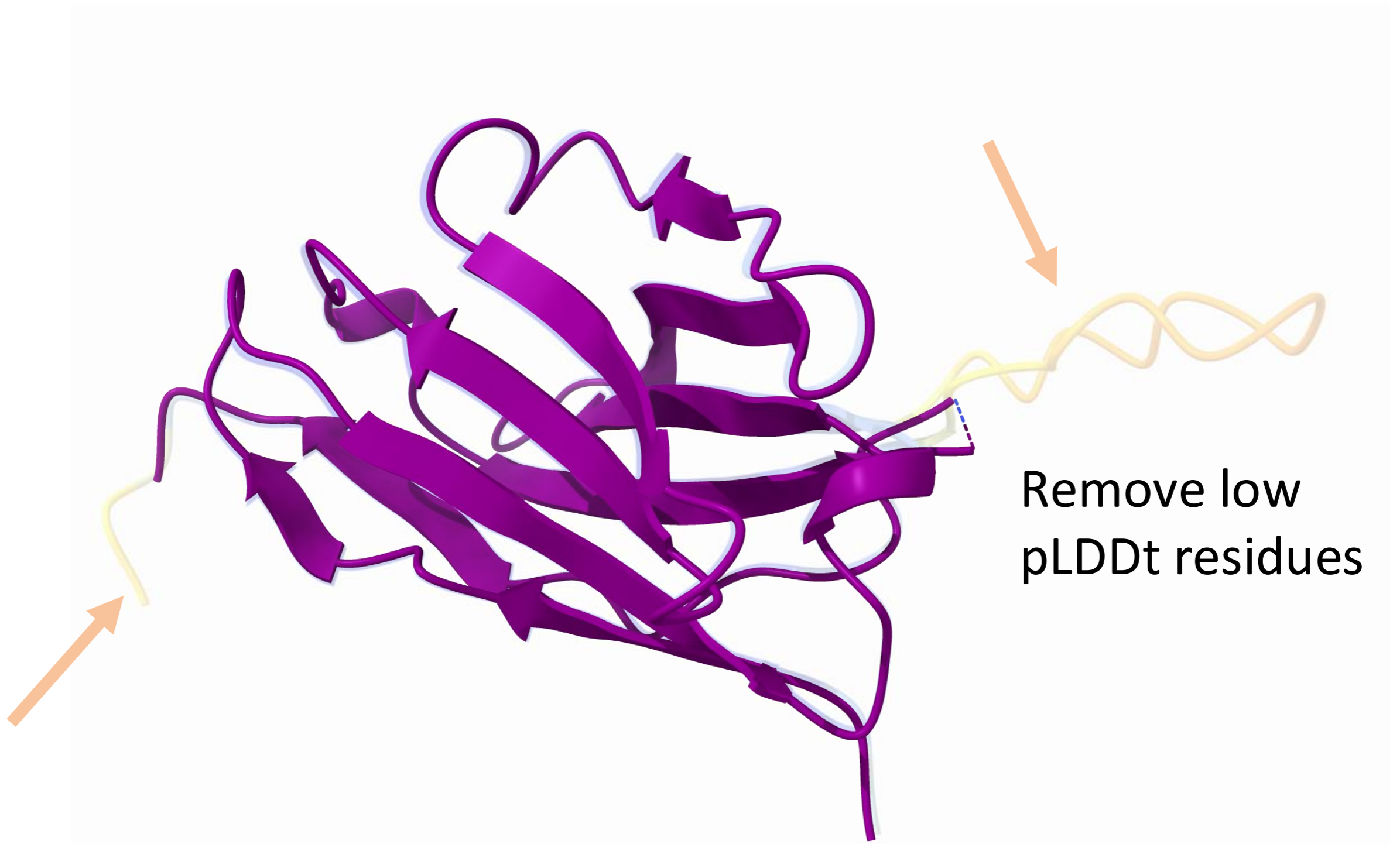


Loop predicted with  
low confidence

Color by pLDDT  
(blue: **good**, red: **bad**)

# Process AF2 prediction

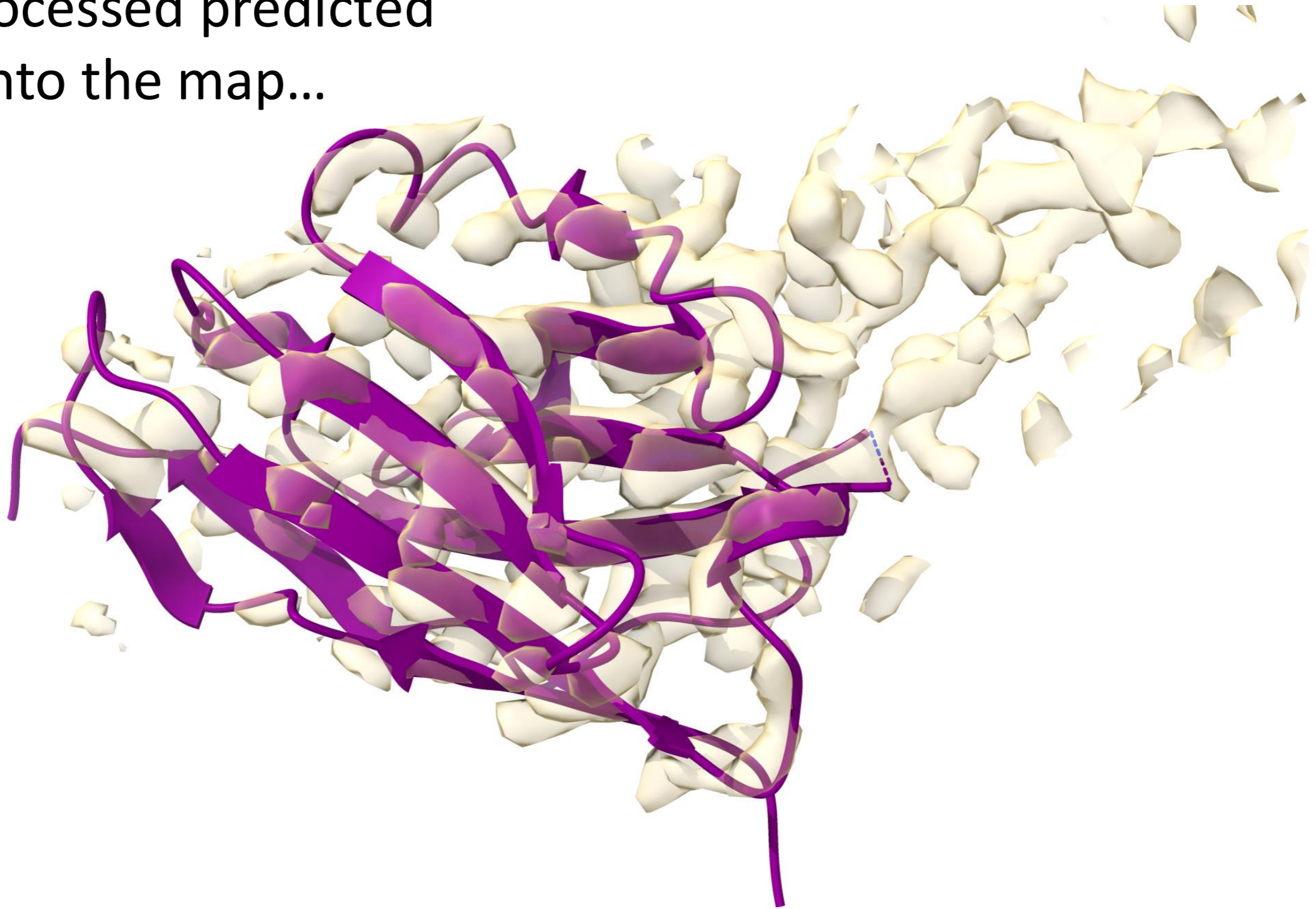
---



# Dock processed model into the map

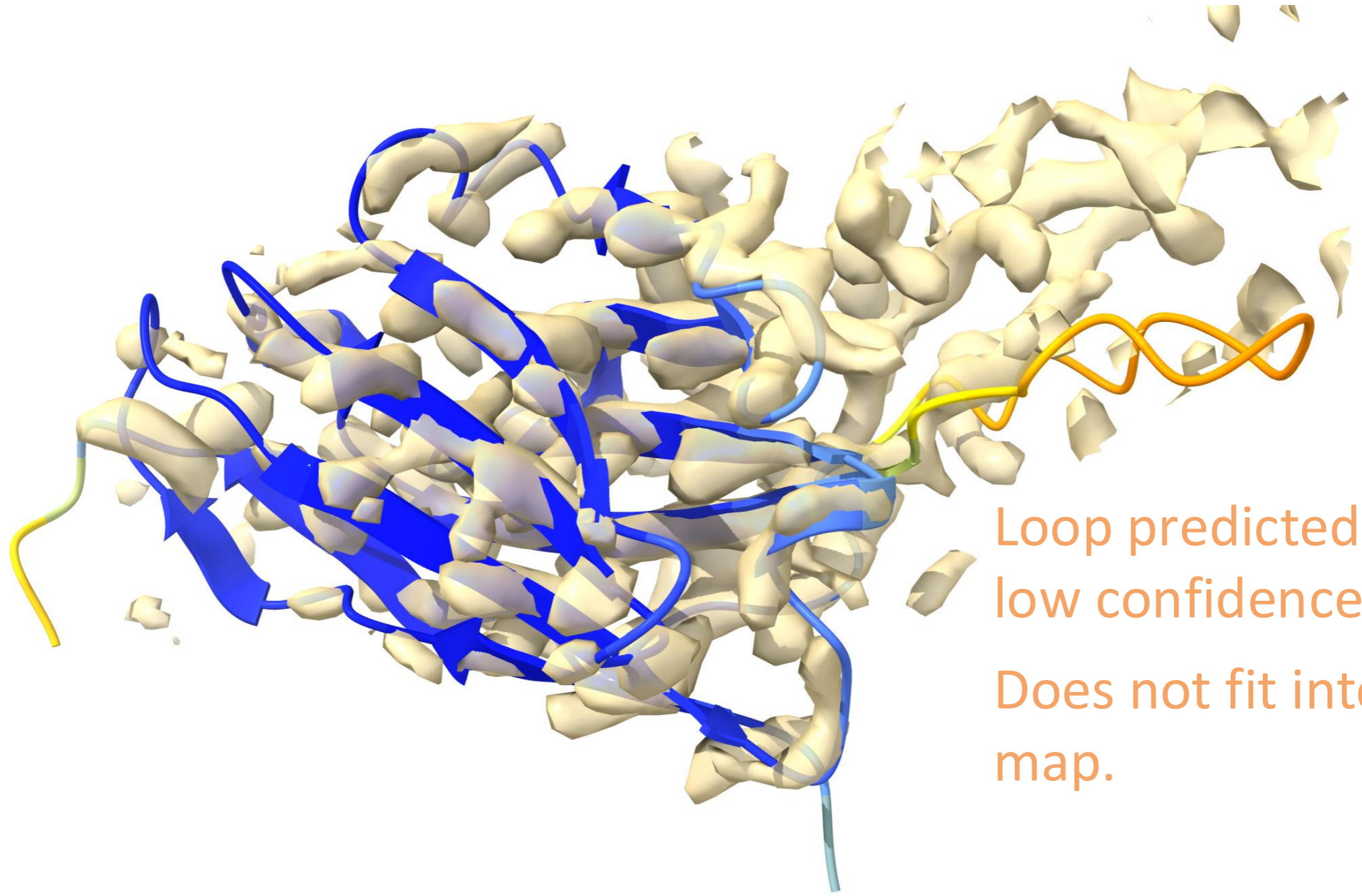
---

Dock processed predicted model into the map...



# AF2 prediction of chain H

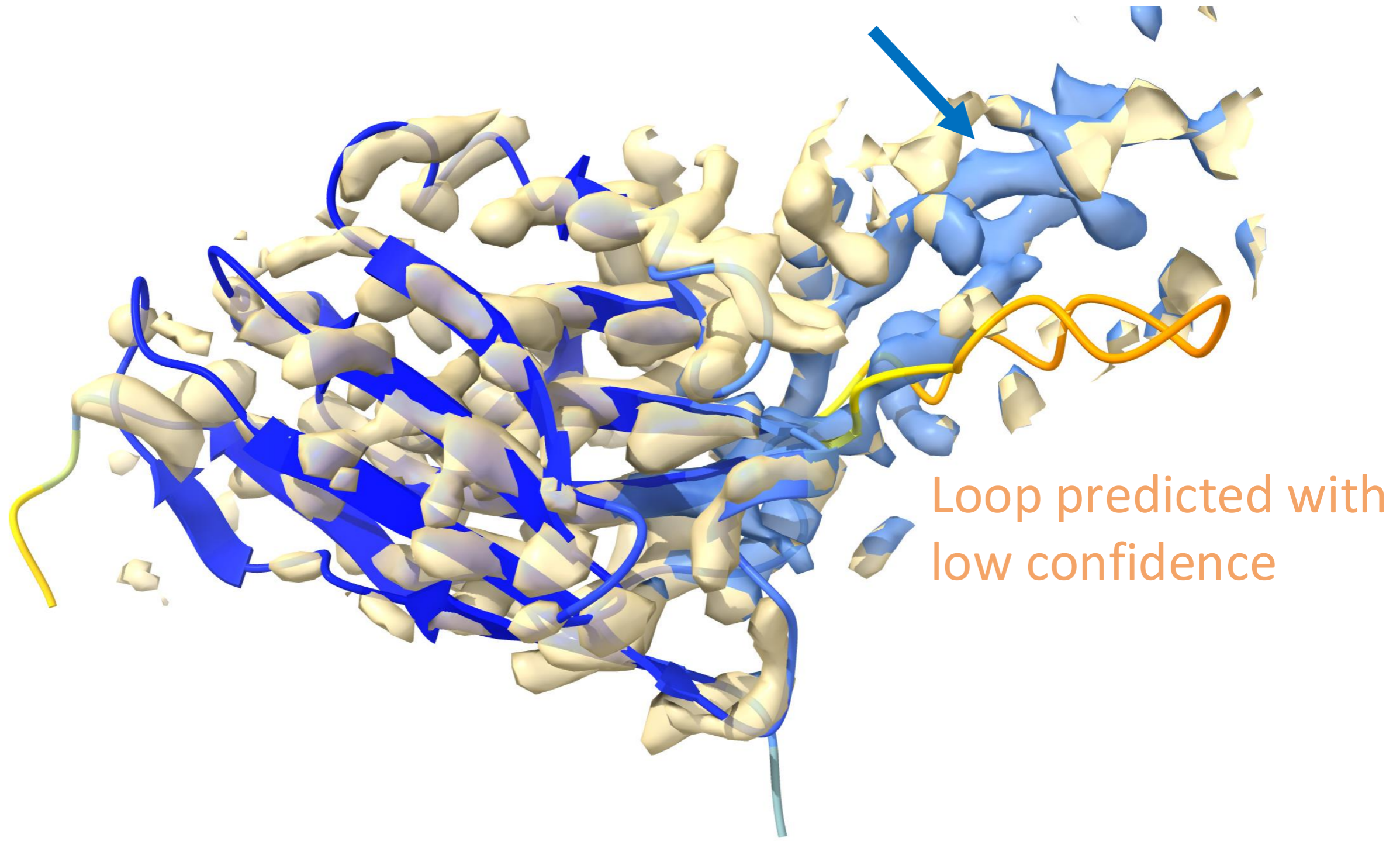
---



Loop predicted with  
low confidence  
Does not fit into  
map.

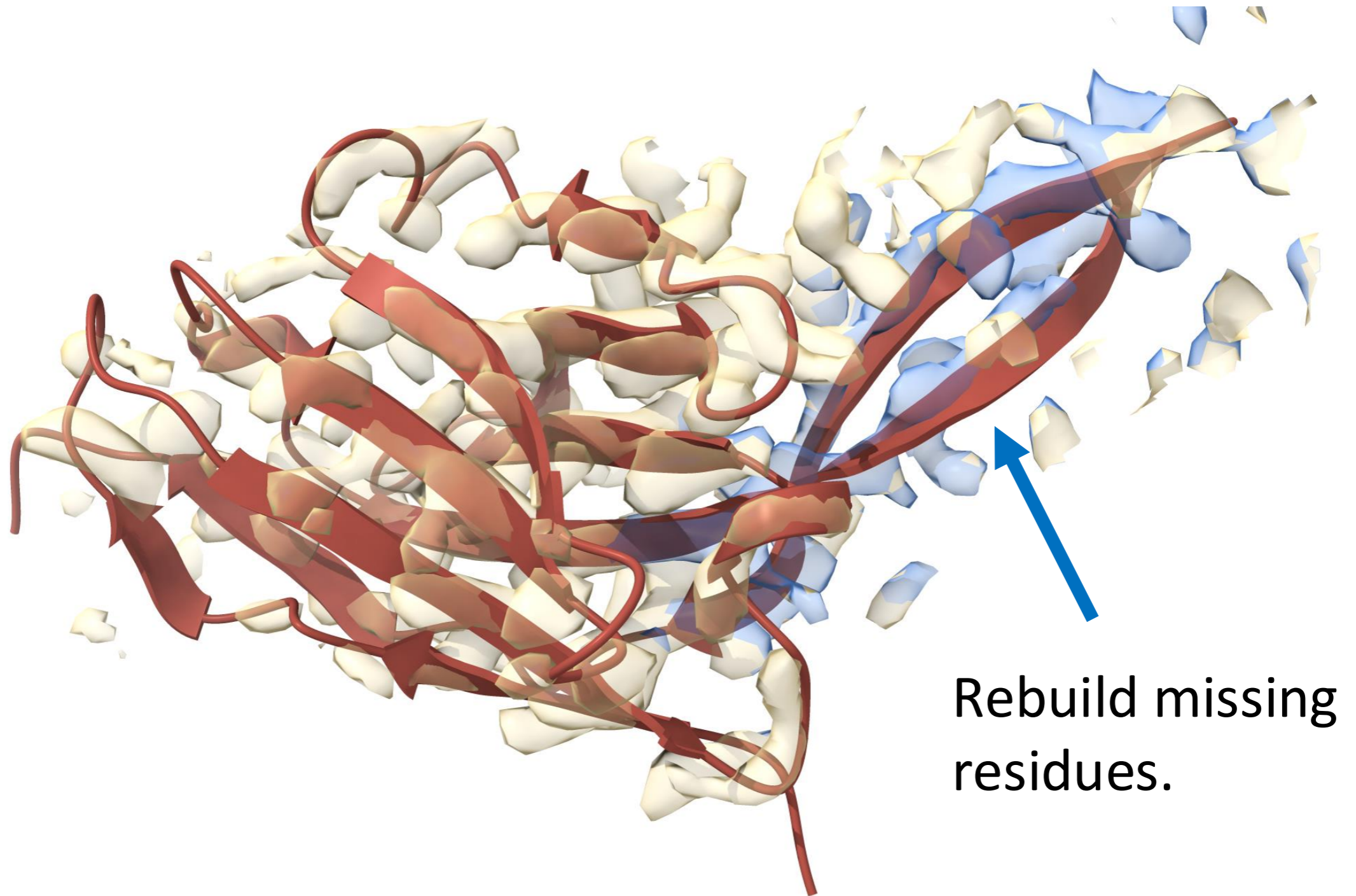
# AF2 prediction of chain H

---



# Dock and rebuild model

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“predicted-processed-docked-rebuilt” model

# Make a new prediction

## Sequence

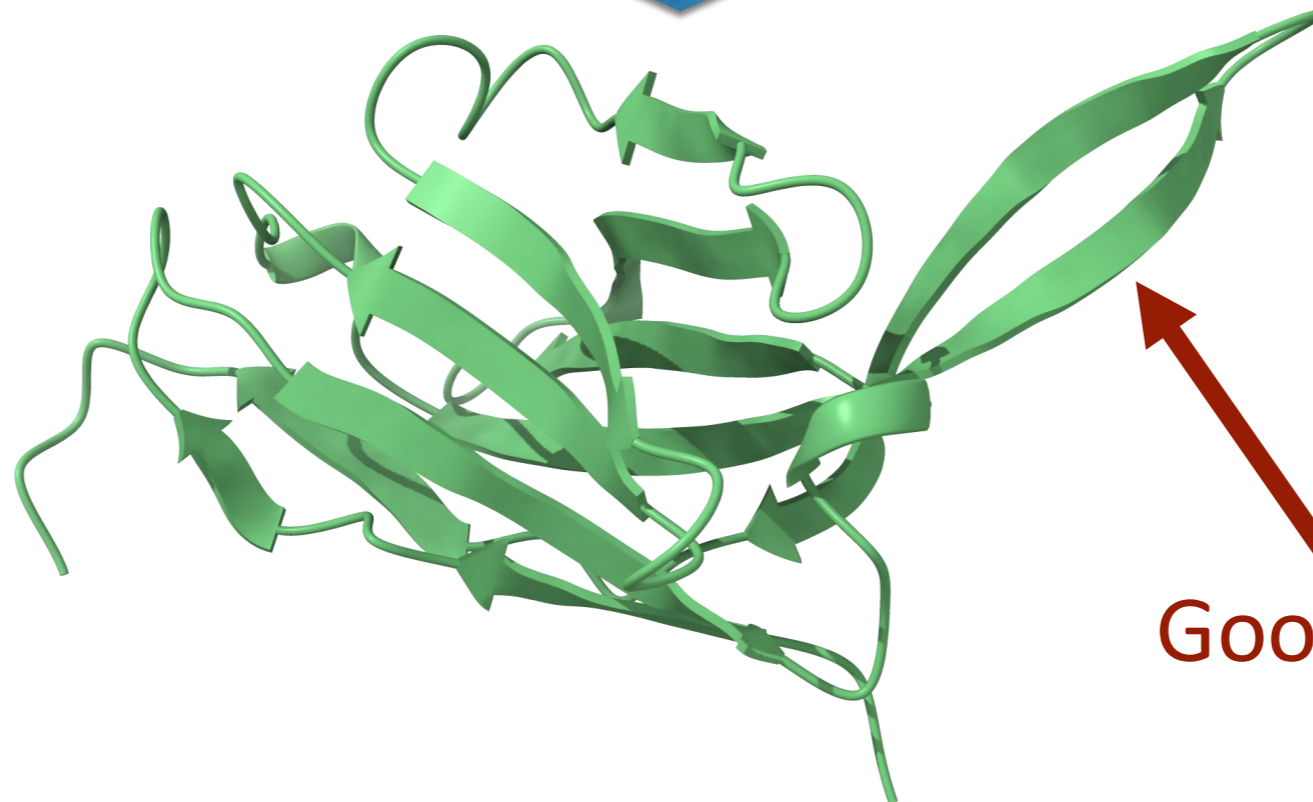
EVQLVESGGGLVQPGGSLRLSCAASGFNIYSSSIHWVRQ  
APGKGLEWVAYIYSYSGYTSYADSVKGRFTISADTSKNTA  
YLQMNSLRAEDTAVYYCARSLEYLYSSGYQYKWATGLDY  
WGQGTLVTVSSAST

## Template



Good loop

AlphaFold



Good loop

# Using a template improves prediction

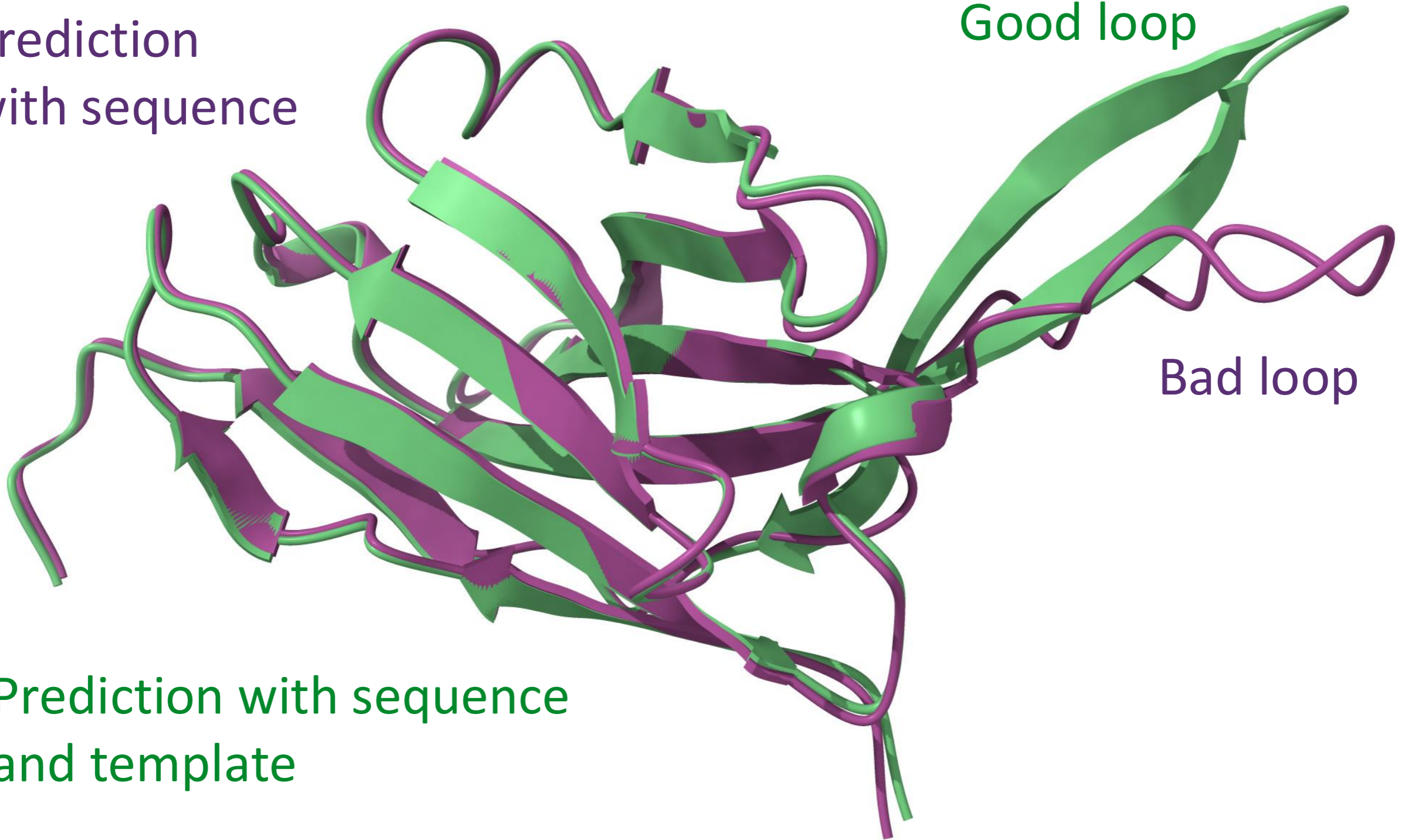
---

Prediction  
with sequence

Good loop

Bad loop

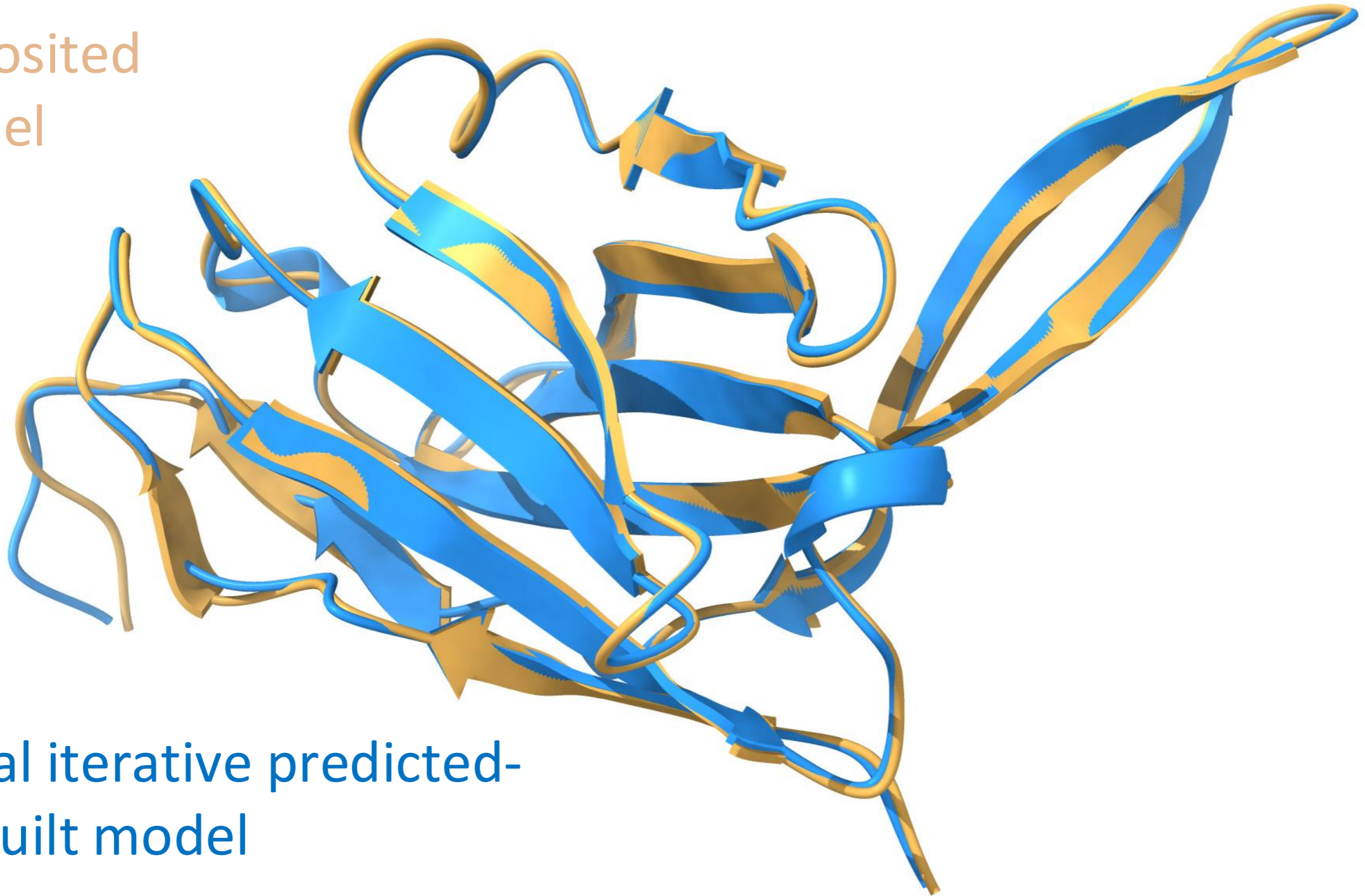
Prediction with sequence  
and template



# Iterate prediction and rebuilding

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Deposited  
model

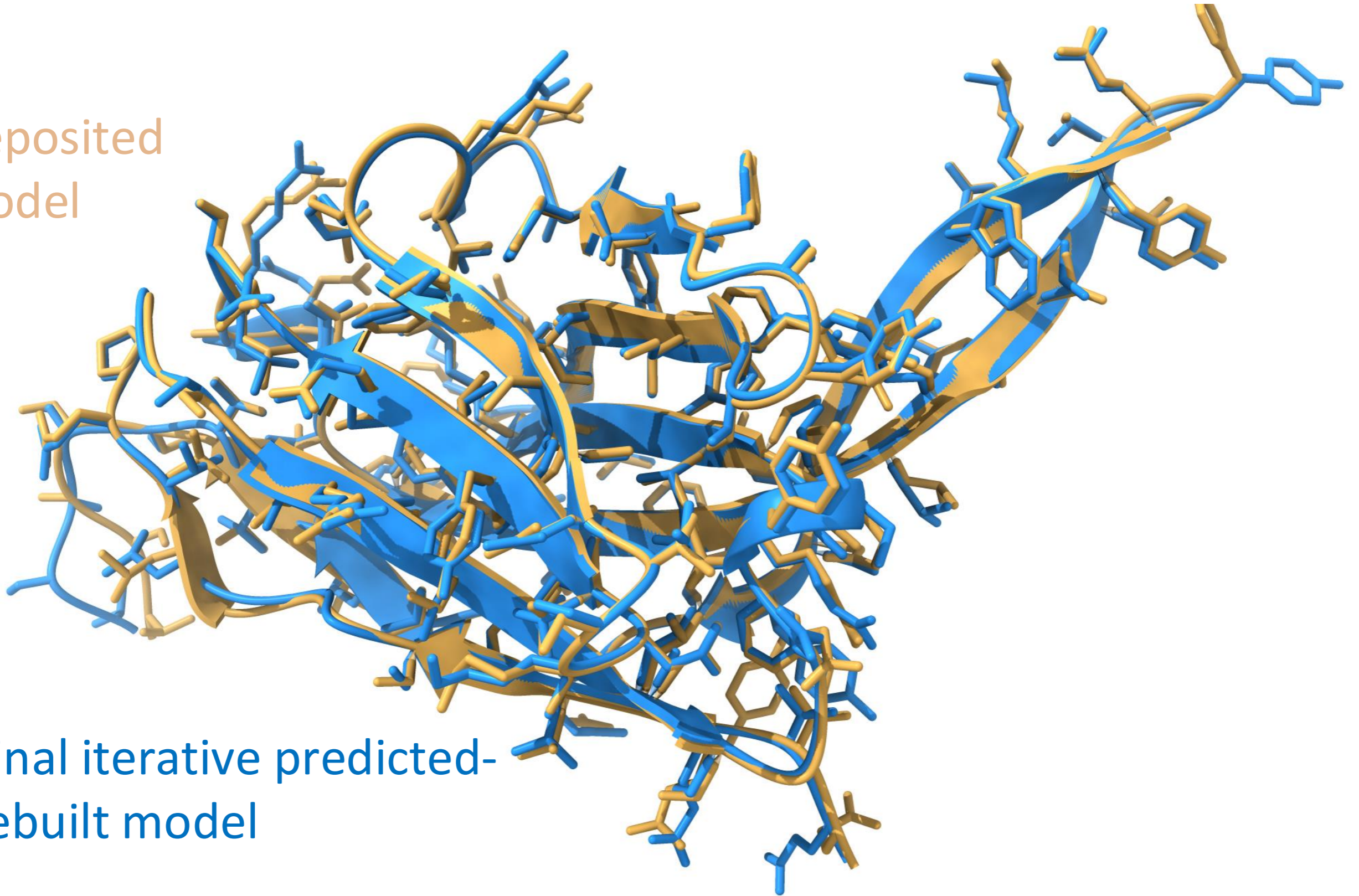


Final iterative predicted-  
rebuilt model

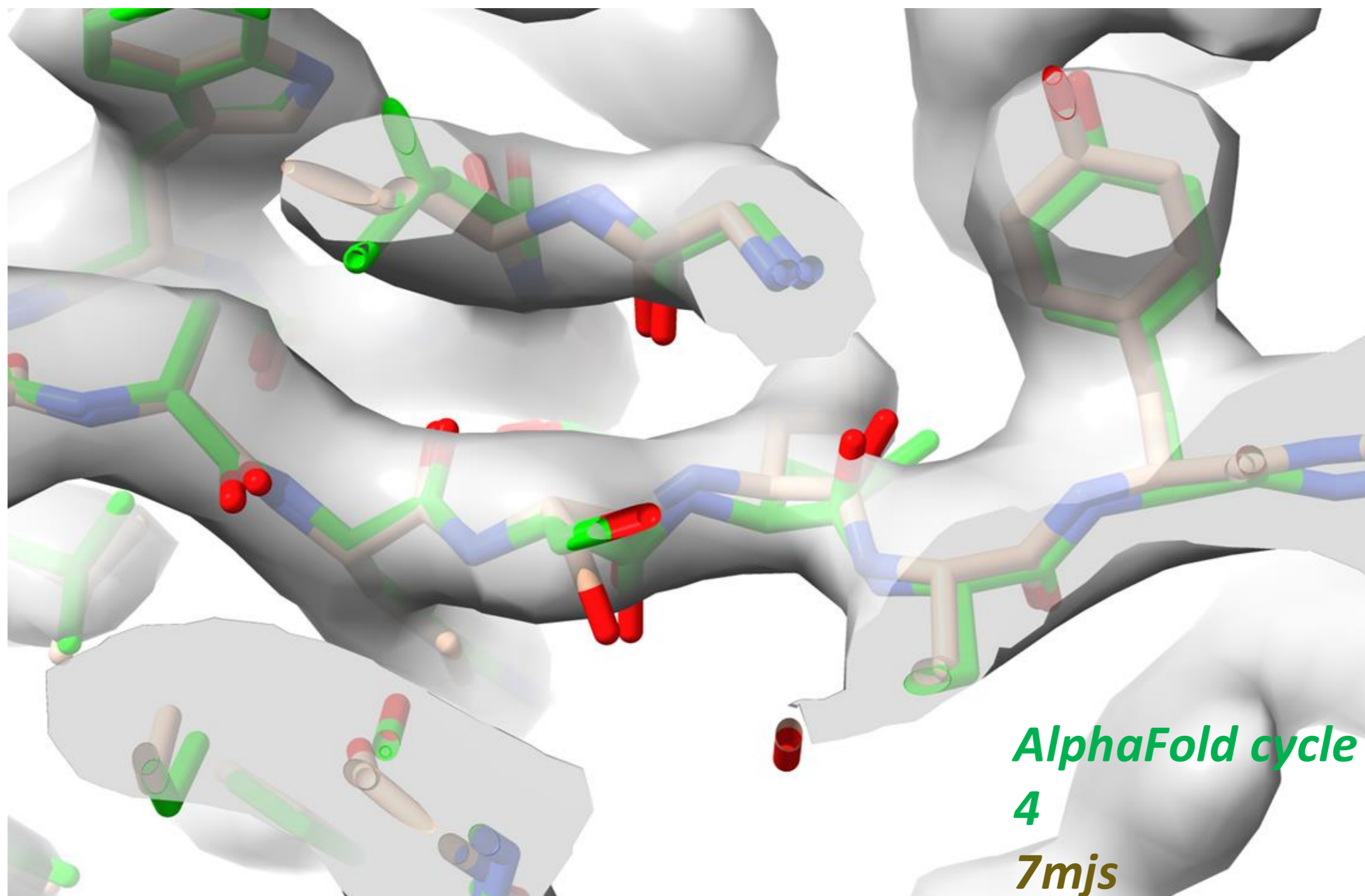
# Iterate prediction and rebuilding

---

Deposited  
model



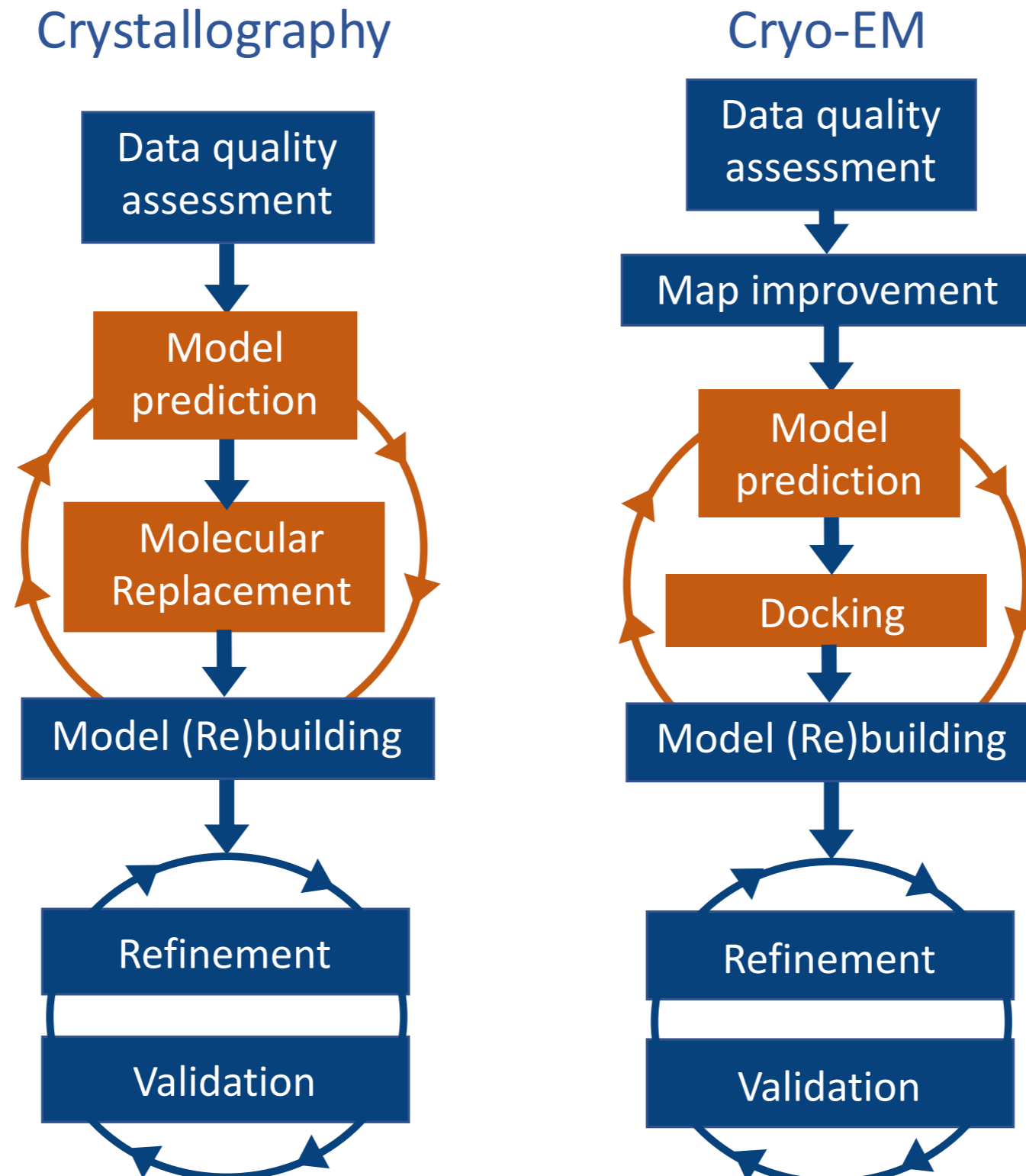
Final iterative predicted-  
rebuilt model



*Data from 7mjs, Cater, R.J., et al. (2021). Nature 595, 315–319*

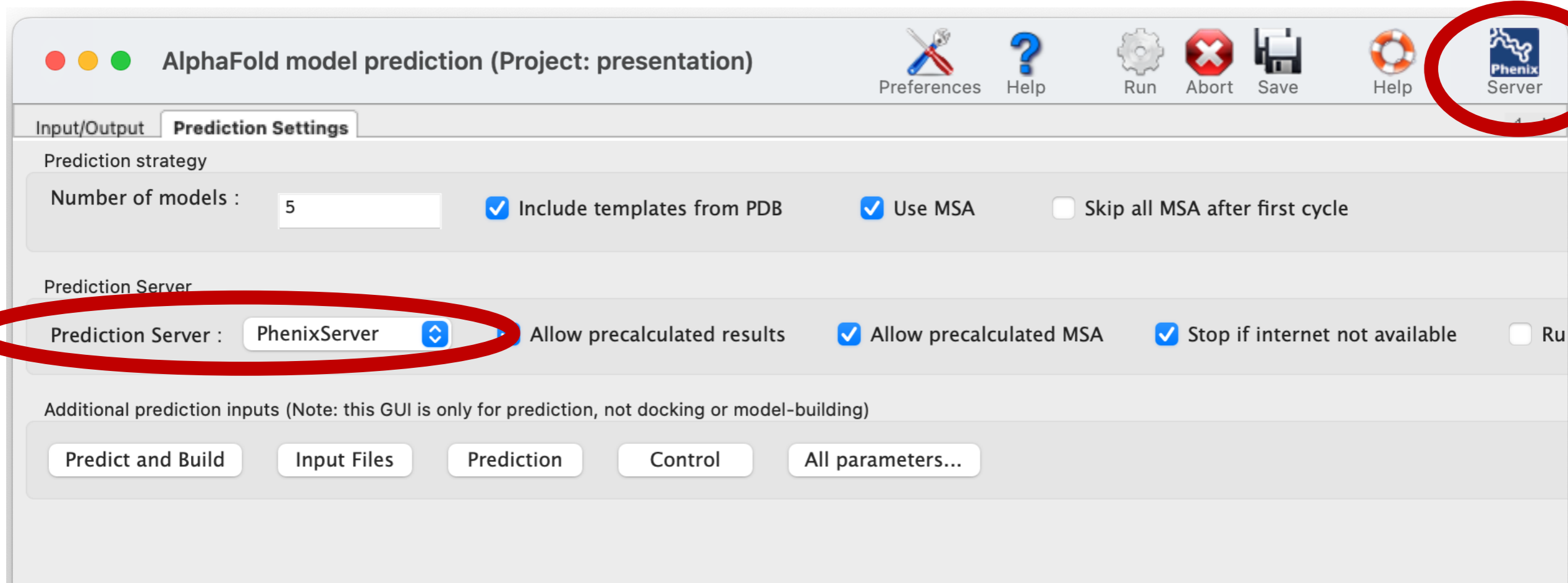
# Using predicted models

## Updated approach: Iterate prediction and model building

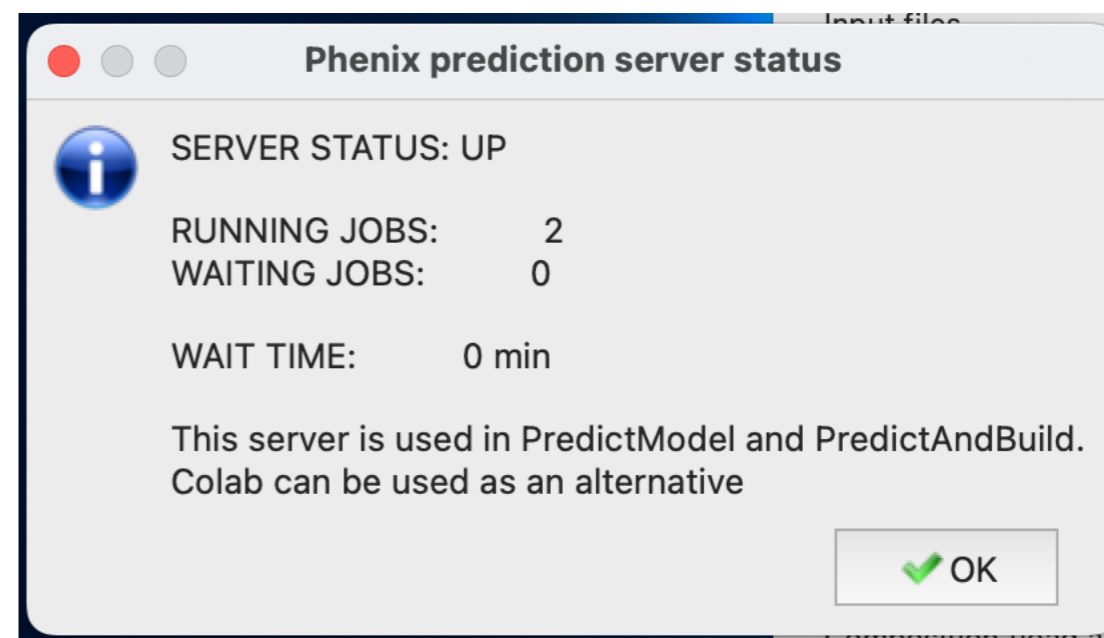




# Phenix Server for running AlphaFold



No need for a local  
AlphaFold installation



# Process predicted model

Process Predicted Model (Project: 7rpq\_AF\_reference\_model)

Preferences Help Run Abort Help

Configure

**ProcessPredictedModel: Prepare predicted model for structure determination**

Replace values in B-factor field with estimated B values.  
Optionally remove low-confidence residues and split into domains.

Inputs: Model file (PDB, mmCIF)

Job title :

**Predicted model**

Input

Note: The B-value field in your model can be pLDDT, RMSD, or B

Predicted model :  Browse...

Contents of B-value field for input models : plddt

**Optional input files**

Output

Output file prefix (optional) :

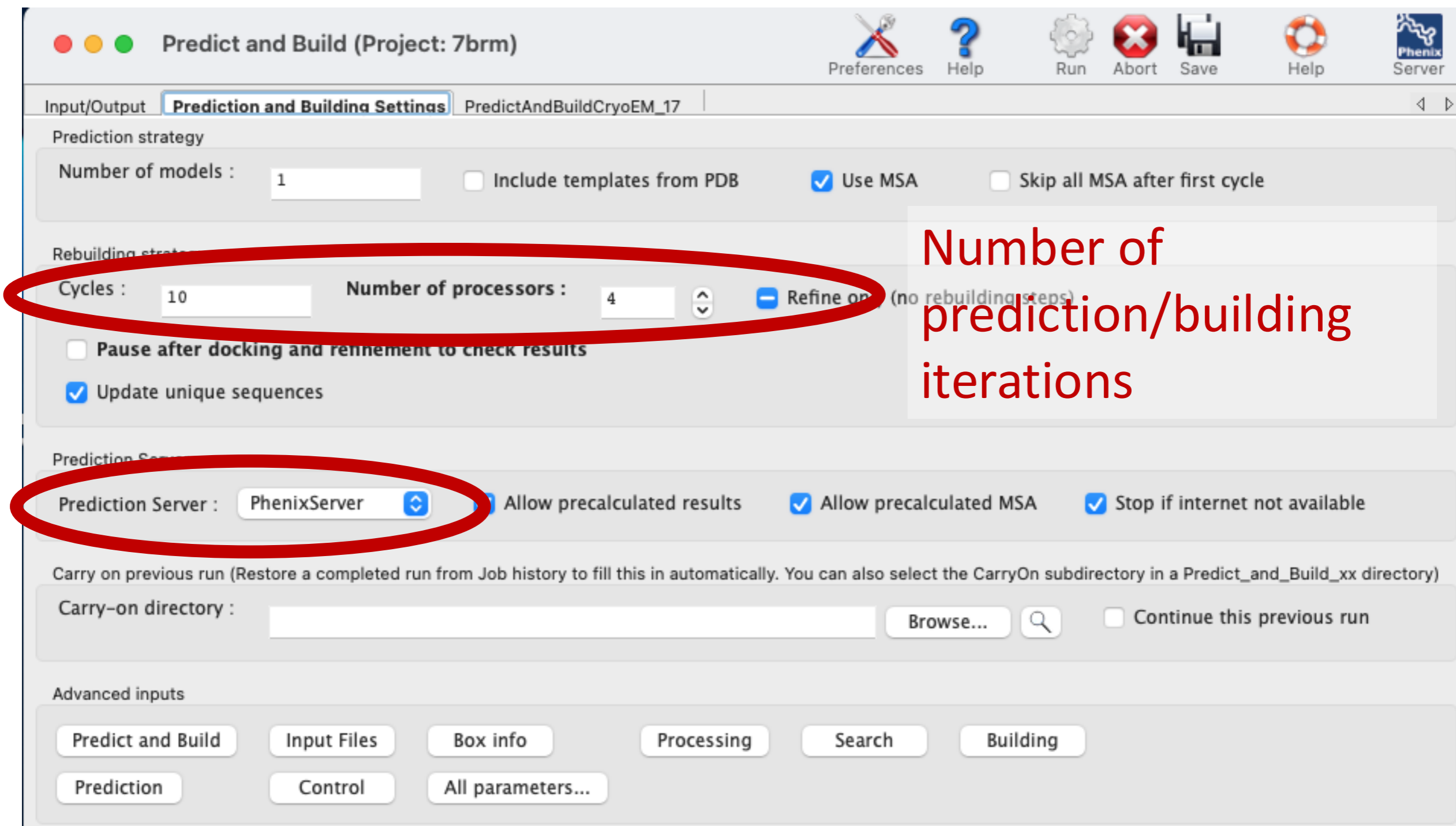
Options

Remove low-confidence residues  Split model into compact regions Maximum output B :

Remove hydrogen  Use only single-letter chain ID  Maintain continuous chain

Processing options All parameters...

# Iterate with Predict and Build



The screenshot shows the Phenix Predict and Build interface for a project named '7brm'. The window title is 'Predict and Build (Project: 7brm)'. The interface includes a toolbar with icons for Preferences, Help, Run, Abort, Save, and Help, along with the Phenix Server logo. The main area is divided into several sections:

- Prediction strategy:** Number of models: 1, Include templates from PDB (unchecked), Use MSA (checked), Skip all MSA after first cycle (unchecked).
- Rebuilding strategy:** Cycles: 10, Number of processors: 4, Refine on (no rebuilding steps) (checked), Pause after docking and refinement to check results (unchecked), Update unique sequences (checked).
- Prediction Server:** Prediction Server: PhenixServer (selected), Allow precalculated results (checked), Allow precalculated MSA (checked), Stop if internet not available (checked).
- Carry-on directory:** Carry-on directory: (empty), Browse... (button), Continue this previous run (unchecked).
- Advanced inputs:** Predict and Build, Input Files, Box info, Processing, Search, Building, Prediction, Control, All parameters... (button).

Two red ovals highlight the 'Cycles' field (set to 10) and the 'Prediction Server' dropdown (set to PhenixServer). A red text box on the right side of the interface reads: 'Number of prediction/building iterations'.

Fully automatic – AF prediction, processing, building, refinement.

# Strategy for structure determination

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## 1. Predict your structure

Design your experiment accordingly

(choose experimental approach, consider trimming at domain boundaries)

## 2. Solve your structure

Cryo-EM: docking

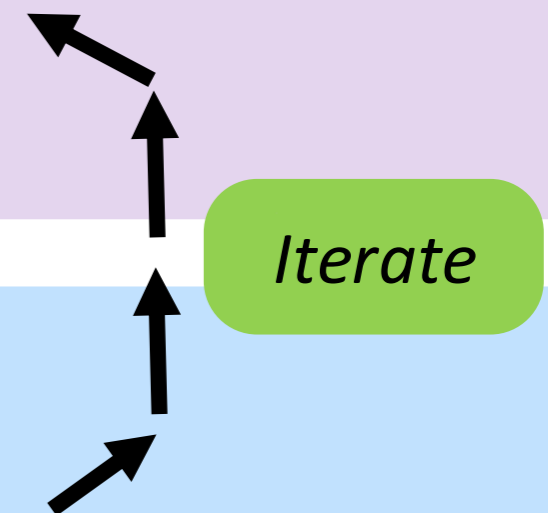
X-ray: MR; SAD

## 3. Update your prediction

Run AlphaFold again with your best model as a template

## 4. Improve your structure

Use your new prediction as hypothesis, rebuild parts



# Further reading/material

## Documentation:

<https://www.phenix-online.org/documentation/>

### Working with AlphaFold2, RoseTTAFold and other predicted models

You can use the predicted models from AlphaFold and other prediction software in Phenix. Using these models can be very helpful in structure determination because the models can be very accurate over much of their length and the models come with accuracy estimates that allow removal of poorly-predicted regions.

### How to use predicted models in Phenix

Use [Predict and build](#) to incorporate predicted models in the structure solution workflow.

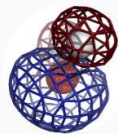
Predict\_and\_build generates predicted models and uses them to solve an X-ray structure by MR or to interpret a cryo-EM map. The tool then carries out iterative model rebuilding and prediction to improve the models. The iterative procedure allows creating more accurate predicted models than can be obtained with a simple prediction.

Predict and build can automatically generate a fairly accurate model starting from just a sequence file and either cryo-EM half-maps or X-ray data. Additionally, it provides morphed versions of unrefined predicted models that can be useful as reference models for refinement.

### Other tools for using predicted models:

- [Overview](#): AlphaFold and Phenix
- [Processing](#) a predicted model
- [Docking](#) a processed predicted model in a cryo-EM map
- [Rebuilding](#) a docked predicted model in a cryo-EM map
- [Processing, docking and rebuilding](#) a predicted model in a cryo-EM map
- [Trimming overlapping parts](#) of models

## Phenix Tutorials



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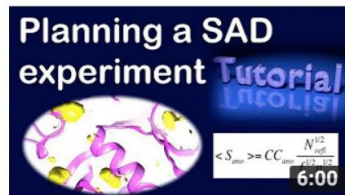
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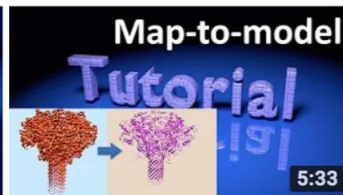


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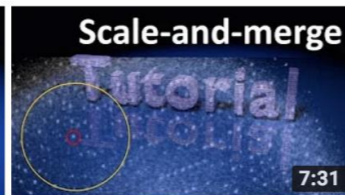
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Automatic map interpretation with map\_to\_model



Scaling and merging anomalous data



Automated map sharpening with phenix.autosharpen

## Video tutorials

<https://www.youtube.com/c/phenixtutorials>

AlphaFold changes everything:  
Incorporating predicted models in X-ray and Cryo-EM structure determination

Presentation available at:  
[phenix-online.org/presentations](https://phenix-online.org/presentations)

January 18, 2022  
RéNaFoBiS Webinar  
Tom Terwilliger  
The New Mexico Consortium  
Los Alamos National Laboratory

Randy Read, Tristan Croll, Claudia Millán, (Cambridge, University), Paul Adams, Billy Poon, Pavel Afonine, Christopher J. Schlicksup (Lawrence Berkeley National Laboratory); Jane Richardson (Duke University)

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New Mexico CONSORTIUM

Tom Terwilliger: AlphaFold changes everything

## Tom Terwilliger: AlphaFold changes everything

<https://youtu.be/ugMPYdPo8Bc?feature=shared>

# The Project



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Program Project

Liebschner D, *et al.*, Macromolecular structure determination using X-rays, neutrons and electrons: recent developments in *Phenix*. *Acta Cryst.* 2019 **D75**:861–877