

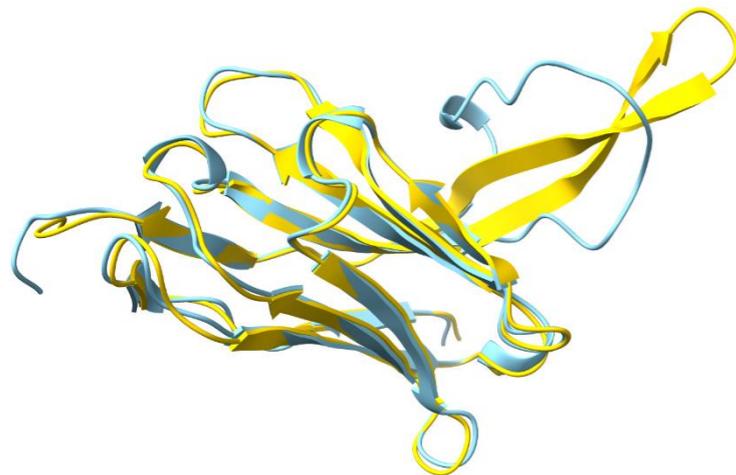
Strategy for structure determination using AlphaFold model

Phenix Workshop
July 18, 2025, ACA Lombard

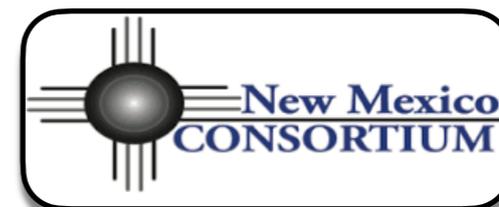
Slides by Tom Terwilliger

The New Mexico Consortium
Los Alamos National Laboratory

Presented by Christopher Williams,
with additional slides

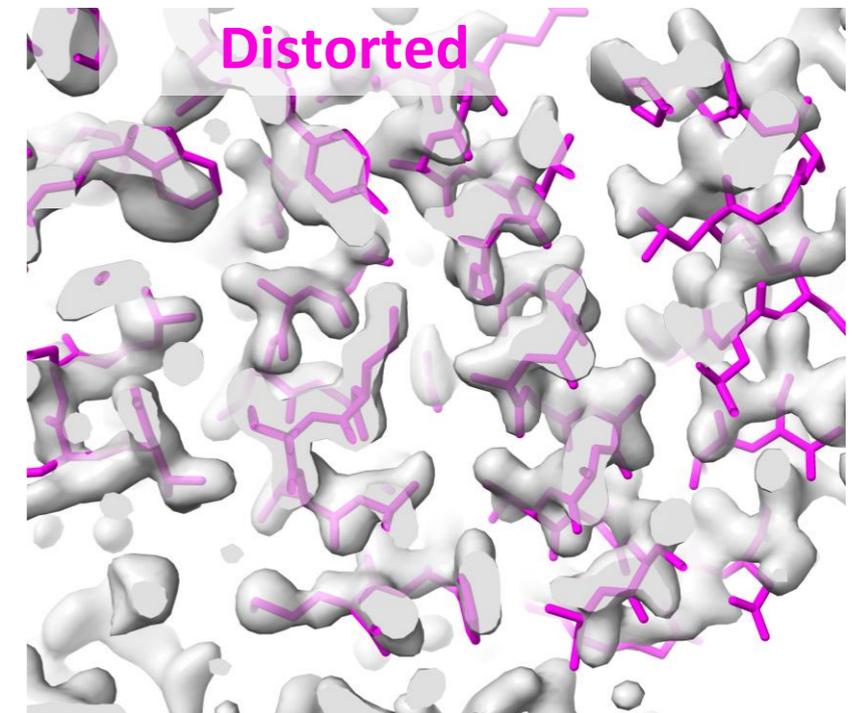
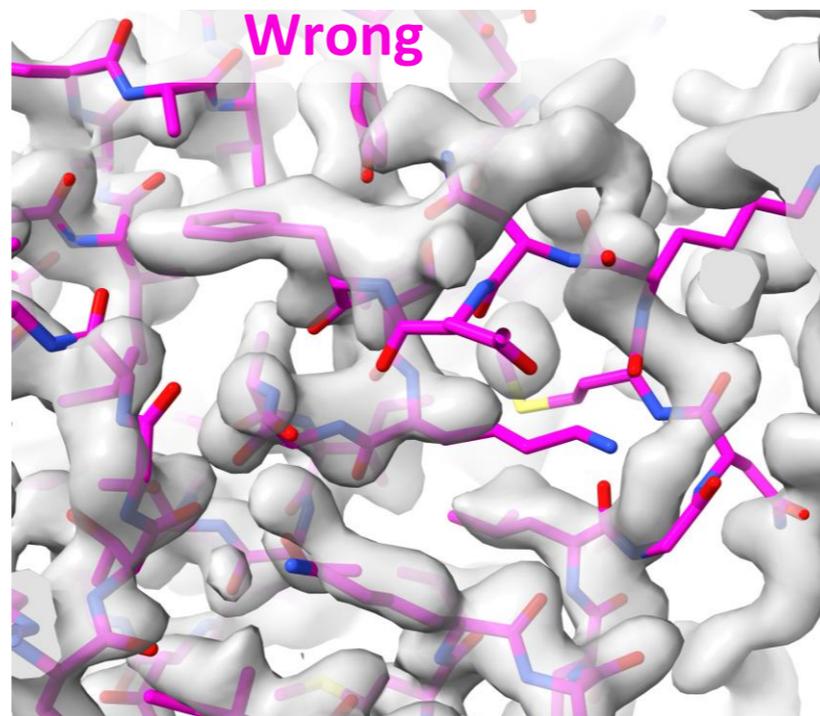
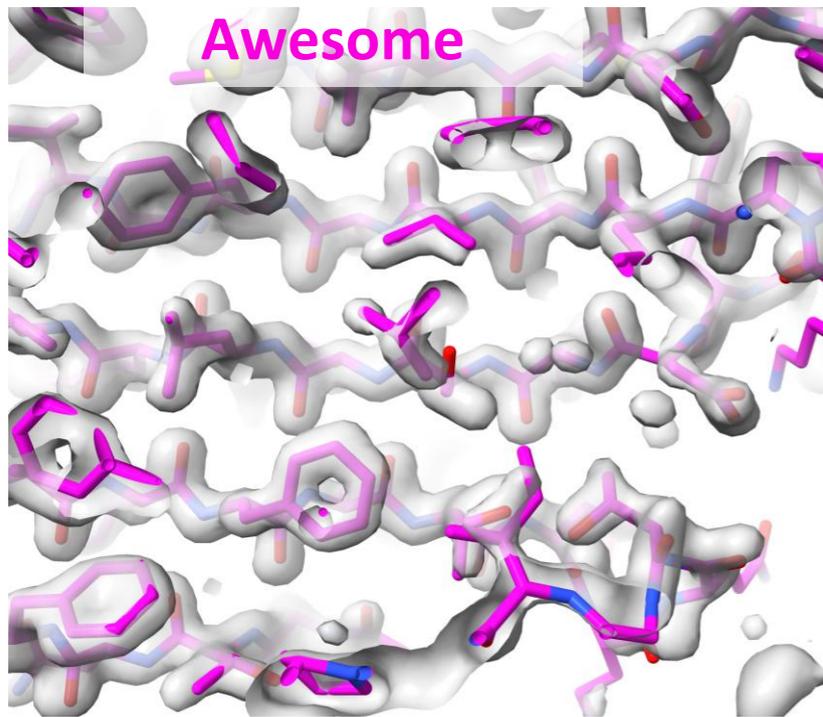


Richardson Lab
Duke University, Biochemistry Department



AlphaFold predictions are great hypotheses

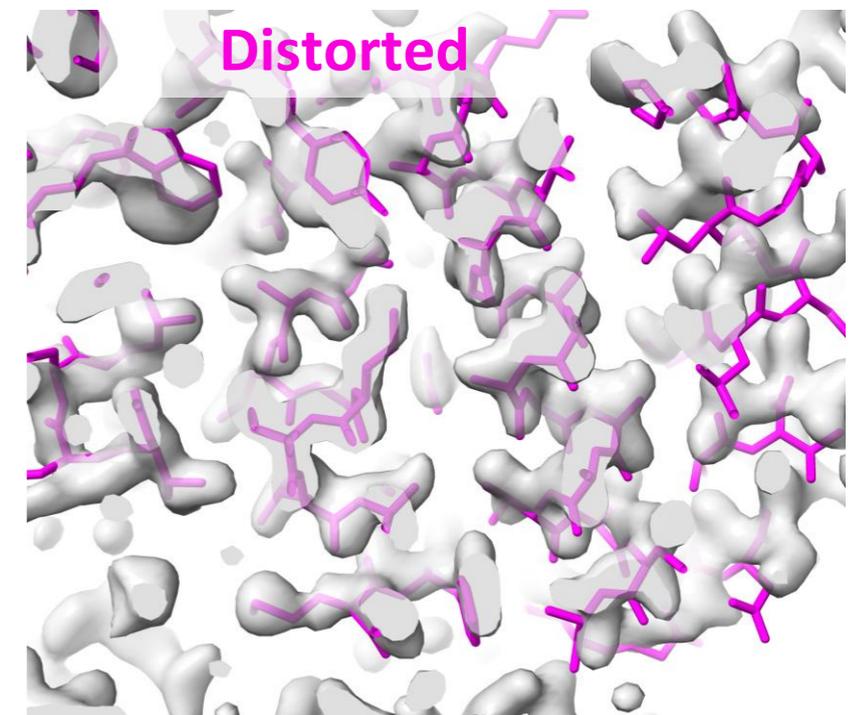
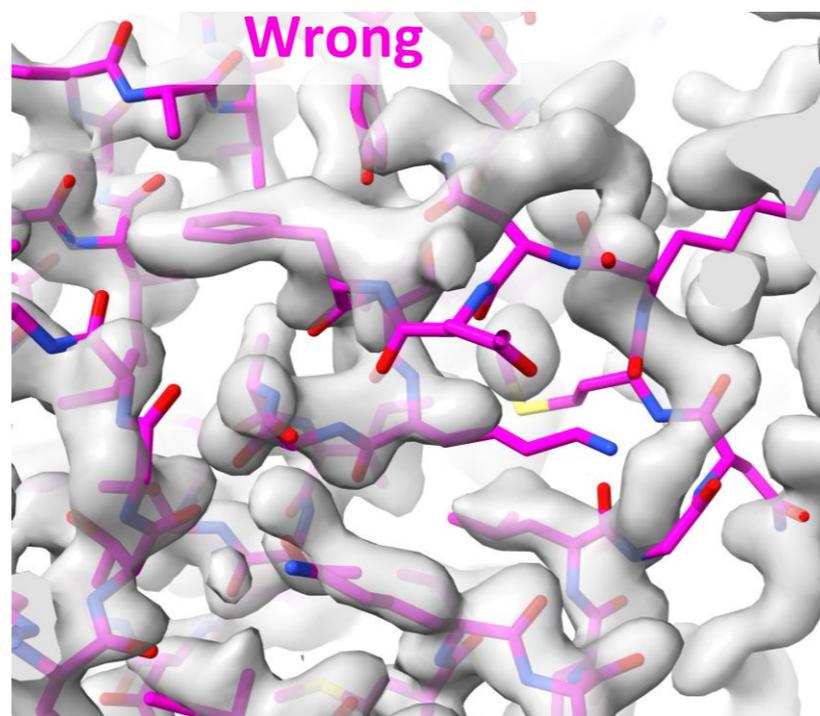
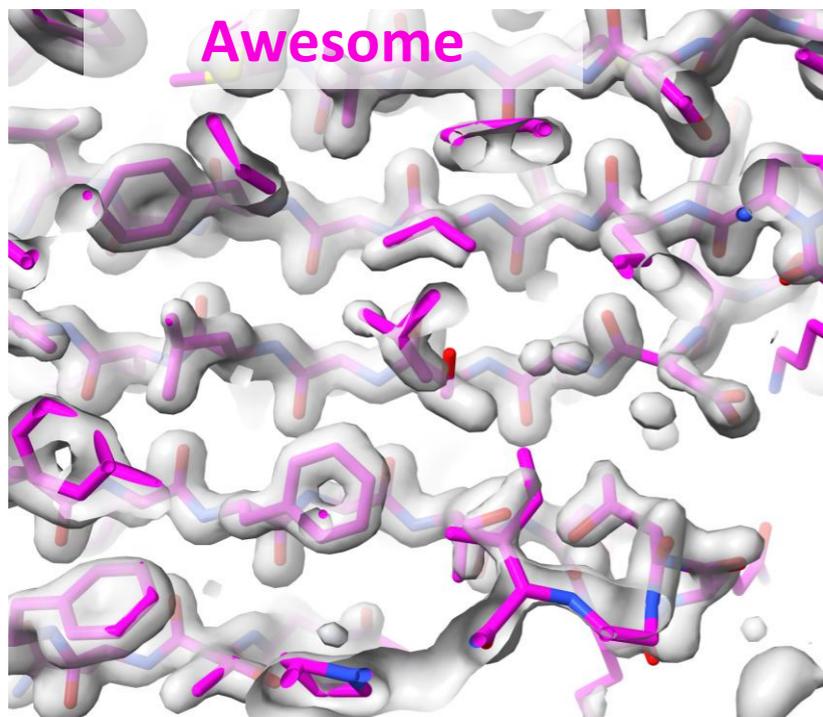
*AlphaFold models
can be*



AlphaFold predictions and confidence estimates

Residue-specific confidence (pLDDT) identifies where errors are more likely

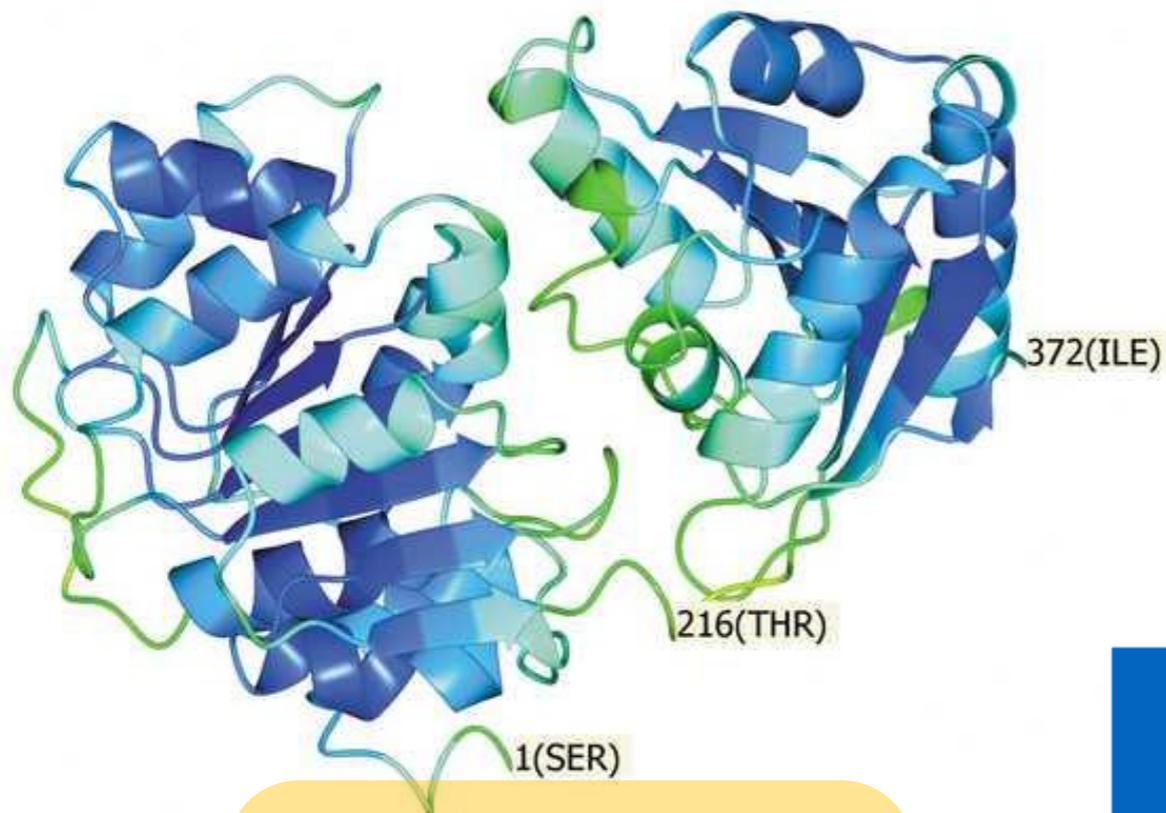
AlphaFold confidence (pLDDT)	Median prediction error (Å)	Percentage with error over 2 Å
>90	0.6	10
80 - 90	1.1	22
70 - 80	1.5	33
<70	3.5	77



Terwilliger, Thomas C., et al. "AlphaFold predictions are valuable hypotheses and accelerate but do not replace experimental structure determination." *Nature Methods* 21.1 (2024): 110-116.

AlphaFold confidence measure

(pLDDT, Predicted difference distance test)



Confidence:

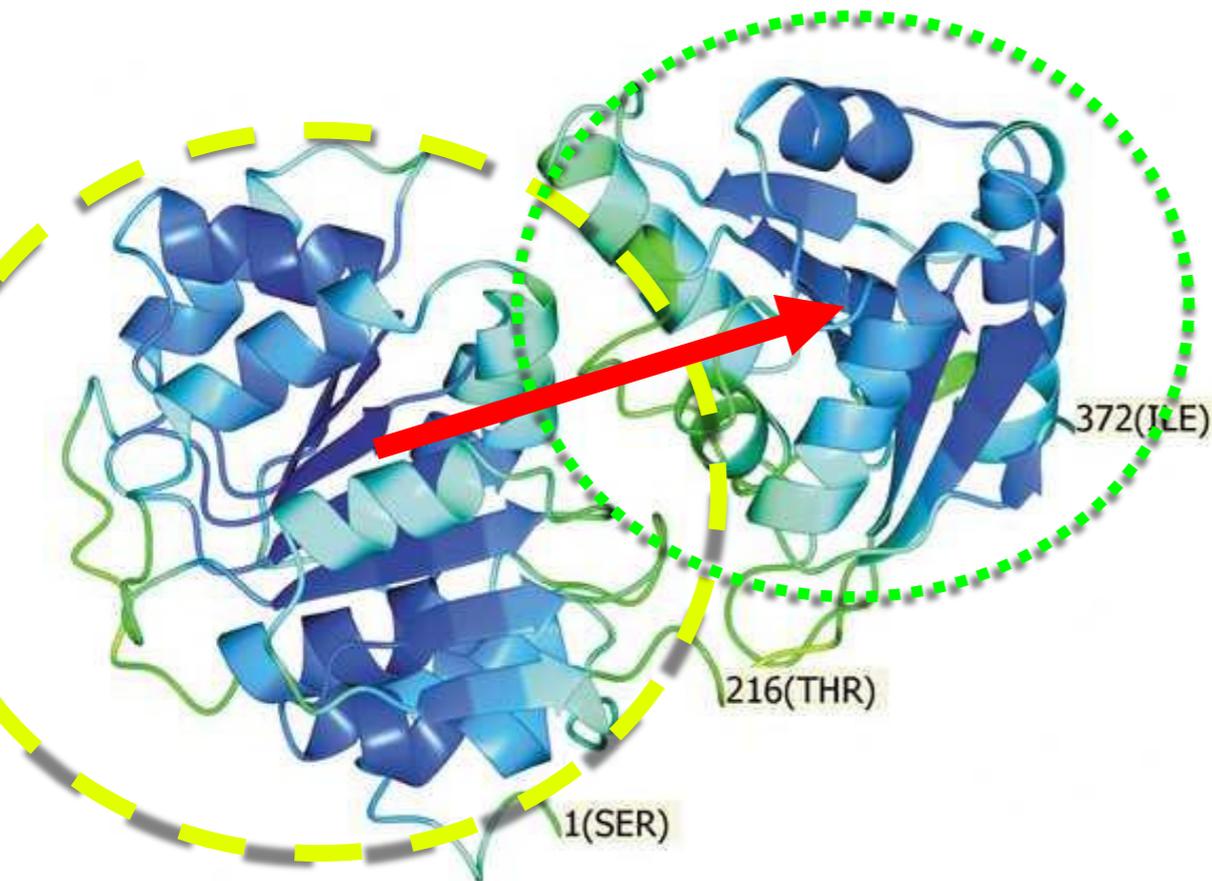
Blue: > 90

Green: 80 - 90

AlphaFold prediction for
RNA helicase
(PDB entry 6i5i)

AlphaFold confidence (pLDDT)	Median prediction error (Å)	Percentage with error over 2 Å
>90	0.6	10
80 - 90	1.1	22
70 - 80	1.5	33
<70	3.5	77

PAE matrix (Predicted aligned error)



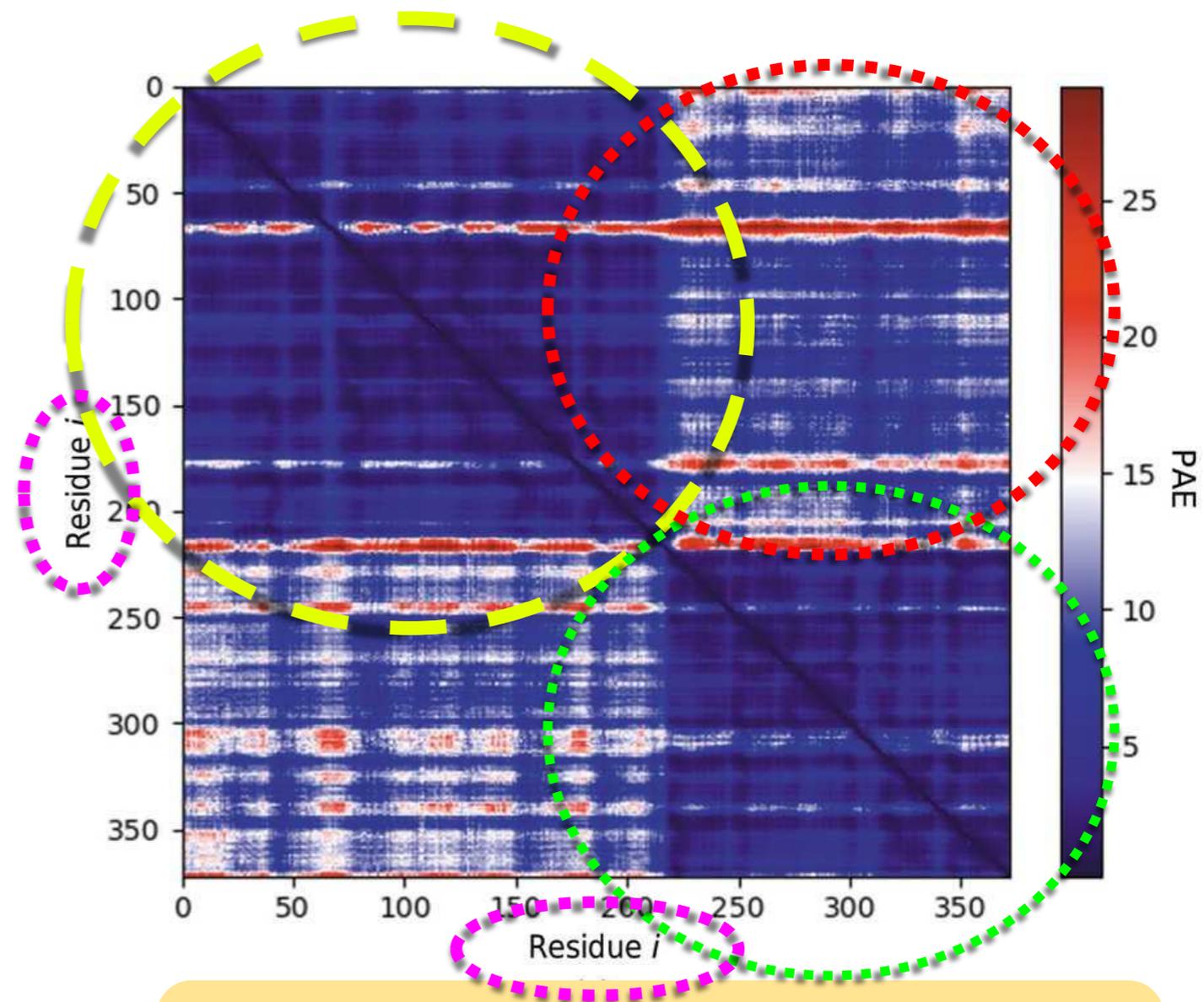
Confidence:

Blue: > 90

Green: 80 - 90

AlphaFold prediction for
RNA helicase
(PDB entry 6i5i)

PAE matrix identifies
accurately-predicted domains



Dark blue: uncertainty in
relative positions < 5 Å

Strategy for structure determination in the AlphaFold era

1. Predict your structure

*Design your experiment based on predicted models
(choose experimental approach, consider trimming at domain boundaries)*

2. Solve your structure

Cryo-EM or X-ray MR with trimmed predicted model, SAD

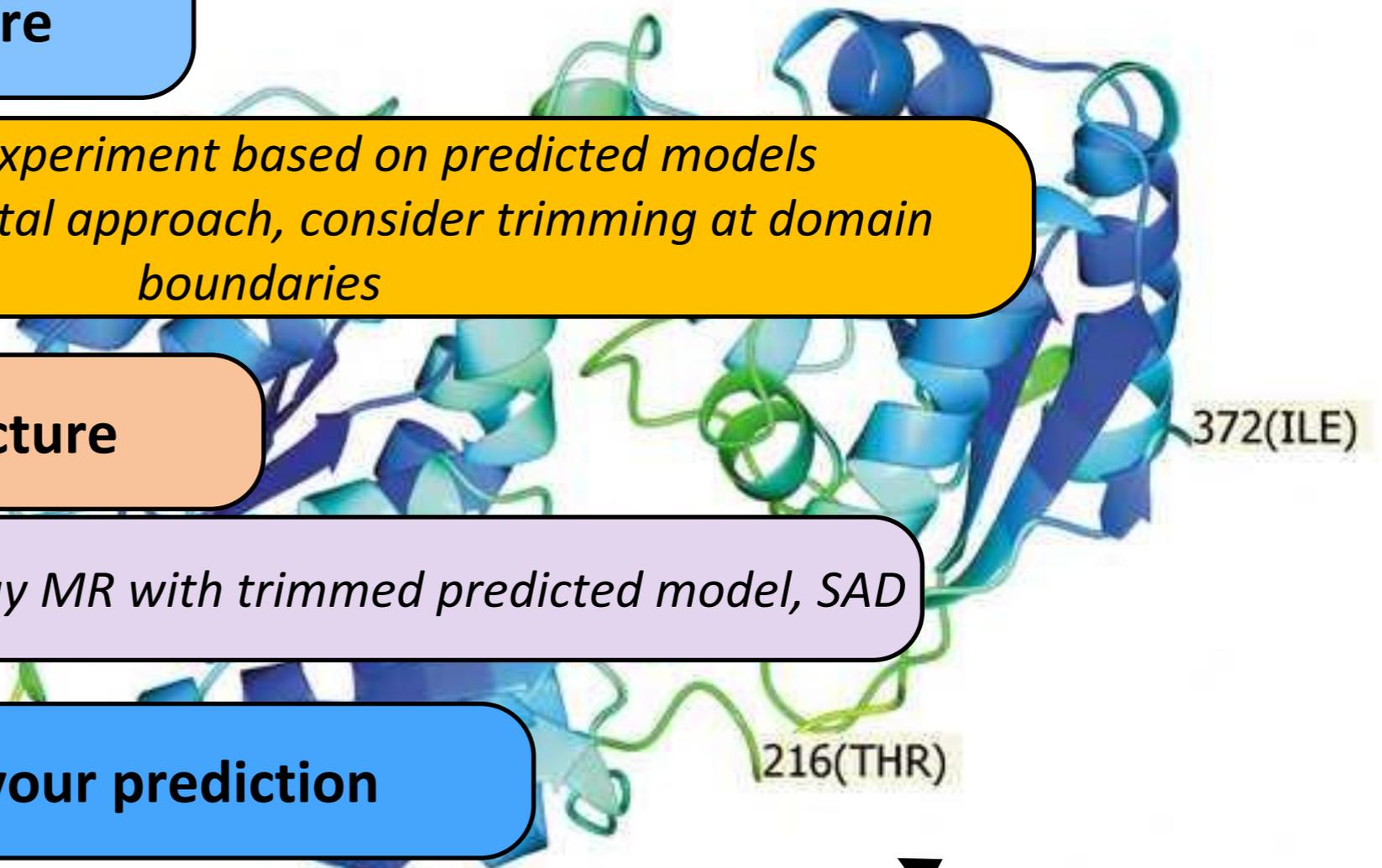
3. Update your prediction

*Run AlphaFold with your best model as a
template*

4. Improve your structure

*Use your new predictions as
hypotheses*

Iterate

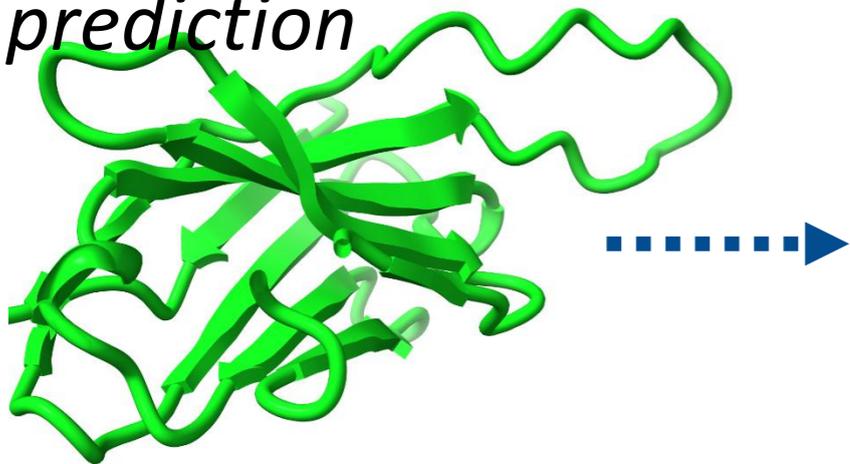


Using your best model as a template in AlphaFold prediction

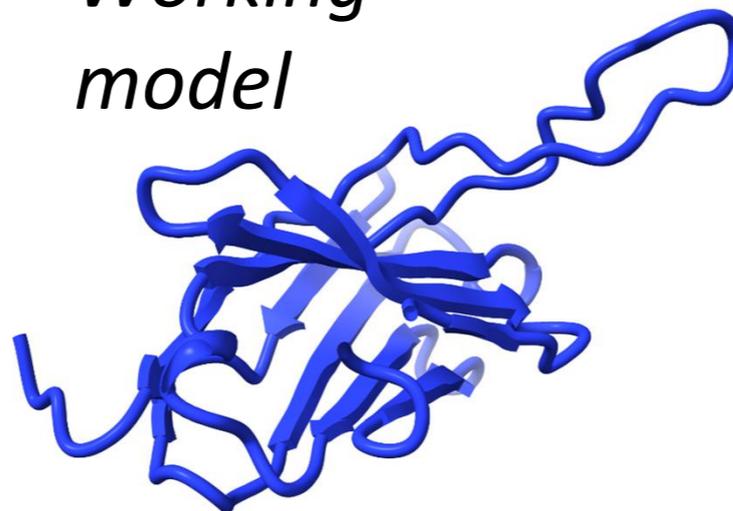
Why?

Because your new prediction might be better than your model ...and better than your original AlphaFold prediction

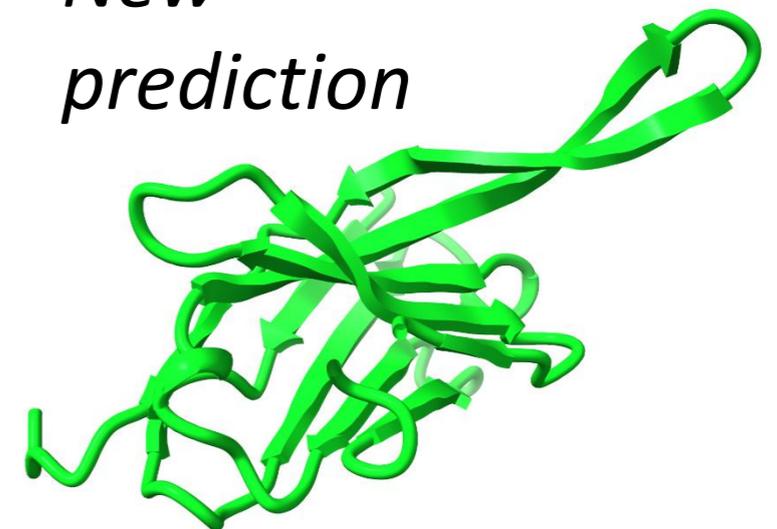
AlphaFold prediction



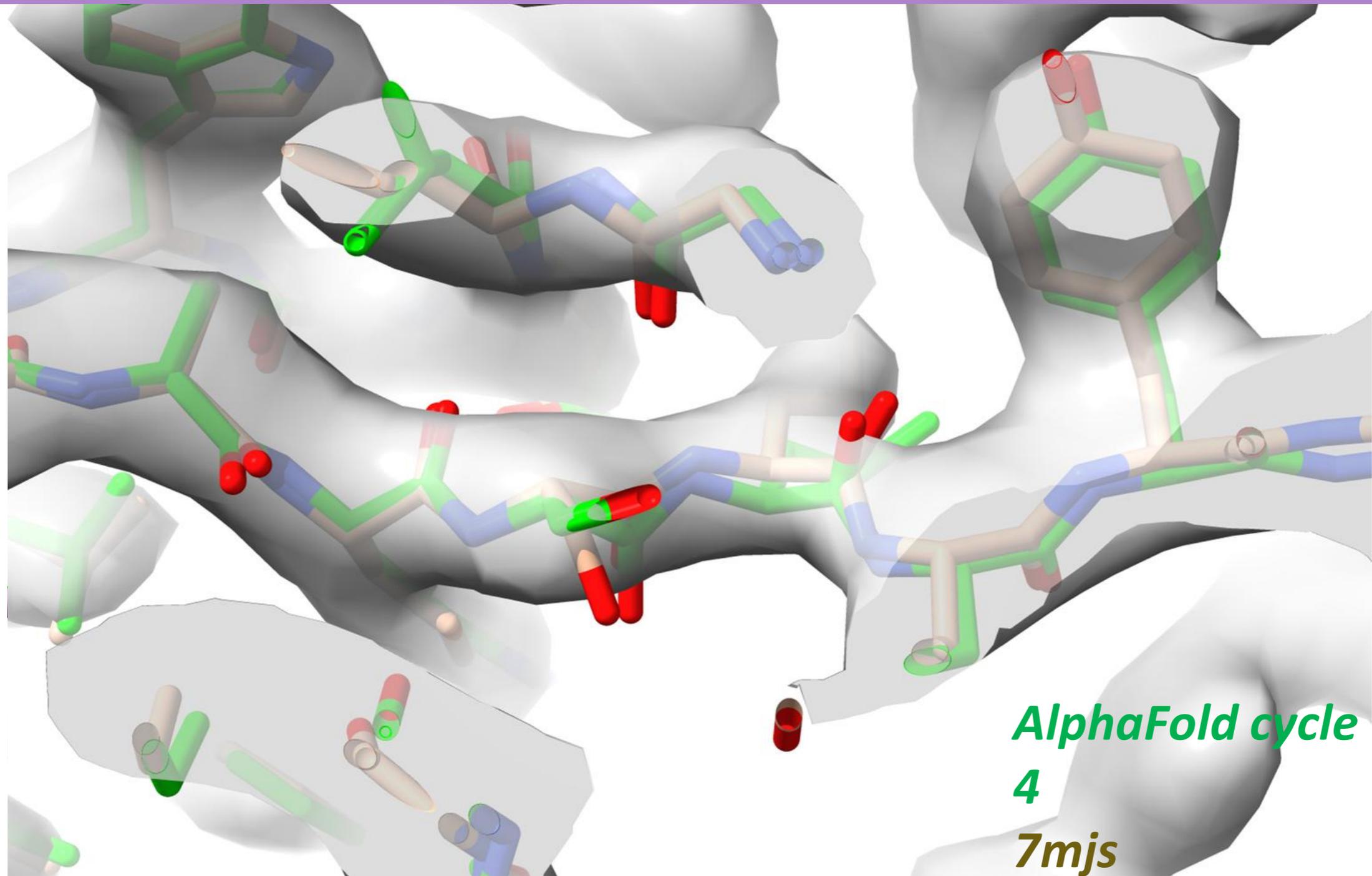
Working model



New prediction

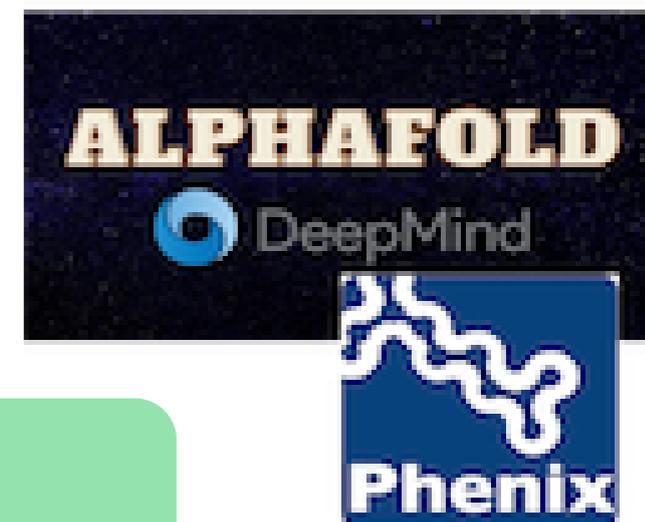


Improving AlphaFold prediction using partial models as templates (Cryo-EM)



Phenix AlphaFold prediction server

Available from the Phenix GUI



*Predicts structures of protein chains
(one at a time)*

Can use a template to guide the prediction

You do not need an MSA (multiple sequence alignment) if you supply a template

The template should not be an AlphaFold model

Many thanks for AlphaFold, ColabFold scripts, and the MMseqs2 server for MSAs

Process predicted model

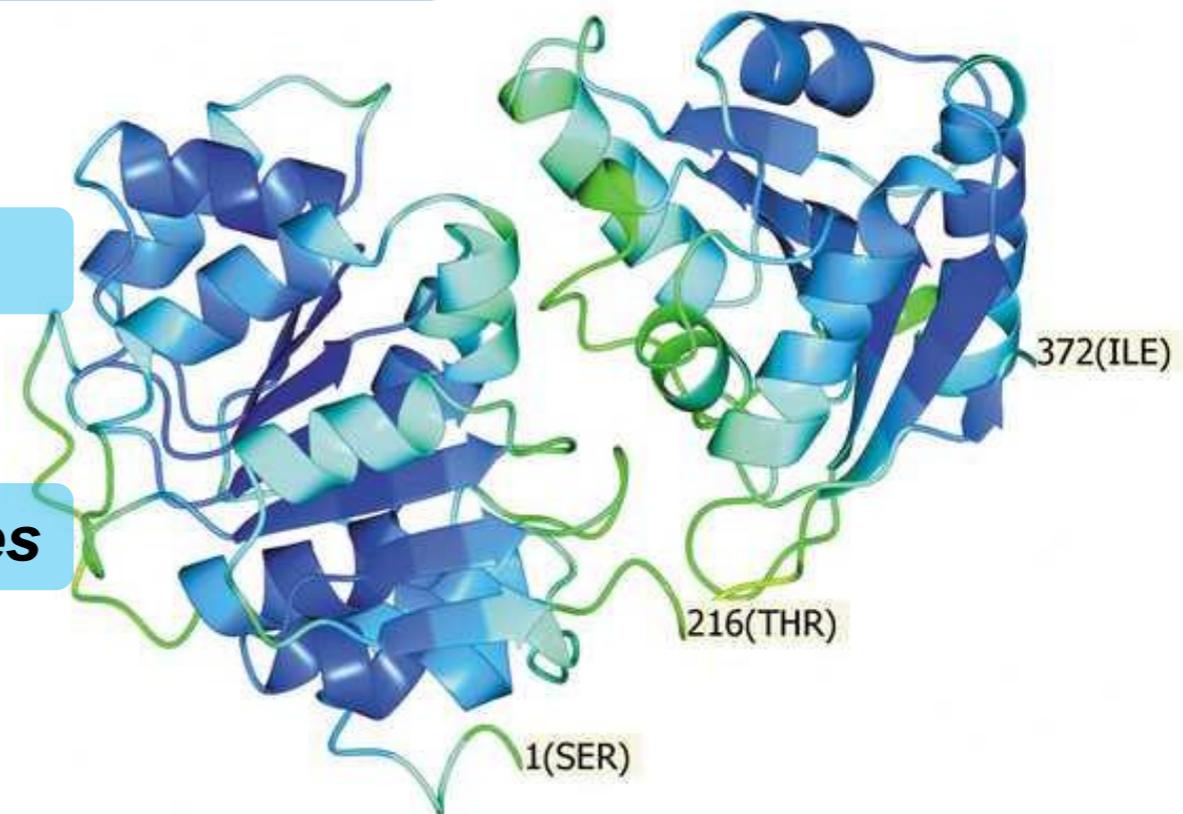
Convert *pLDDT* to *B-value*

Trim low-confidence parts of model

Identify high-confidence domains

Compact high-confidence regions

Groupings of residues with low PAE values



Phenix tools for structure determination with AlphaFold

PredictModel (Predict with AlphaFold)

*AlphaFold
models*

ProcessPredictedModel (Trim and identify domains)

ResolveCryoEM, LocalAnisoSharpen (map improvement)

EMPlacement, DockInMap (Docking of single, multiple chains)

Cryo-EM

DockAndRebuild (Morphing and rebuilding)

RealSpaceRefine (Refinement)

Phaser-MR (Molecular replacement)

AutoBuild (Density modification and rebuilding)

X-ray

Phenix.refine (Refinement)

PredictAndBuild (Prediction and structure determination)

*Full
automation*

Low-pLDDT AlphaFold predictions

- Most of the time, AlphaFold predictions are high-confidence and easy to interpret
- Most of the time, `phenix.process_predicted_model` is all you need
- So, let's talk about the other times . . .

When automation struggles, Use visualization

<https://alphafold.ebi.ac.uk/entry/Q5VSL9>

3D viewer

The screenshot displays the AlphaFold 3D viewer interface. At the top, the sequence of the protein AF-Q5VSL9-F1 is shown, with residue numbers 1, 11, 21, 31, 41, 51, 61, 71, 81, 91, 101, 111, 121, 131, 141, 151, 161, 171, 181, 191, and 199. The structure is visualized as a blue ribbon with a yellow tail. The interface includes a 'Structure Tools' panel on the right with sections for 'Structure', 'Quick Styles', 'Components', and 'Measurements'. The 'Structure' section shows the protein name and type. The 'Quick Styles' section has buttons for 'Default', 'Stylized', and 'Illustrative'. The 'Components' section shows the protein name and a 'Preset' button. The 'Measurements' section has an 'Add' button.

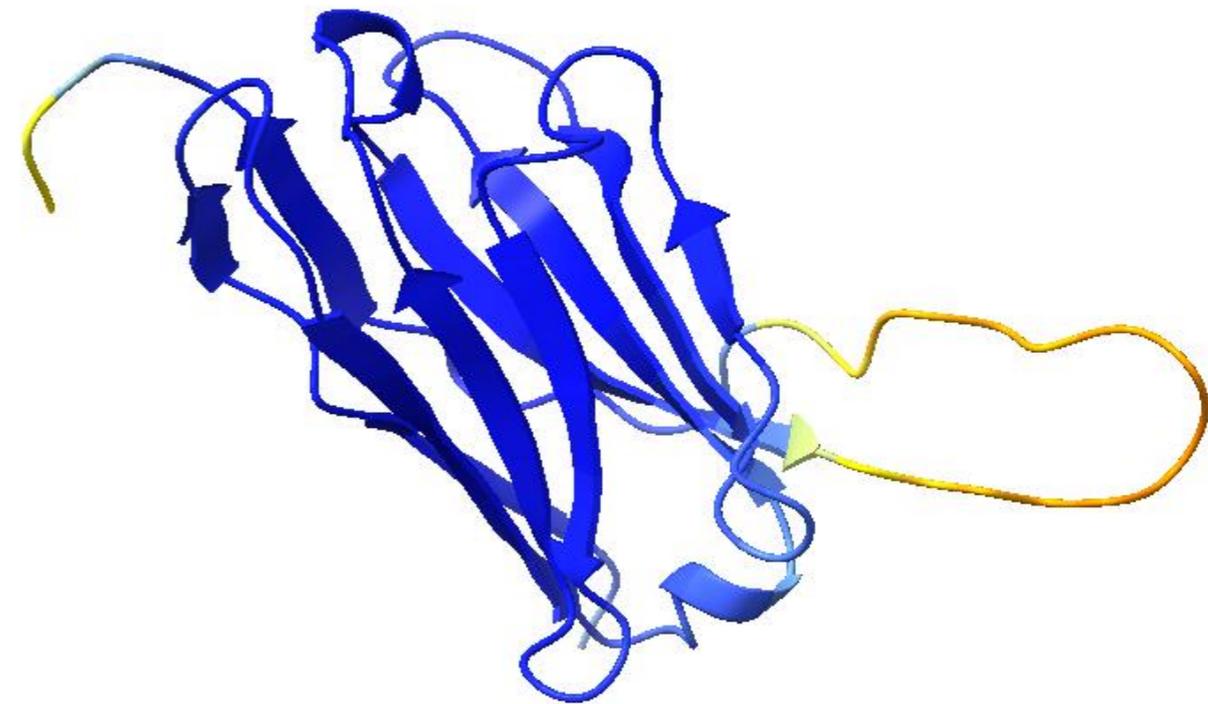
Model Confidence

- Very high (pLDDT > 90)
- High (90 > pLDDT > 70)
- Low (70 > pLDDT > 50)
- Very low (pLDDT < 50)

AlphaFold produces a per-residue model confidence score (pLDDT) between 0 and 100. Some regions below 50 pLDDT may be unstructured in isolation.

Mol* viewer at
<https://alphafold.ebi.ac.uk>

or
PDB



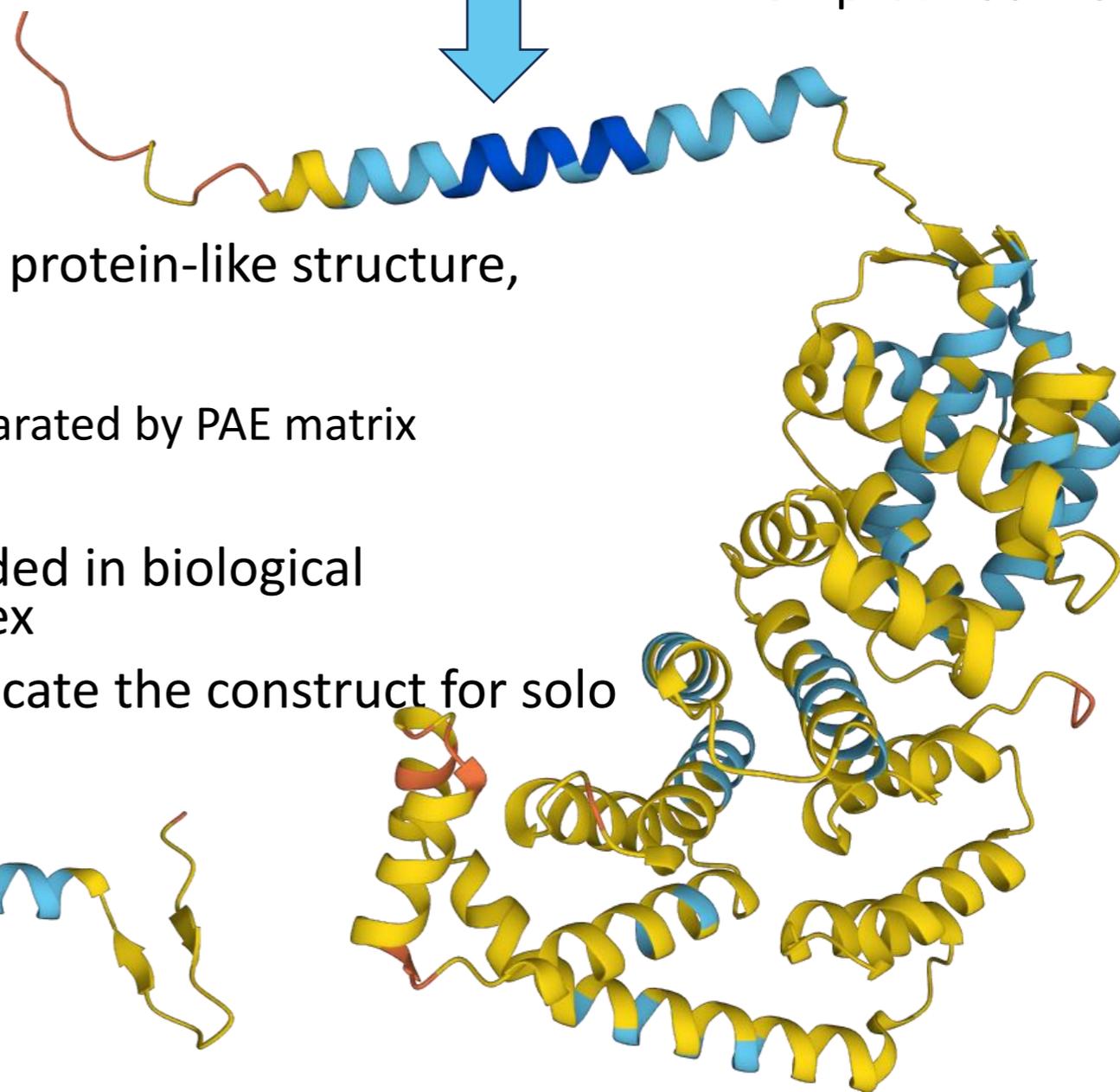
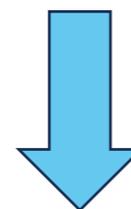
ChimeraX: “color bfactor palette alphafold”

Features to watch for

- High pLDDT
 - Unpacked helices
- Low pLDDT
 - Non-predictive “barbed wire”
 - Pseudostructure regions similar to secondary structure
 - Near-predictive packed regions

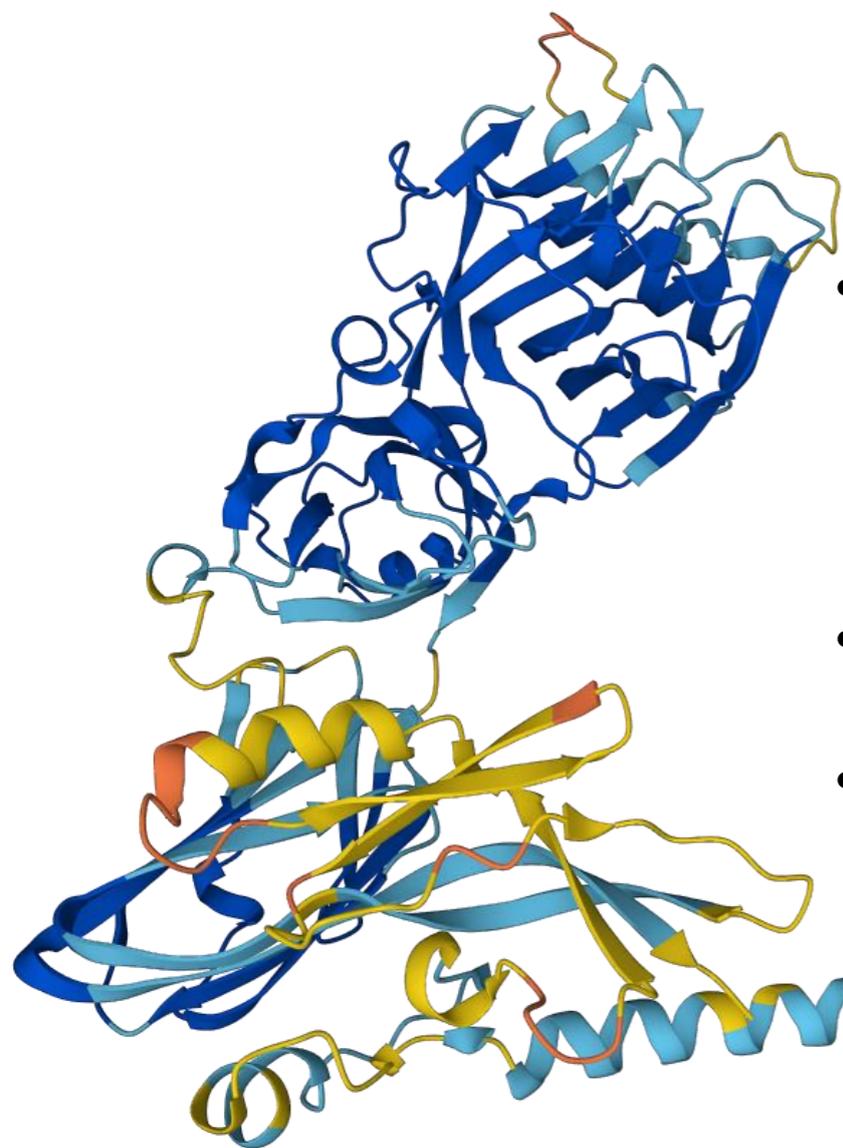
Unpacked high pLDDT

Homo sapiens
Uniprot **P60228**



- High-confidence, protein-like structure, touching nothing
 - Often helix
 - Often well-separated by PAE matrix

- Conditionally folded in biological multimer/complex
- May have to truncate the construct for solo crystallization



M. Jannaschii
Uniprot **Q58865**

AlphaFold predictions and confidence estimates

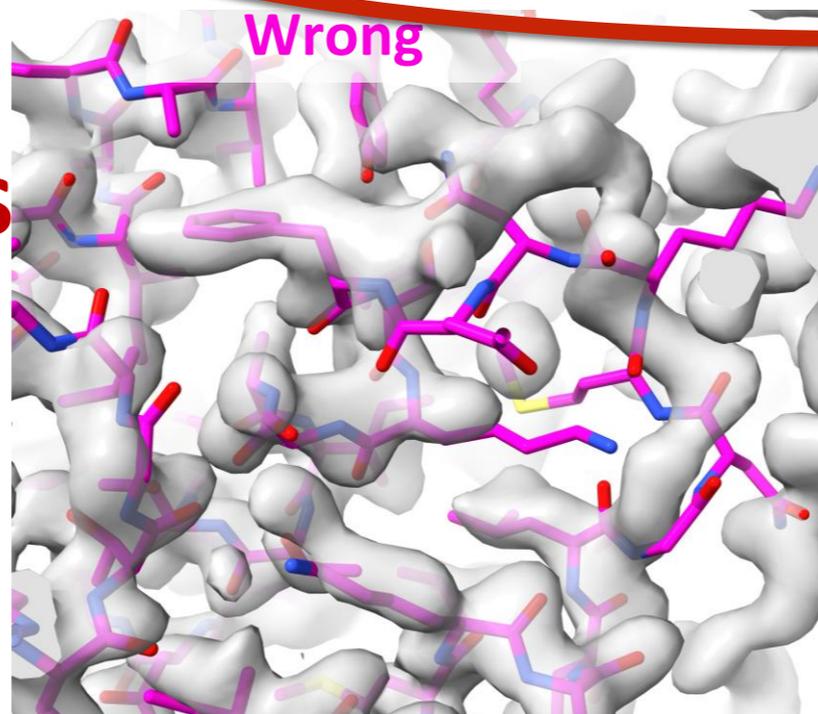
Residue-specific confidence (pLDDT) identifies where errors are more likely

AlphaFold confidence (pLDDT)	Median prediction error (Å)	Percentage with error over 2 Å
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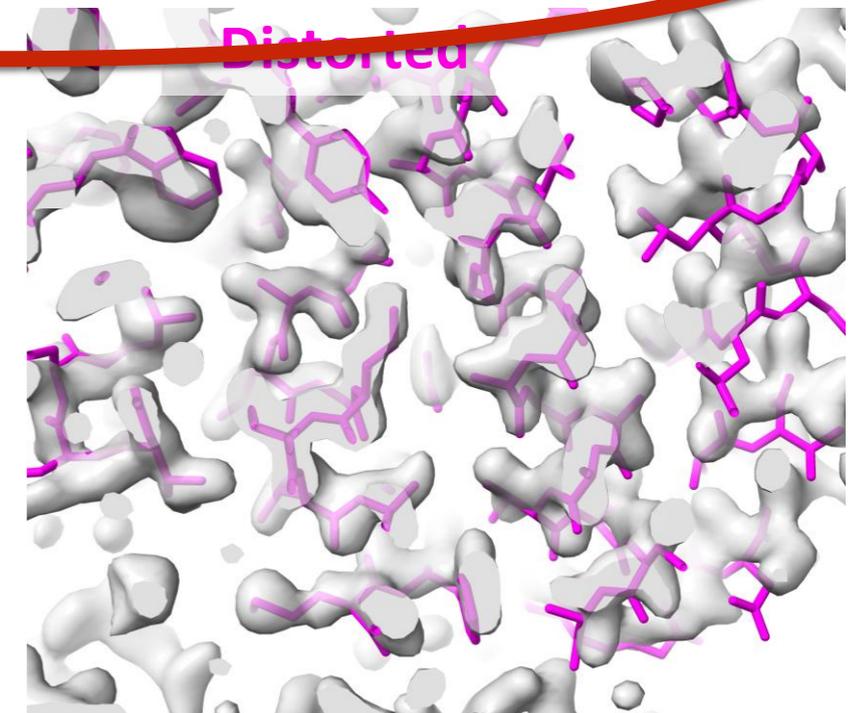
Awesome

The low-pLDDT regime contains multiple behaviors

Wrong



Distorted



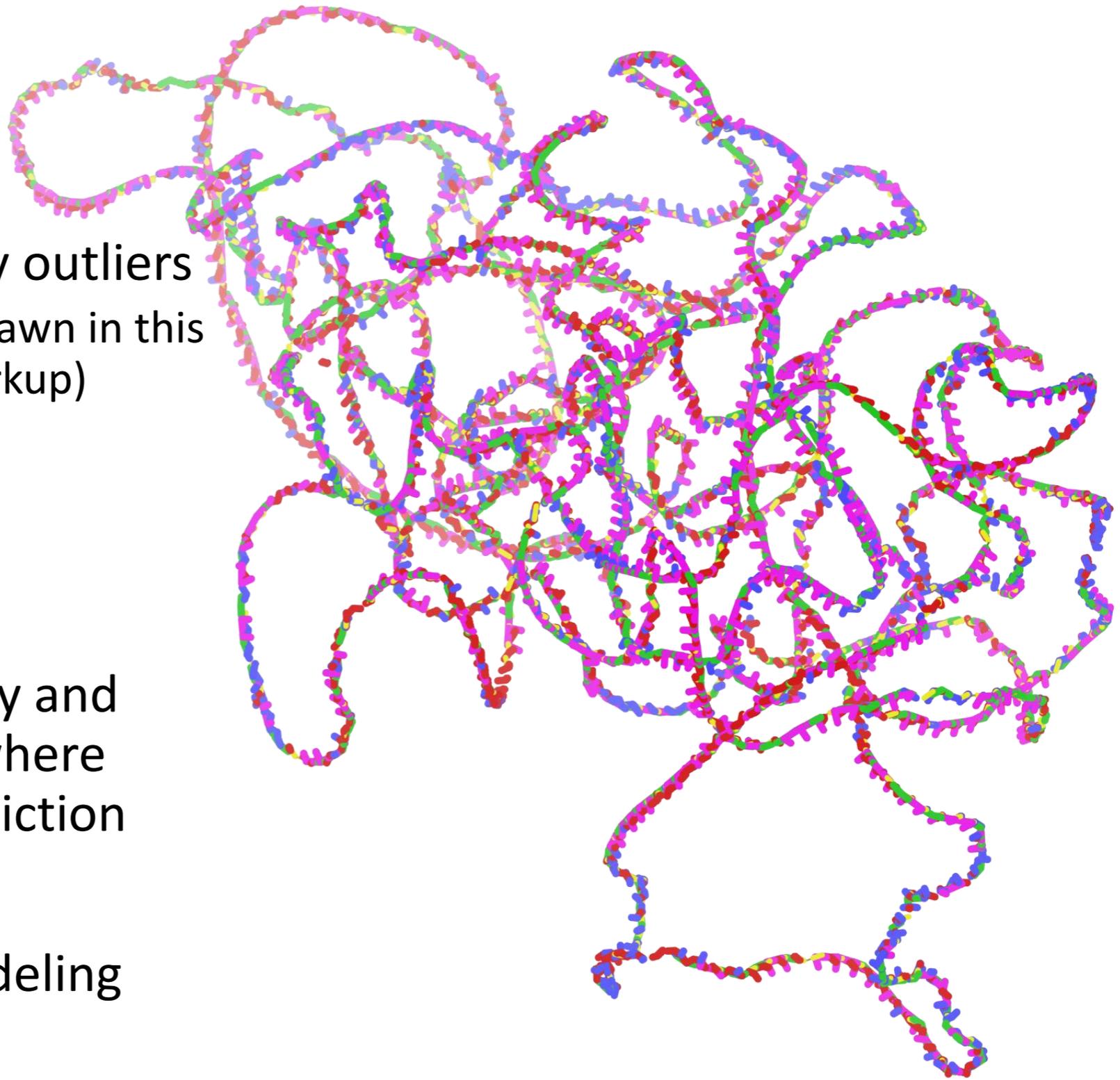
Low pLDDT - Barbed wire



Low-confidence AlphaFold predictions often have wide coils like concertina wire

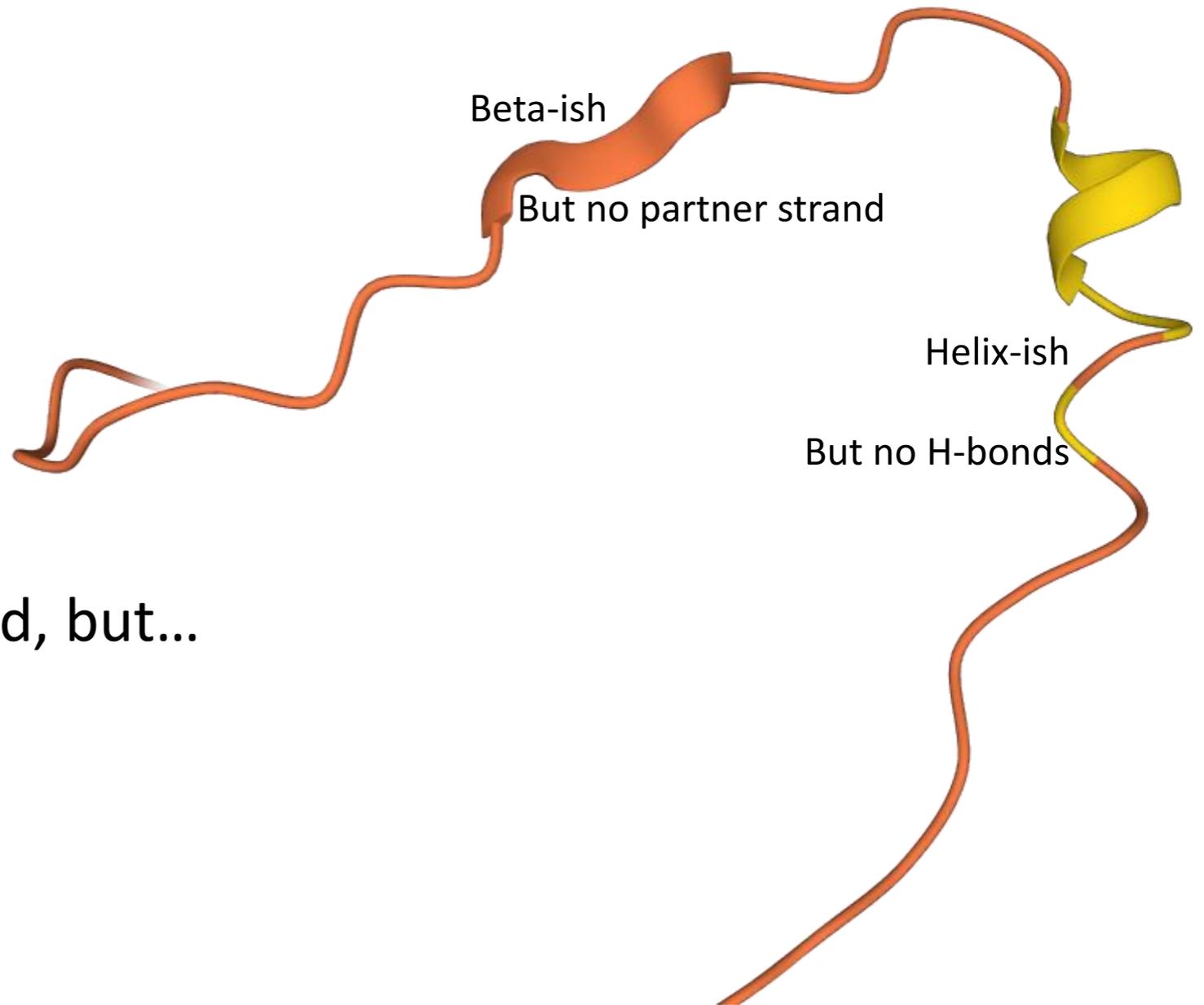
Barbed wire

- Extreme density of geometry outliers
 - (The protein is not actually drawn in this image, just the validation markup)
- This is a good thing!
- Along with pLDDT, this clearly and consistently marks regions where AlphaFold has made no prediction
- Different from “normal” modeling errors



Pseudostructure

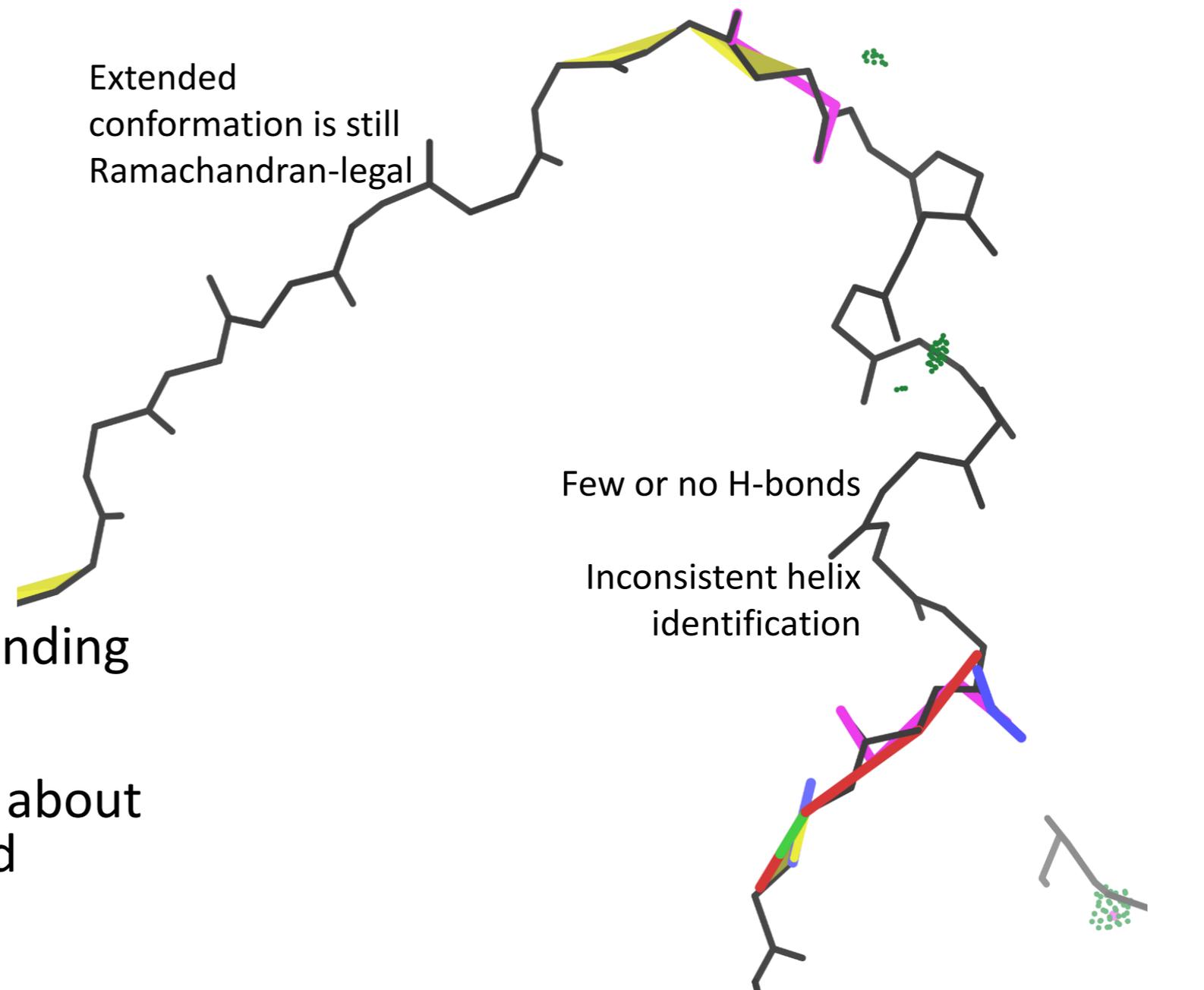
- Somewhat protein-like conformations
- Unpacked and unidealized, but...



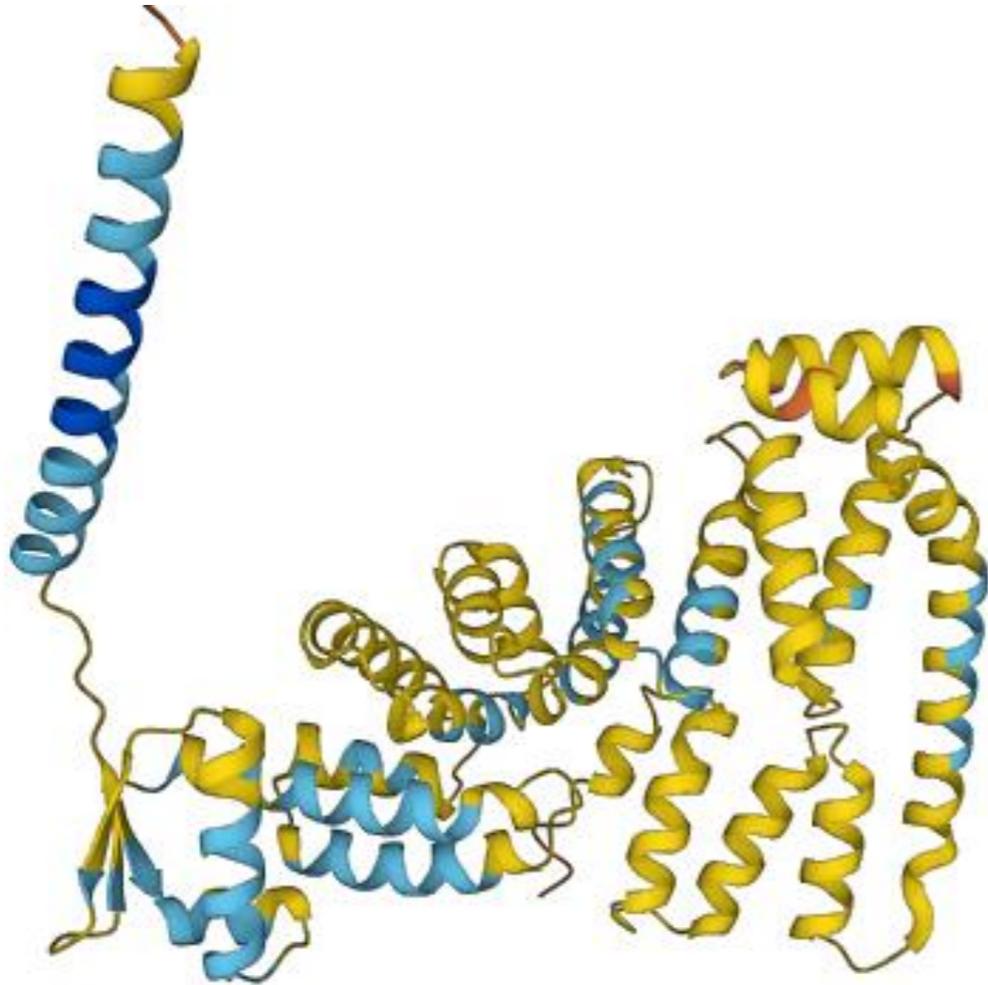
Zinc finger CCCH domain-containing protein 13
Residues 70-100
Homo sapiens
Uniprot **Q5T200**

Pseudostructure

- Lacks validation outliers!
- Also lacks good hydrogen bonding
- May have some information about disorder (Signal peptides and disorder-to-disorder binding regions)...
- ...but the *appearance* of structure is probably an “AI hallucination”



Near-predictive

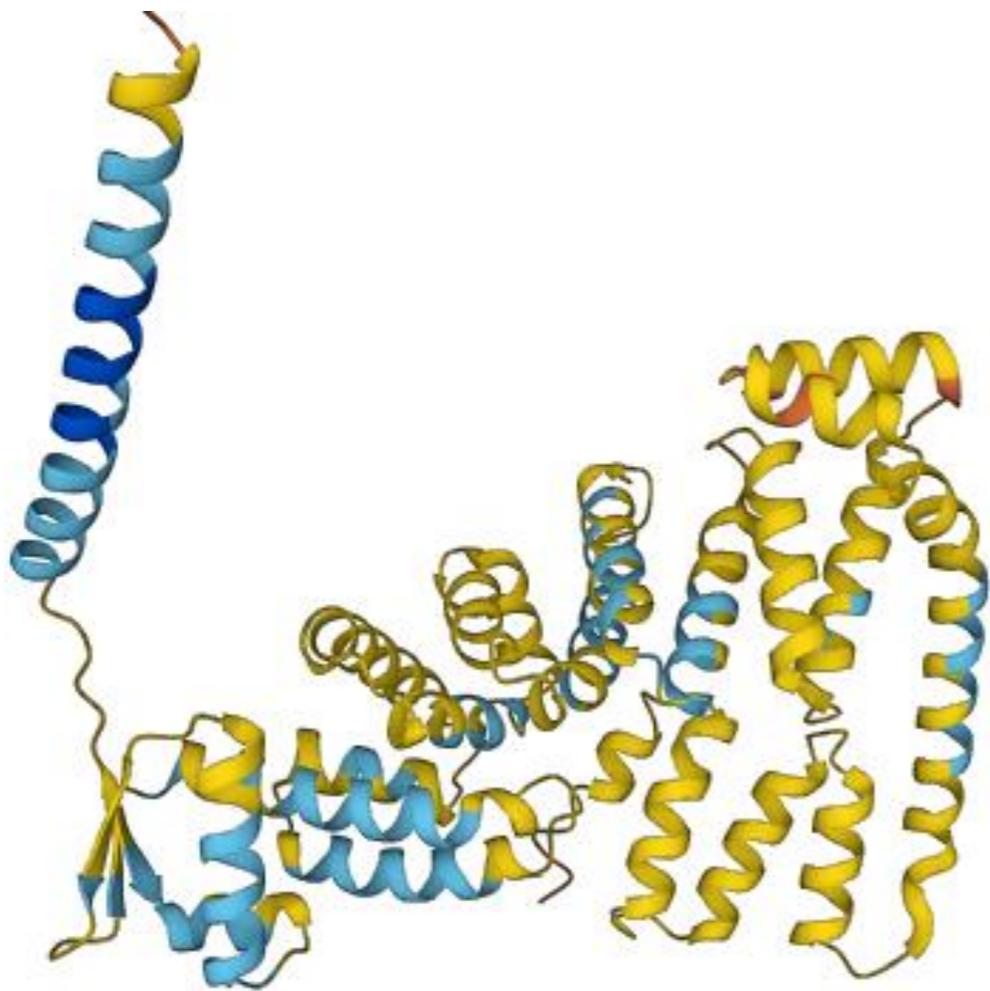


- Low pLDDT, but . . .
- Well-packed
- Protein-like fold
- Protein-like local geometry

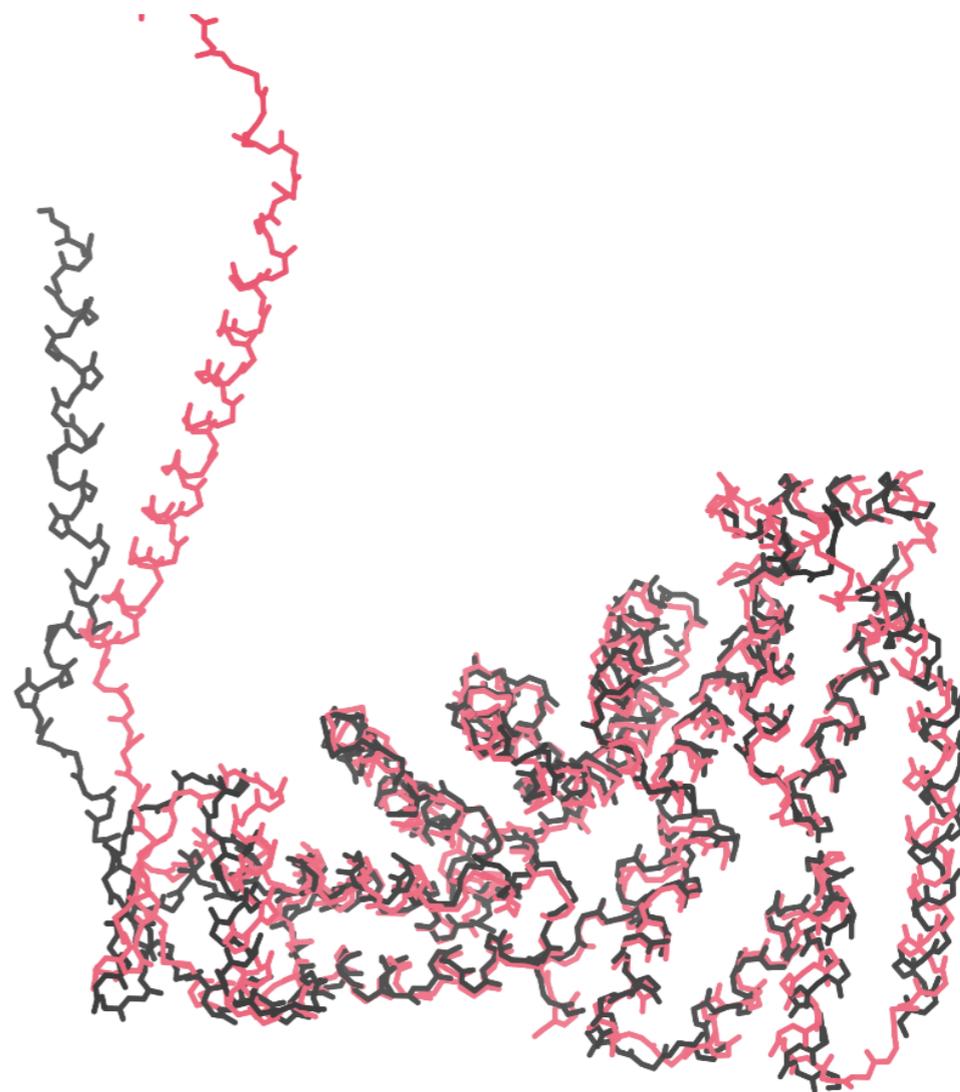
- Sometimes, these traits indicate a region with conditional folding, and sometimes . . .

Homo sapiens
Uniprot **P60228**

Near-predictive



Homo sapiens
Uniprot **P60228**

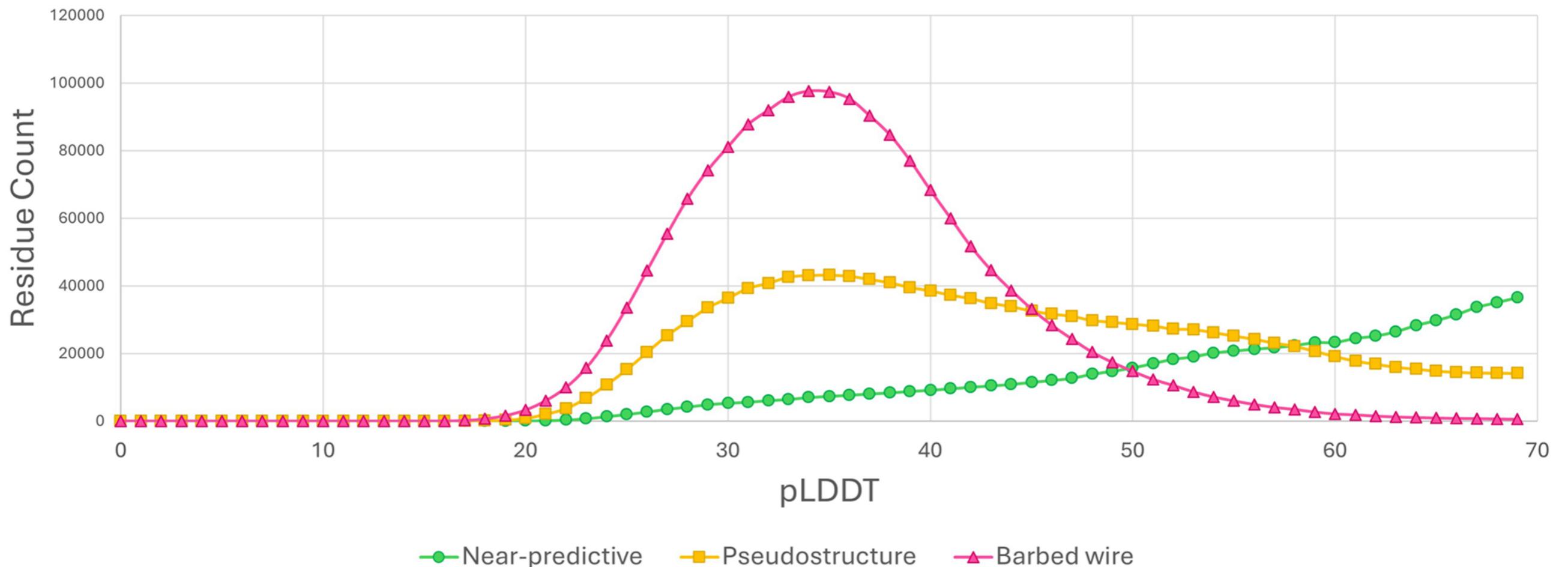


6zon.pdb, chain E

P60228 AlphaFold
prediction

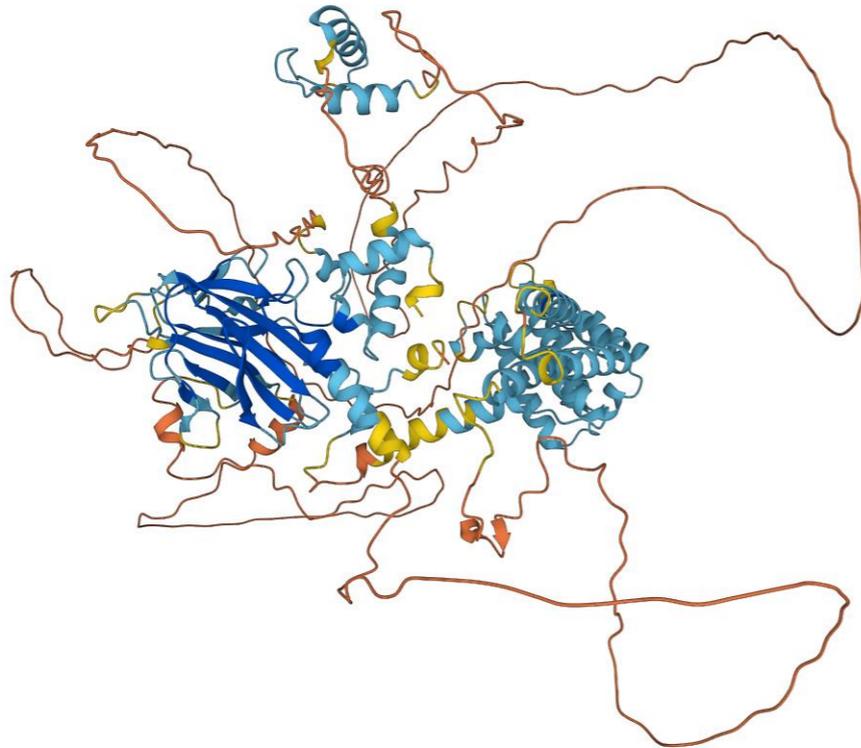
pLDDT comparison

Distribution of residue pLDDT for low-pLDDT prediction modes in AlphaFold2



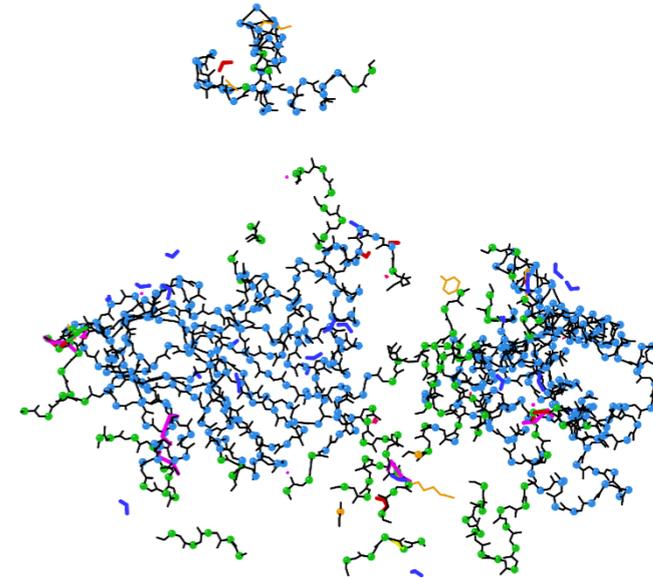
Low pLDDT contains multiple behaviors
Protein-like regions with pLDDT ~45-70 *may* still be usable!

Whole-model statistics may be misleading



Clashscore, all atoms:	0.54	
Clashscore is the number of serious steric overlaps (> 0.4 Å) per 1000 atoms.		
Poor rotamers	27	3.12%
Favored rotamers	791	91.55%
Ramachandran outliers	133	13.91%
Ramachandran favored	702	73.43%
Rama distribution Z-score	-3.50 ± 0.24	
MolProbity score [^]	1.87	
C β deviations >0.25Å	72	7.97%
Bad bonds:	0 / 7731	0.00%
Bad angles:	241 / 10452	2.31%
Cis Prolines:	3 / 28	10.71%
Cis nonProlines:	30 / 929	3.23%
Twisted Peptides:	152 / 957	15.88%
CaBLAM outliers	149	15.6%
CA Geometry outliers	144	15.09%
Tetrahedral geometry outliers	10	

Barbed wire present, validation says
“probably unusable”



Clashscore, all atoms:	0.54	
Clashscore is the number of serious steric overlaps (> 0.4 Å) per 1000 atoms.		
Poor rotamers	7	1.34%
Favored rotamers	509	97.32%
Ramachandran outliers	4	0.75%
Ramachandran favored	505	94.22%
Rama distribution Z-score	-0.75 ± 0.33	
MolProbity score [^]	1.17	
C β deviations >0.25Å	7	1.28%
Bad bonds:	0 / 4757	0.00%
Bad angles:	30 / 6407	0.47%
Cis Prolines:	0 / 18	0.00%
Cis nonProlines:	1 / 554	0.18%
Twisted Peptides:	6	1.2%
CaBLAM outliers	1	0.20%
CA Geometry outliers	0/707	

Barbed wire removed, validation says
“needs work”

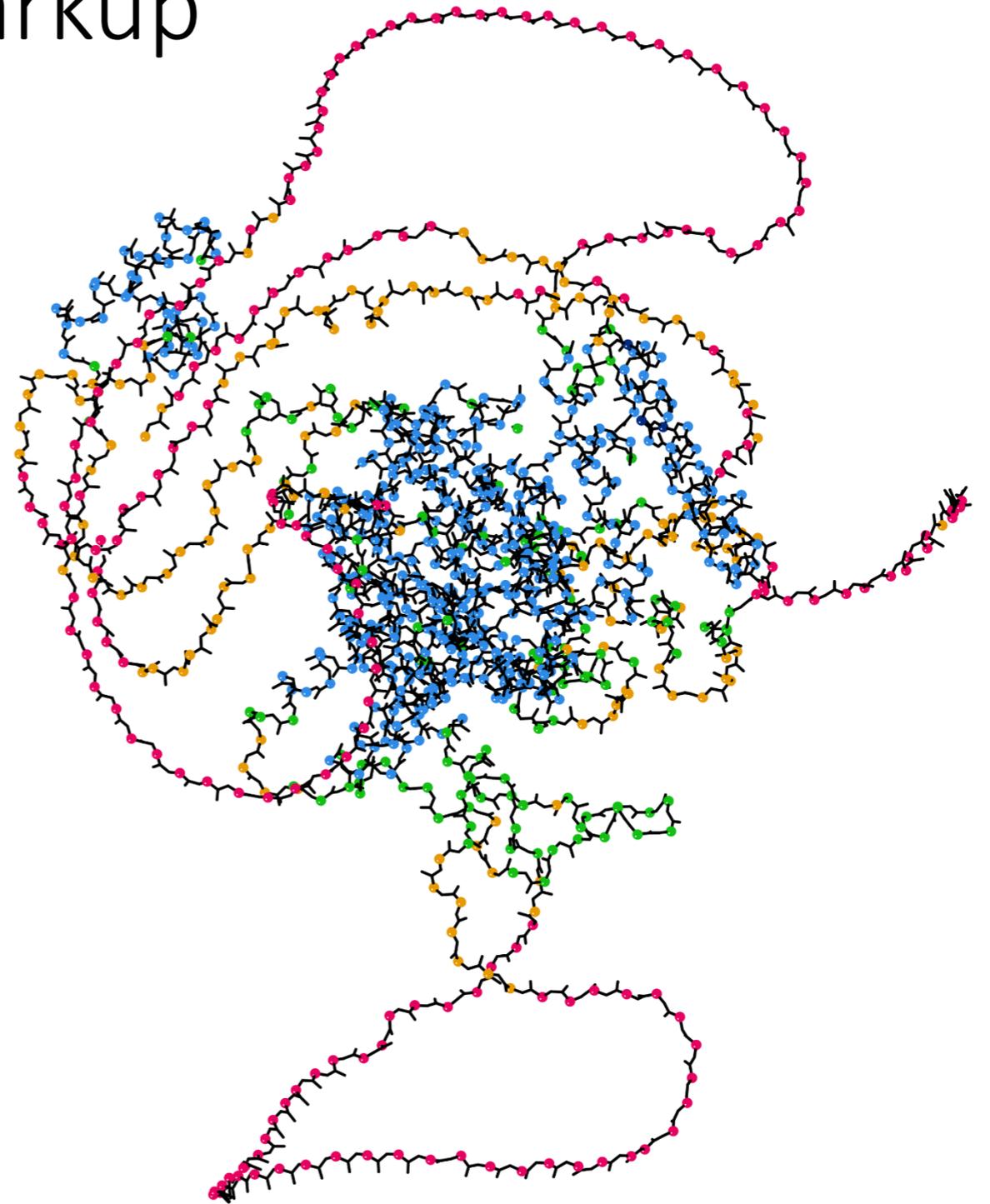
Low-pLDDT tool in Phenix

- Barbed wire analysis combines:
 - pLDDT score
 - Packing quality
 - Ignores contacts within secondary structure
 - Ignores sequence-local contacts
 - Density of barbed wire-like validation problems

- `phenix.barbed_wire_analysis`
- `phenix.barbed_wire_analysis output.type=kin`
 - Colored balls kinemage markup
- `phenix.barbed_wire_analysis output.type=selection_file`
 - PDB-format file of just the Predictive and Near-predictive parts of the input

Low-pLDDT kinemage markup

- Predictive (blue)
 - Unpacked high pLDDT (gray)
 - Near-predictive (green)
 - Pseudostructure (gold)
 - Barbed wire (hot pink)
-
- This markup only available in KiNG/kinemage format for now.



What about AlphaFold3?

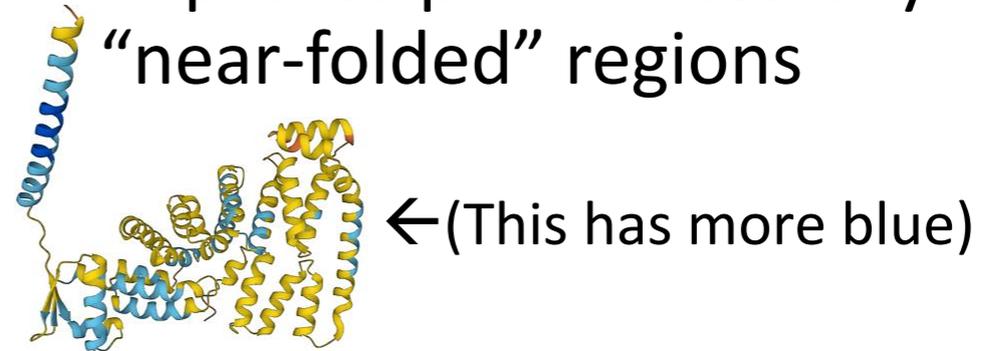
- This presentation concerns AlphaFold2

- AlphaFold3 has now been released

- Abramson, J., Adler, J., Dunger, J. *et al.* Accurate structure prediction of biomolecular interactions with AlphaFold 3. *Nature* **630**, 493–500 (2024). <https://doi.org/10.1038/s41586-024-07487-w>

- Offers centralized support for predicting ligands, multimers, modified residues, etc.

- Improves pLDDT accuracy for “near-folded” regions



- AF3 is not yet available in a form we can use for iterative prediction
- Stay tuned for developments



The Project



Lawrence Berkeley Laboratory

Paul Adams, Pavel Afonine,
Dorothee Liebschner, Nigel
Moriarty, Billy Poon,
Oleg Sobolev,
Christopher Schlicksup



Los Alamos National Laboratory New Mexico Consortium

Tom Terwilliger, Li-Wei Hung



University of Cambridge

Randy Read, Airlie McCoy,
Alisia Fadini



UTHealth

Matt Baker



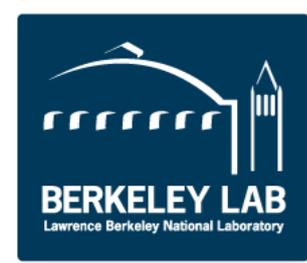
Duke University

Jane Richardson, Vincent
Chen, Michael Prisant,
Christopher Williams



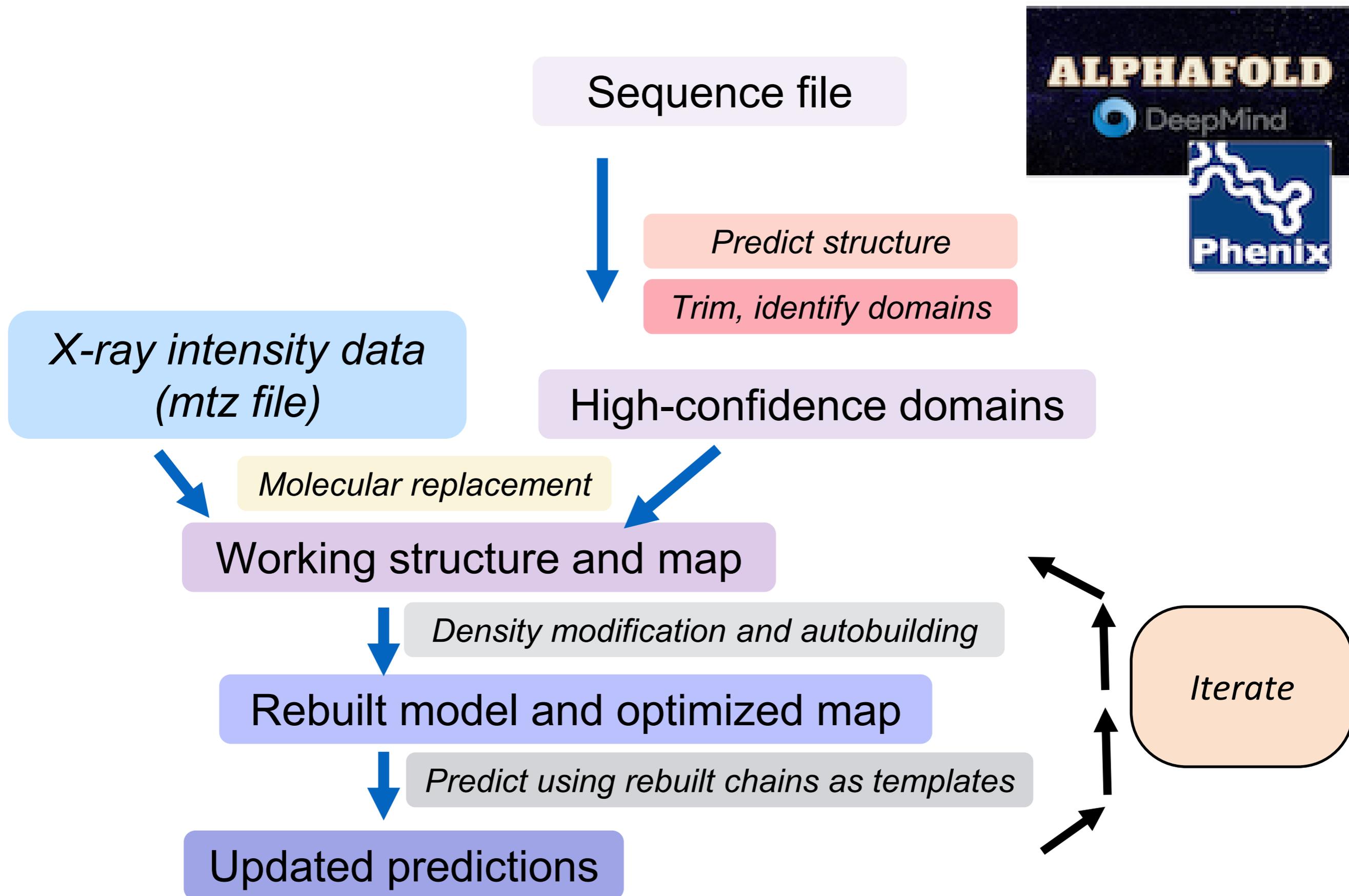
An NIH/NIGMS funded
Program Project

Liebschner D, *et al.*, Macromolecular structure determination using X-rays, neutrons and electrons: recent developments in *Phenix*. Acta Cryst. 2019 **D75**:861–877



Sample workflows

X-ray structure determination with AlphaFold



Cryo-EM structure determination with AlphaFold

Half-maps (optional processed map)

Density modification
or
Anisotropic sharpening

Optimized map



Dock domains in map

Docked domains



Morph full prediction onto domains and rebuild

Rebuilt model



Predict using rebuilt chains as templates

Updated predictions

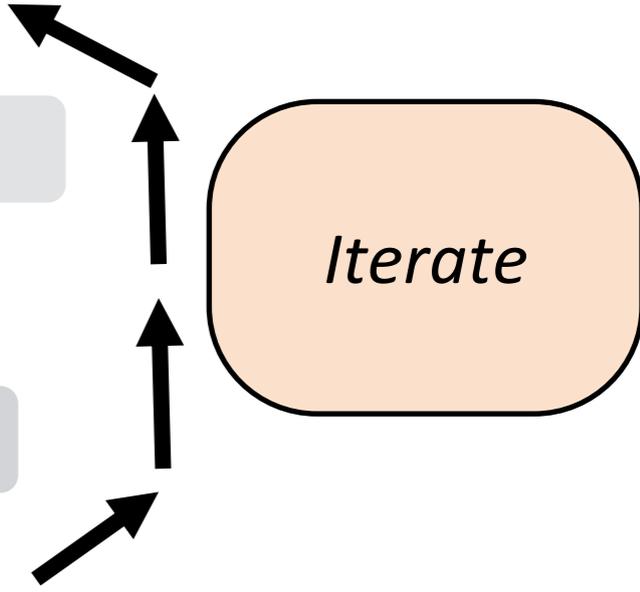
Sequence file



Predict structure

Trim, identify domains

High-confidence domains



Iterate

Input and output from structure determination with AlphaFold

Input

Experimental data (maps or X-ray data)

Contents of asymmetric unit (sequence file)

Output

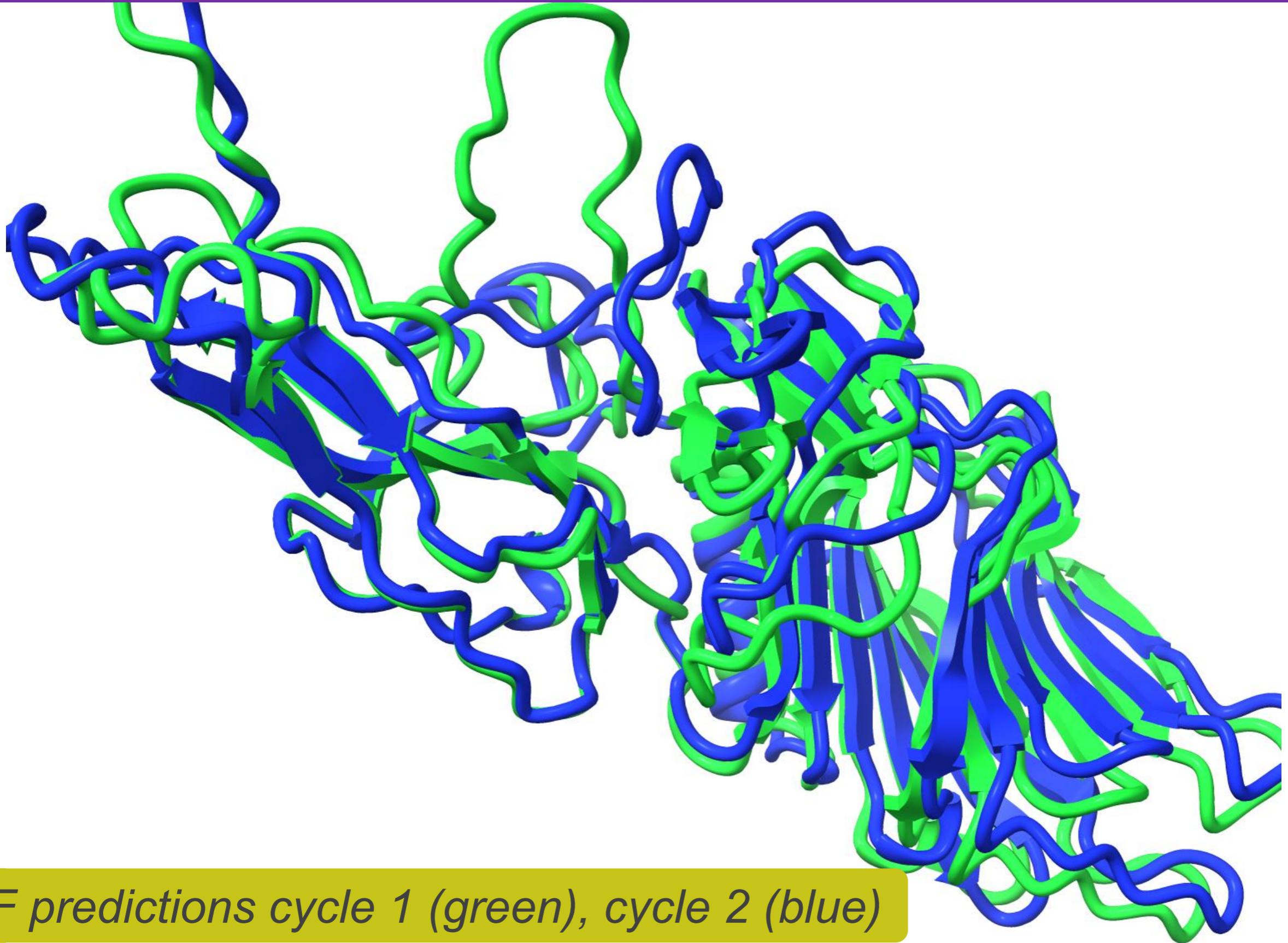
*Rebuilt model
Optimized map*

*Map and model ready
for next steps*

Docked predicted models

*Useful as high-quality
reference models*

*Improving AlphaFold prediction using partial models as templates
(X-ray crystallography)*



AF predictions cycle 1 (green), cycle 2 (blue)