

# Ligands in Phenix

Generating & modifying for all scenarios

Nigel W. Moriarty

User's Meeting  
ACA, July 2025

# What are restraints?

- Restraints are harmonic functions that provide (mostly) chemical information via residuals & gradients
- Needed because experimental data is rarely sufficient for structure determination
- Weighted with the experimental information to find the optimal result

# Resolution dependence

- Ultra-hi res – Not needed
- Hi res – Can have large deviations because the experimental data dominates
- Lo res – Generally approaches the ideal values
  - If not, large scale problems

# Restraints in Action

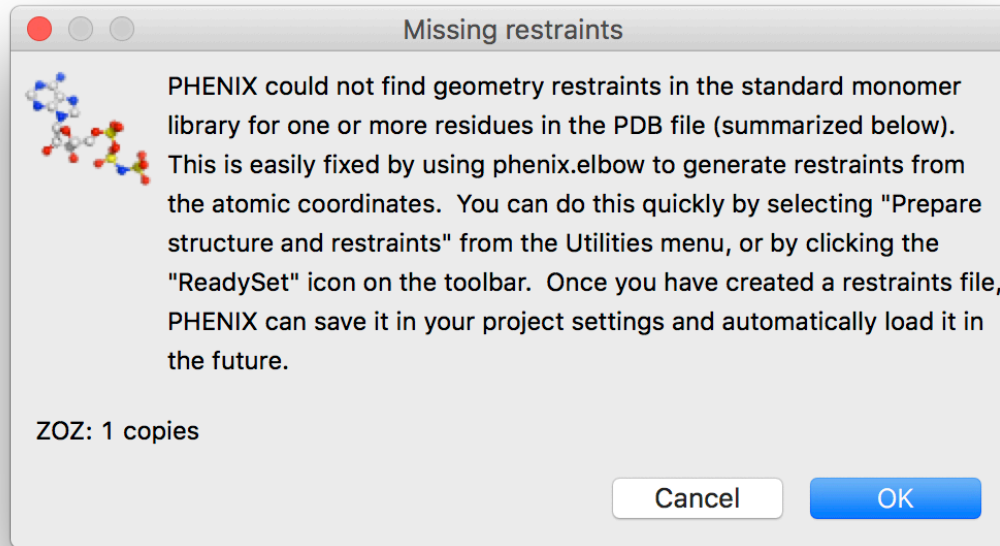
- Libraries
  - Monomer Library
  - GeoStd
- Algorithms
  - Polymer
  - Links



# GeoStd

- All standard amino acids
- Current list of non-standard amino acids
- All standard RNA/DNA
- Current list of non-standard RNA/DNA
- Others – 37k Mogul validated restraints using PBEh-3c/CPCM and higher QM

# What you will see



# Or

Sorry: Fatal problems interpreting model file:

Number of atoms with unknown nonbonded energy type symbols: 21

Please edit the model file to resolve the problems and/or supply a CIF file with matching restraint definitions, along with `apply_cif_modification` and `apply_cif_link` parameter definitions if necessary.

# CIF

- Crystallographic Information File
- mmCIF – macro-molecular CIF
- Used for
  - Model
  - Data
  - Maps
  - Ligands
    - Information
    - Restraints

# Confusion

- All depositions of X-ray model use mmCIF from 1 July 2019
- “I need a CIF file.”
  - But what do you really need?

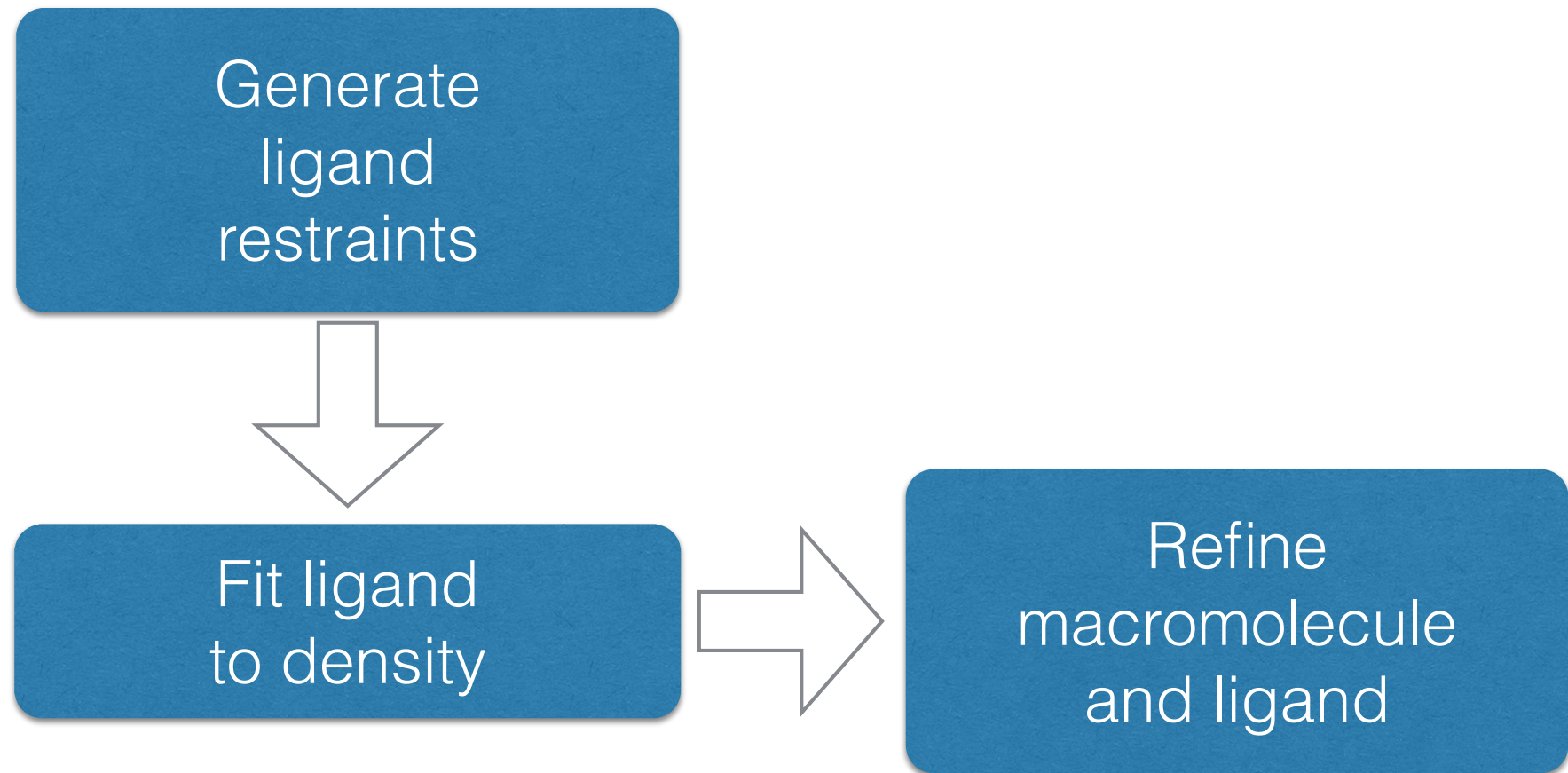
# Restraints?

- Provide a reasonable geometry during refinement particularly at low resolution
  - Bonds, angles, dihedrals, chirals, planes, ...
- Must be weighted against the experimental information

# Overview

- eLBOW - electronic Ligand Builder & Optimisation Workbench
- ReadySet! - One-stop preparation for your refinement needs
- REEL - Restraints Editor Essentially Ligands

# Ligands in crystallography

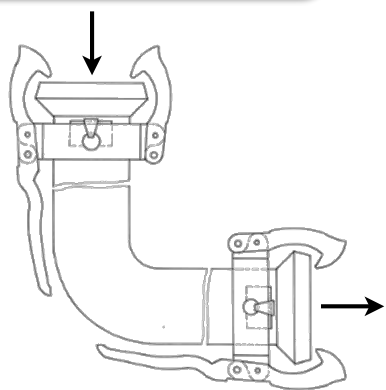




# eLBOW goals

- Fast, simple and flexible procedure to include ligands
- Reduce the tedium of building 3D ligand models
- Automate generation of restraints for ligands
- Comparison of ligand structures

Chemical input



Chemical restraints (CIF)  
Cartesian coordinates (PDB)

Reflection data  
Protein information

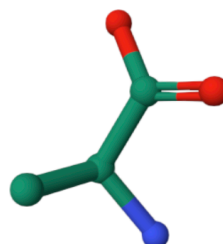
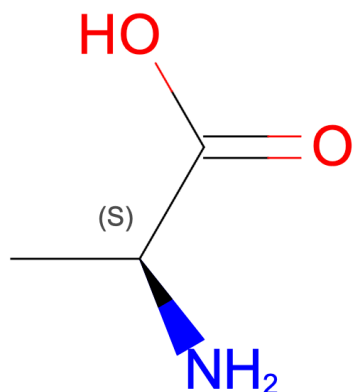
refinement

N. W. Moriarty, R. W. Grosse-Kunstleve, P. D. Adams, (2009). Acta Cryst. D 65, 1074-1080.

# Ligand Expo / Chemical Component Library

- List of all the entities in the Protein Data Bank
  - Amino acids, Nucleic acids
  - Ligands, Small molecule
  - Metal clusters
- In CIF format
  - Contents chemical information
    - SMILES, atom names, bonds
  - Not restraints

# Amino Acid



Toggle Hydrogen

Toggle Labels

 Display Files ▼

 Download Files ▼

 Data API

## ALA

### ALANINE

Find entries where: ALA

- ☒ is present as a standalone ligand in [172 entries](#)
- ☒ as a non-polymer is covalently linked to polymer or other heterogen groups [58 entries](#)
- ☒ is present in a polymer sequence [210,908 entries](#)

Find related ligands:

[Similar Ligands \(Stereospecific\)](#)

[Similar Ligands \(including Stereoisomers\)](#)

[Similar Ligands \(Quick Screen\)](#)

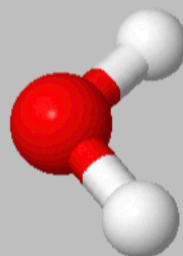
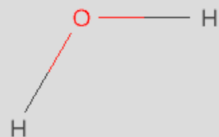
[Similar Ligands \(Substructure Stereospecific\)](#)

[Similar Ligands \(Substructure including Stereoisomers\)](#)

# Water

**Chemical Details** **Geometry** **Atom Nomenclature** **Downloads** **Related Resources**

## PDB Chemical Component HOH



JSmol

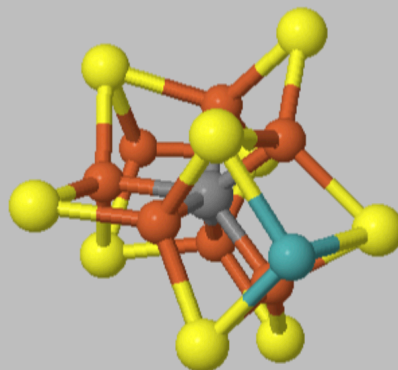
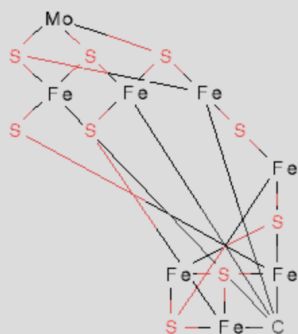
Ideal Model

## Chemical Description

**Name** WATER  
**Formula** H2 O  
**Formal charge** 0  
**Molecular weight** 18.015 g/mol  
**Component type** NON-POLYMER

# Metal clusters

## PDB Chemical Component ICS



JSmol

Ideal Model

# 5-letter codes

**RCSB PDB** PROTEIN DATA BANK

217,157 Structures from the PDB  
1,068,577 Computed Structure Models (CSM)

▼ 3D Structures ? Enter search term(s), Entry ID(s), or sequence Include CSM ?

[Advanced Search](#) | [Browse Annotations](#) [Help](#)

PDB-101

[Display Files ▼](#) [Download Files ▼](#) [Data API](#)

## A1LU6

5-(3-azanyl-1~{H}-indazol-6-yl)-1-[(3-chlorophenyl)methyl]pyridin-2-one

Find entries where: A1LU6  
☒ is present as a standalone ligand in **1** entries  
[search](#)

Find related ligands:  
[Similar Ligands \(Stereospecific\)](#)  
[Similar Ligands \(including Stereoisomers\)](#)  
[Similar Ligands \(Quick Screen\)](#)  
[Similar Ligands \(Substructure Stereospecific\)](#)  
[Similar Ligands \(Substructure including Stereoisomers\)](#)

[Toggle Hydrogen](#) [Toggle Labels](#)

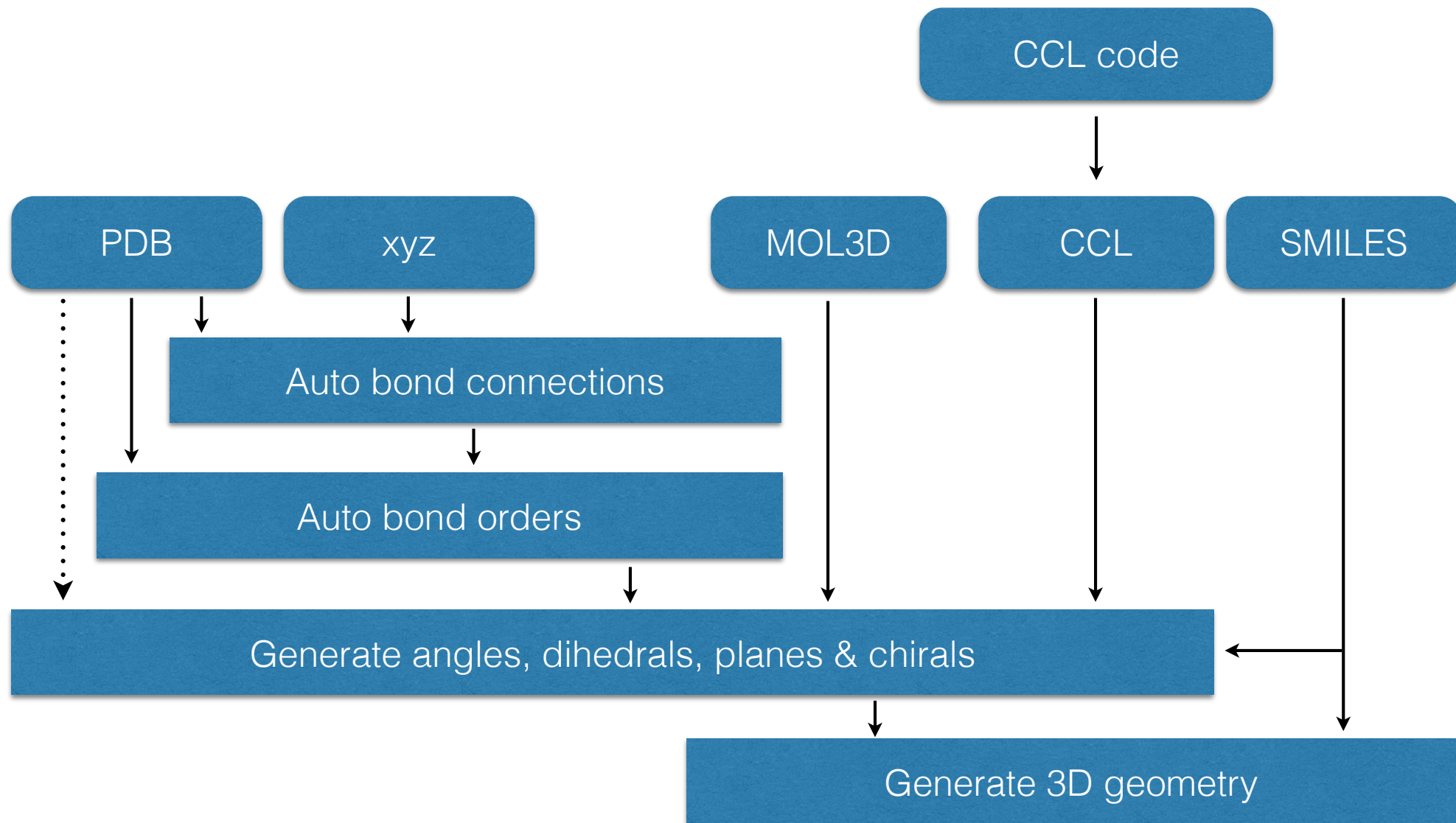
- 49k combinations for 3-letter codes

# Human readable

Standard		Human readable	
Uppercase	Lowercase	Uppercase	Lowercase
10I0	1oi0	1oi0	1oi0
1IJJ	1ijj	1iJJ	1ijj
4OCL	4ocl	4oCL	4ocL
5SS2	5ss2	5ss2	5ss2

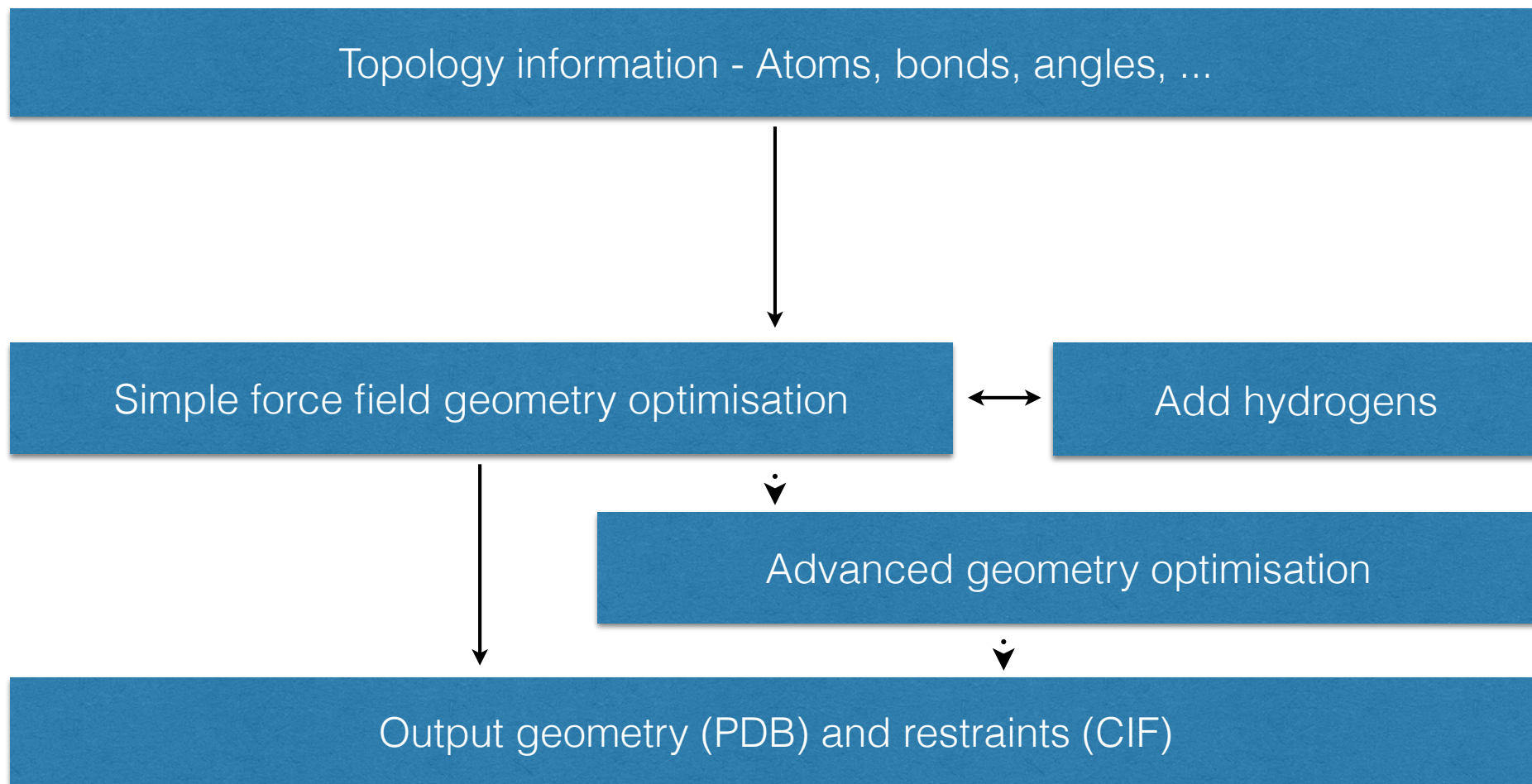
- Confusable letters are case-forced
  - L is always uppercase
  - i, o are always lowercase
- Somewhat confusable – “5” and “S”

# Topology





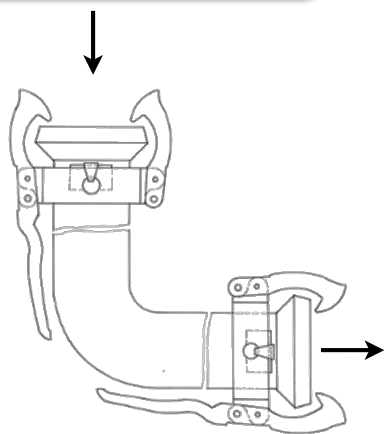
# Optimisation



# Getting ready to refine

- Many details needed to prepare for structure refinement

Chemical input



Chemical restraints (CIF)  
Cartesian coordinates (PDB)

Protein Information

ReadySet!

Experimental data

refinement

# ReadySet!

- Add hydrogens
  - Default: adds hydrogens to protein, ligands
    - Protein - Reduce
    - Ligands - eLBOW
  - Add hydrogens to water
  - Add deuteriums instead of hydrogens
  - Add hydrogen & deuteriums appropriately
- Generate restraints

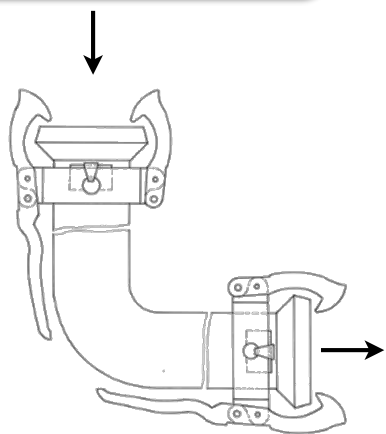
# ReadySet!

- Restraints CIF filename
- Restraints CIF directory
- LINKS to “edits”
- --dry-run to show ligand process pathway
- Metal coordination

# Restraints editing

- Visualisation of primary and secondary restraints
- Adjusting restraints to user preference

Chemical input



REEL

Chemical restraints (CIF)  
Cartesian coordinates (PDB)

Protein Information

ReadySet!

Reflection data

phenix.refine

**Phenix**

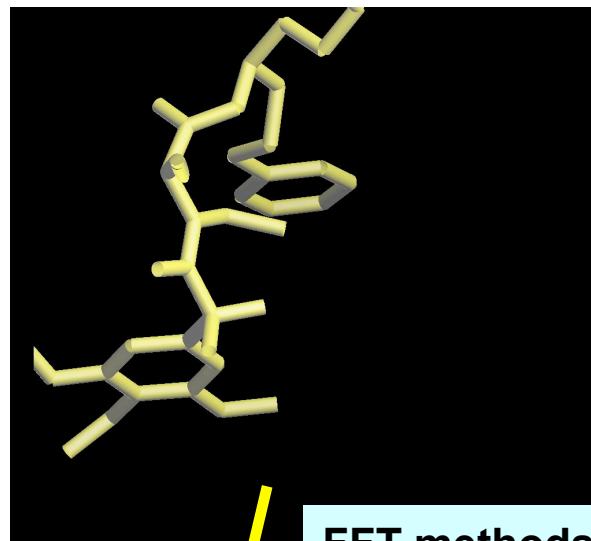
# Ligand Fitting

# Approach

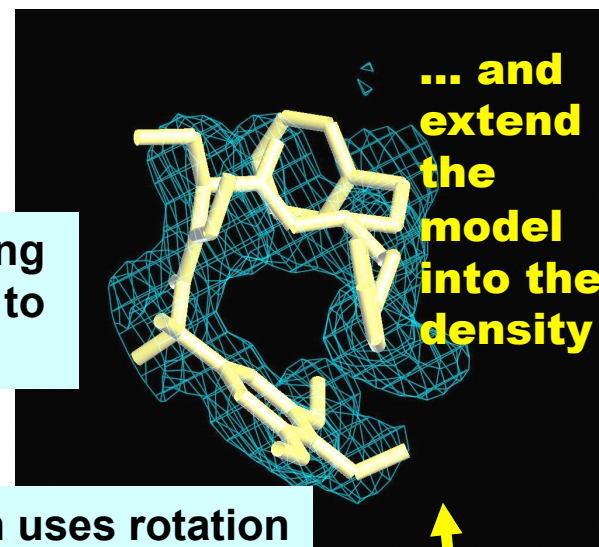
- Where is the ligand?
  - Choose the largest region of contiguous density
- What are rotatable bonds?
  - Analyze ligand for allowed rotations
- What is the orientation of the ligand?
  - Fit core of ligand
- What is the conformation of the ligand?
  - Trace the ligand out from the core



# Automated Ligand Fitting



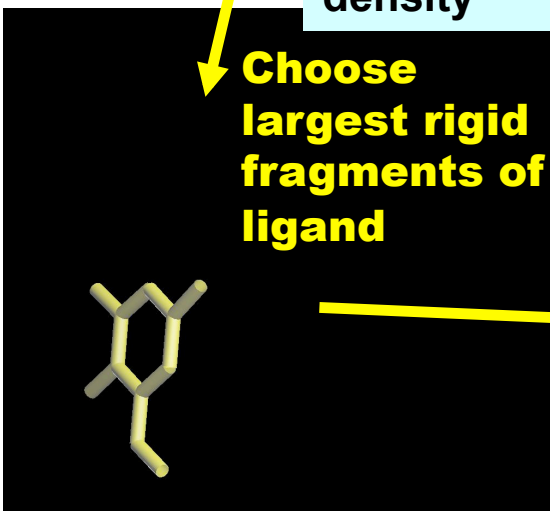
The best fit is measured using the correlation of the model to the electron density



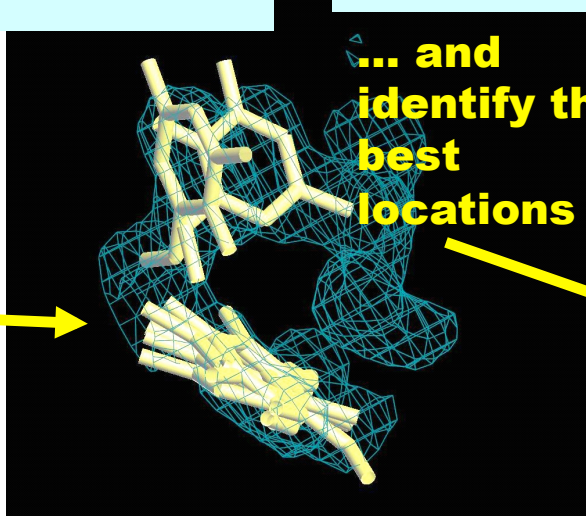
FFT methods are used to rapidly fit fragments to density

The extension uses rotation around torsion angles to find the best fit

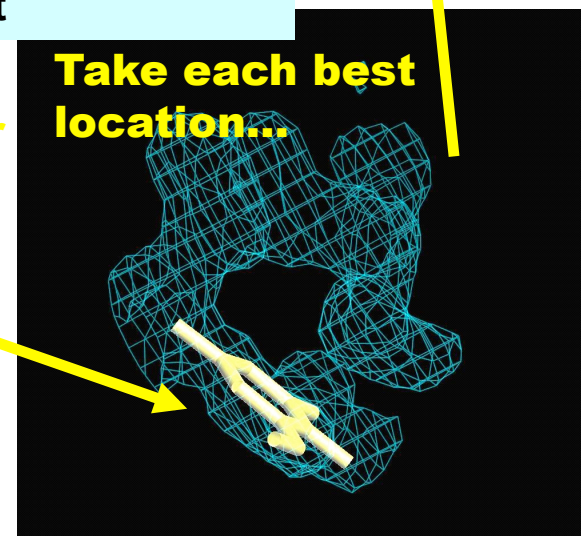
Choose largest rigid fragments of ligand



... and identify their best locations



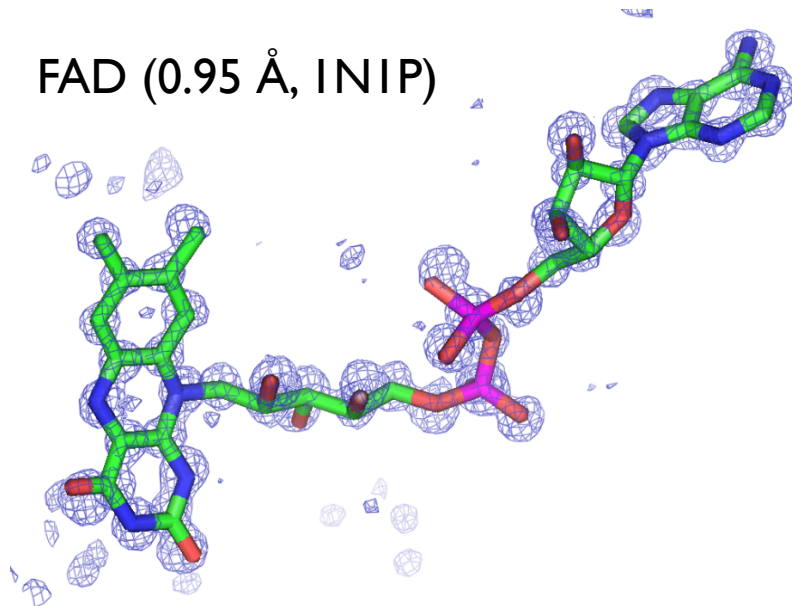
Take each best location...



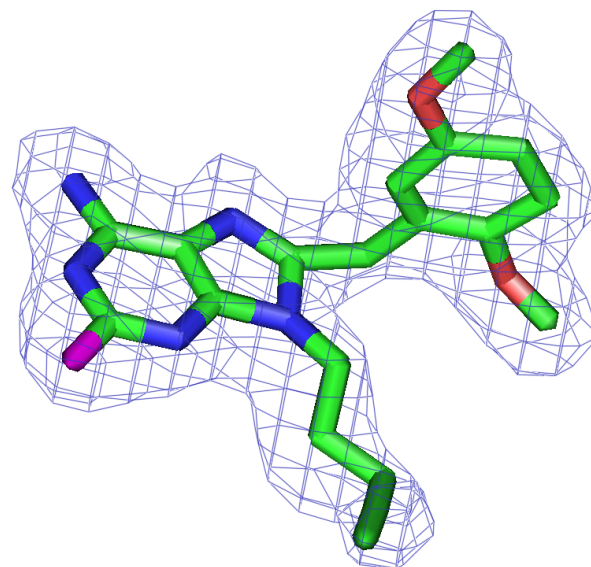


# Fitting Over a Range of Resolutions

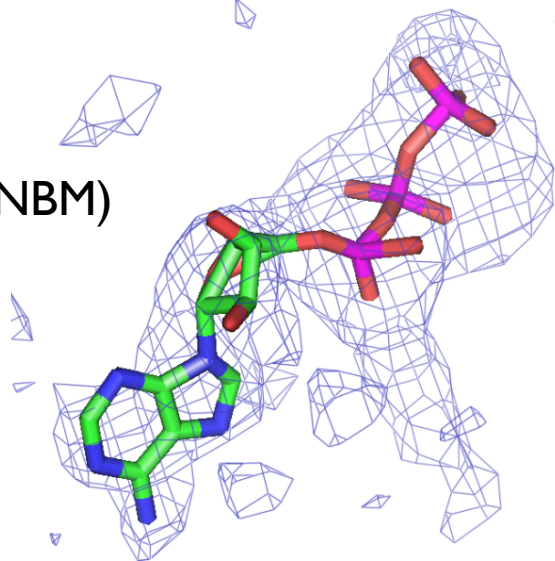
FAD (0.95 Å, 1NIP)



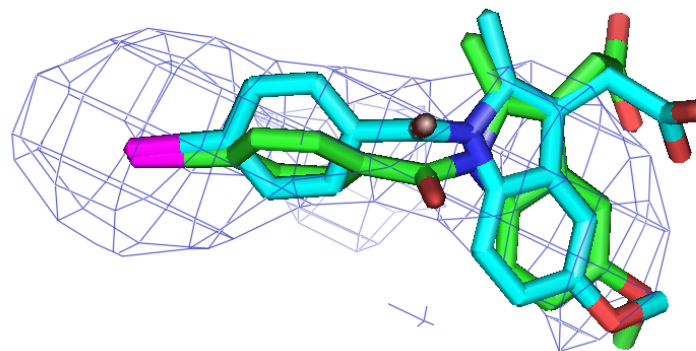
8-(2,5-DIMETHOXY-BENZYL)-2-FLUORO-9-PENT-9H-PURIN-6-YLAMINE (2.2 Å 1UYI)



ATP (3 Å, 1NBM)



(1-(4-IODOBENZOYL)-5-METHOXY-2-METHYL-INDOLE-3-ACETIC ACID (4.5 Å, 1PGF)



**Phenix**

# Restraints in phenix.(real\_space\_)refine

- LINK records have no impact
- Automatically accesses the “standard” residues restraints
- Automatically links the “standard” residues
- Parameter “link\_all=True” links
  - Covalent ligands
  - Carbohydrates
  - Metal ions

# phenix.(real\_space\_)refine (continued)

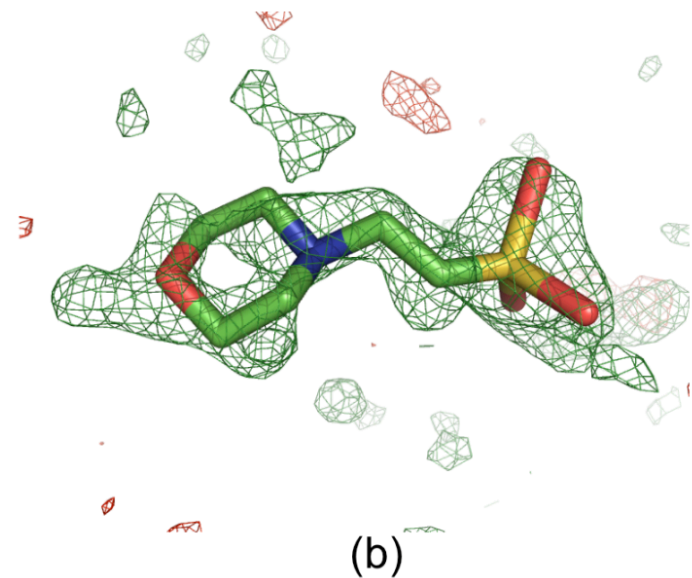
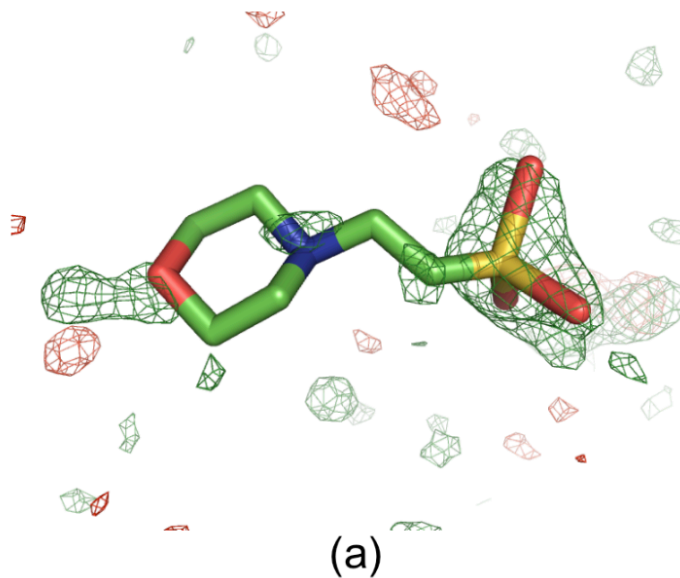
- RNA/DNA restraints
  - Base pair hydrogen bonding
  - Base pair planarity
  - Base stacking (parallelity)
- Secondary Structure restraints
- NCS restraints
- Custom bonds & angles using edits
- Restraints are written to .geo file including non bonded interactions

# Ligands? Water?

- Be careful when inheriting a model
- Can contain ligands and water molecules
- Must be able to “see” them in the map

# Ligands

# Polder OMIT Maps



Solvent molecule MES88 in structure 1ABA. The positive and negative mFobs-DFmodel OMIT difference density is displayed in green and red, respectively. (a) OMIT map contoured at  $\pm 3\sigma$ . (b) Polder map contoured at  $\pm 3\sigma$ . In the OMIT map, there is only some density for the O, N and S atoms. The polder map shows difference electron density for the entire molecule

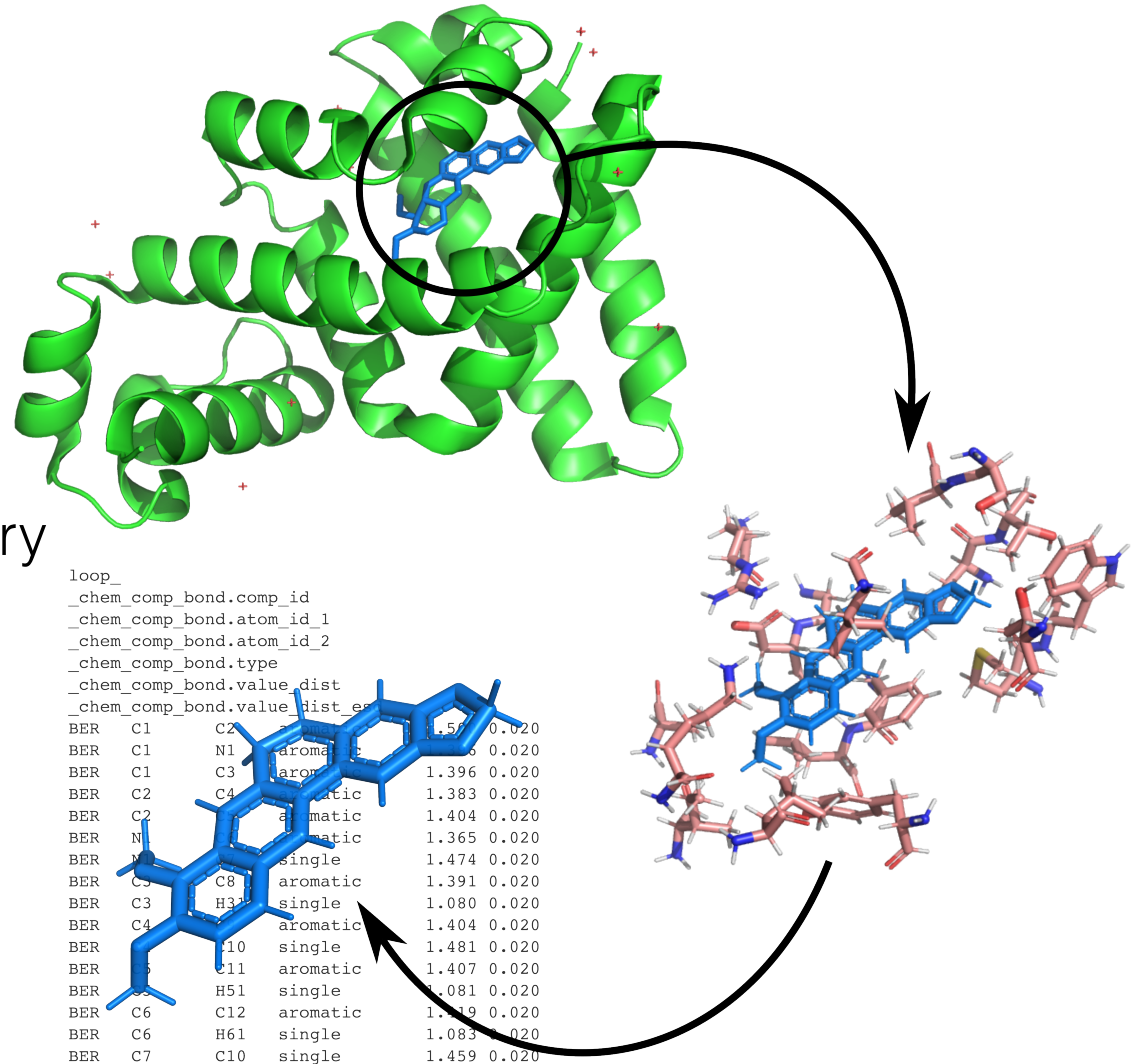
# QM Restraints

- Generates restraints of ligands using Quantum Mechanics *in situ*
- There are two ways of using QMR
  - In *phenix.refine*
  - In a standalone program *mmtbx.quantum\_interface*
- Python3 installers from the bottom of the download page
- For ORCA, set \$PHENIX\_ORCA

# QMR

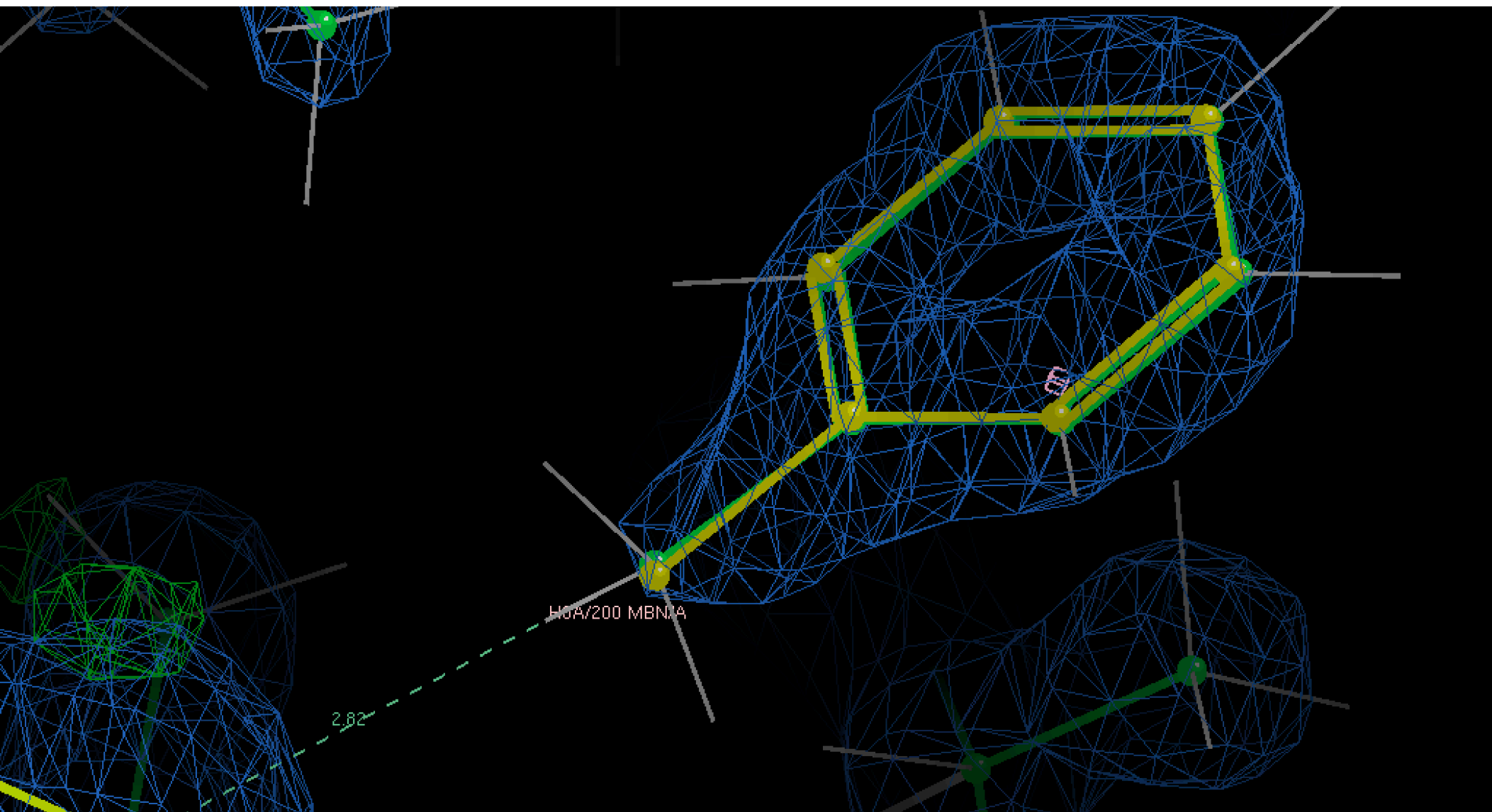
*In situ* restraints generation

- Carve out the ligand environment
- Minimise the ligand geometry *in situ*
- Transfer geometry values to restraints (and write to disk)
- Refinement with modified restraints

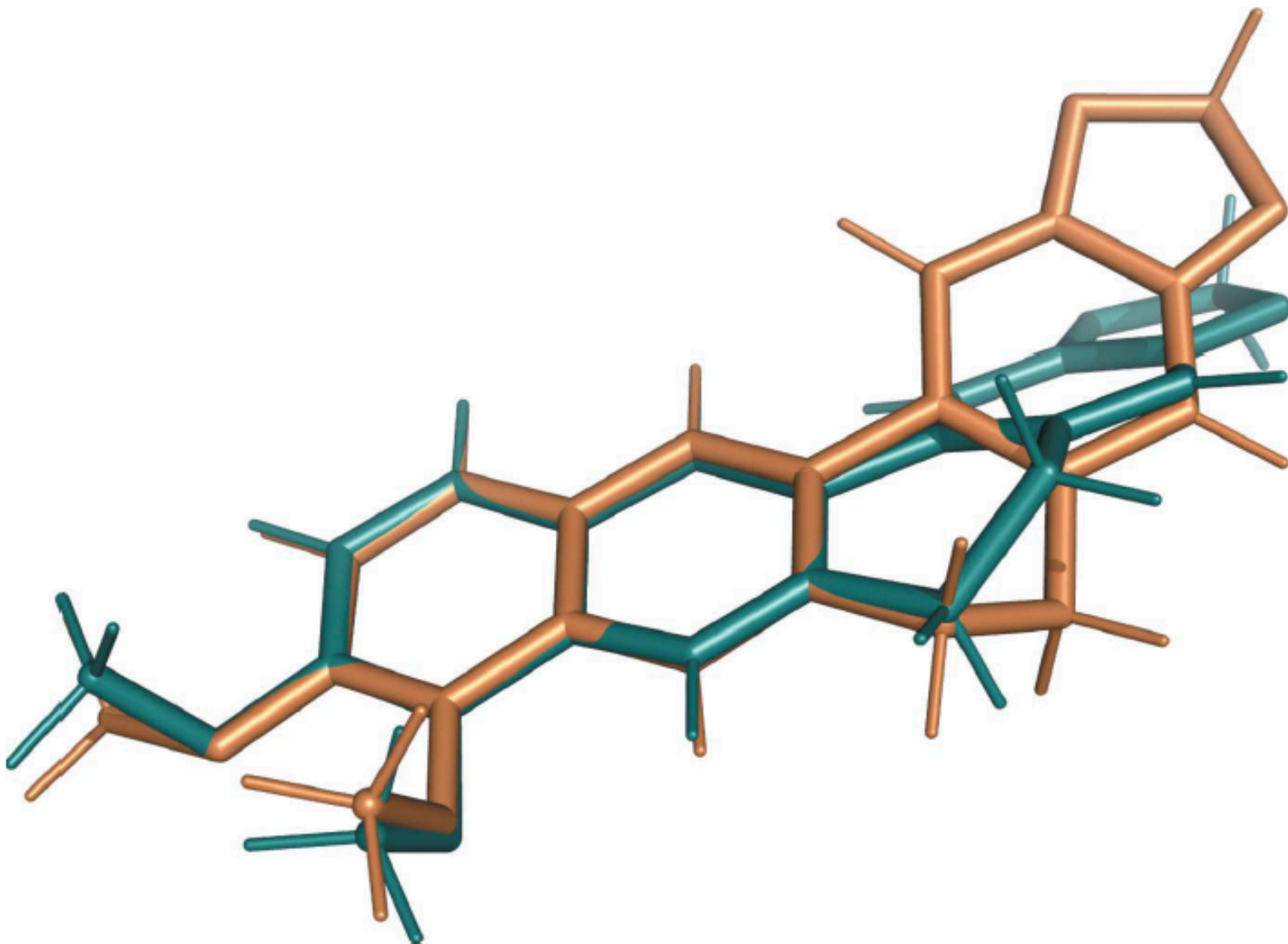




# *In situ*



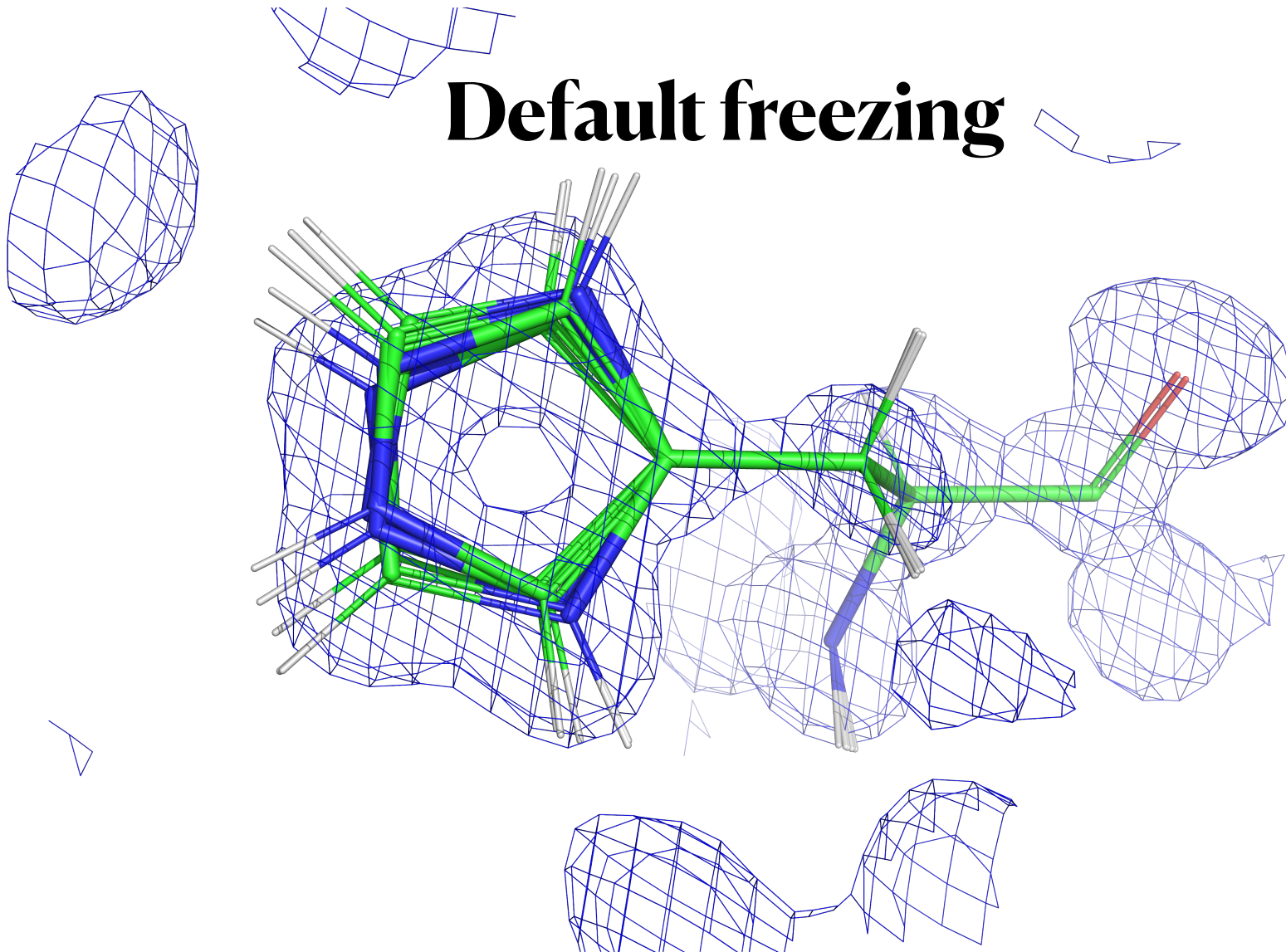
# BER in 3vw2



# QM Flipping

- Generate the three pronation states of HIS
- Flip  $\chi_2$  180 for total of six configurations
- Perform a QM geometry minimisation of the side-chain while freezing the heavy atoms (non-H) for rest of model
- Compare metrics
  - Energy
  - H-bonds
  - RMSD

# Default freezing

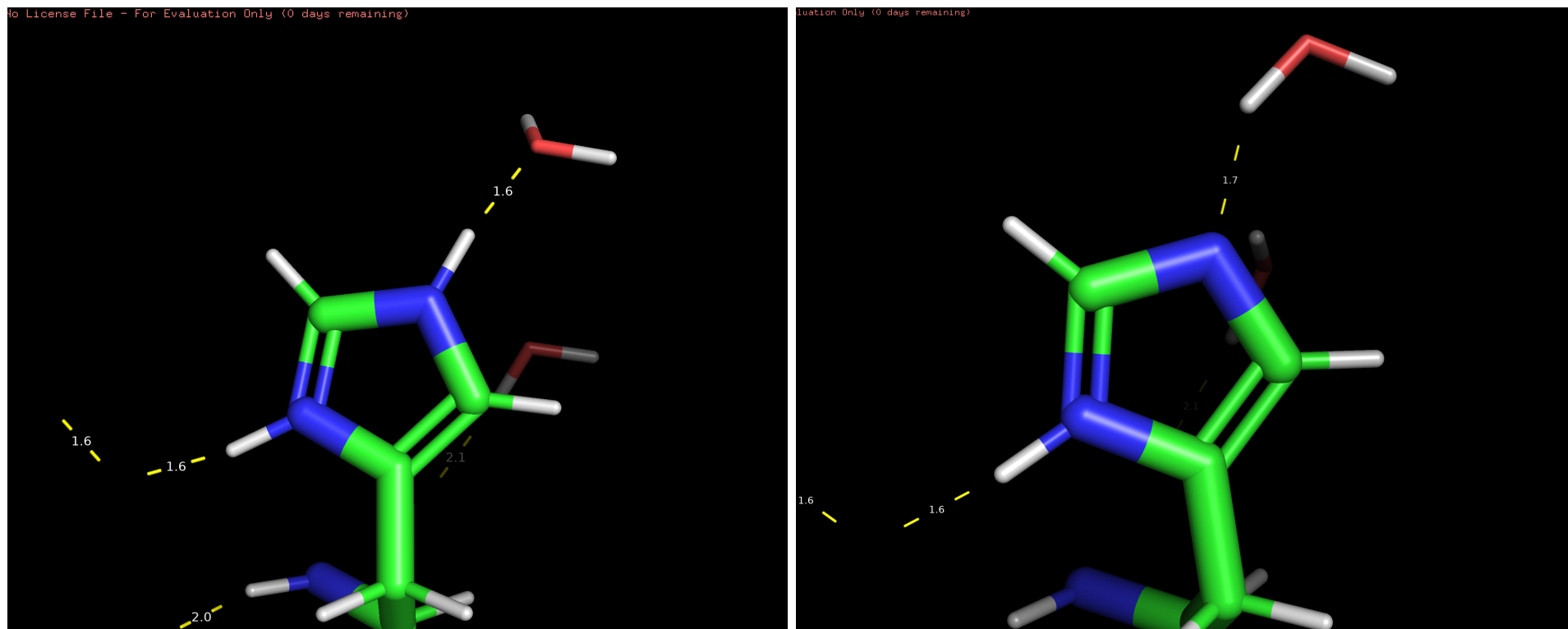


# Metrics

	Configuratio	Energy (kcal/	$\Delta E$	H bonds	r.m.s.d.	Rotamer
0	HD1, HE2					m90
1	HD1, HE2	-1019.6	2.6	14	0.04	m90
2	HD1 only	-1022.2	0.0	14	0.04	m90
3	HE2 only	-1003.0	19.3	13	0.05	m90
4	HD1, HE2	-1004.0	18.3	12	0.29	m-70
5	HD1 only	-1004.4	17.9	12	0.38	m-70
6	HE2 only	-1009.4	12.9	11	0.32	m-70

The QMF results for the histidine resseq 4 in chain A of PDB 4rj2

## Minimised geometry of two histidine configurations of histidine in chain A and resid 4 of PDB 4rj2.



# Summary

- eLBOW & ReadySet! perform better when provided with better input. (GIGO)
- Need to know something about the ligand
  - Hierarchy of input file value
- Check your .geo file for confirmation of restraints