



Computational Crystallography Initiative

Quick Facts About PHENIX, CCI APPS and Structure Refinement with phenix.refine

01 – February – 2008

PHYSICAL BIOSCIENCES DIVISION

The PHENIX project is a collaboration between several groups to develop highly integrated tools for automated crystallography:

- Los Alamos National Lab
Tom Terwilliger, Li-Wei Hung (SOLVE / RESOLVE)
Paul Langan, Marat Mustyakimov, Benno Schoenborn (Tools for Neutron crystallography) (separate funding, MNC)
- Cambridge University, UK
Randy Read, Airlie McCoy (PHASER)
- Duke University
Jane & David Richardson, Ian Davis (MolProbity)
- Lawrence Berkeley National Lab
Paul Adams, Pavel Afonine, Ralf Grosse-Kunstleve, Nigel Moriarty, Nicholas Sauter, Peter Zwart (CCI APPS)
- Texas A&M University
Tom Ioerger, Jim Sacchettini, Erik McKee (TEXTAL)

Tools for automated structure solution:

- **Automated analysis of data quality and integrity**
 - phenix.xtriage (detect twinning and more)
- **Rapid substructure determination** – Hybrid Substructure Search (HySS)
- **Scoring of substructure enantiomorphs** – Analysis of maps calculated with each hand
- **Phasing** – SOLVE, PHASER
- **Density modification** – Statistical density modification (RESOLVE)
- **Automated model building**
 - Pattern matching methods (RESOLVE or TEXTAL)
 - Building and refinement of multiple models
- **Structure refinement**
 - phenix.refine (complete and automated structure refinement at any resolution)
- **Ligand building and fitting** – eLBOW (cif files), AutoLigand
- **Comprehensive PDB file manipulations, models superposition**
 - phenix.pdbtools, phenix.superpose_pdbs
- **Structure validation**
 - MolProbity, Reduce (conformations and contacts, hydrogen atoms)

CCI APPS: Subset of PHENIX components developed by the Computational Crystallography Initiative (CCI) at LBNL:

Computational Crystallography Initiative (LBNL)

-Paul Adams, Ralf Grosse-Kunstleve, Pavel Afonine

-Nigel Moriarty, Nicholas Sauter, Peter Zwart



CCI APPS: Command-line tools only. “One click” installation on most of platforms (list of supported platforms: http://phenix-online.org/download/cci_apps/). Does not require installation of anything else. Can be obtained and installed separately from PHENIX package.

CCI APPS include:

phenix.refine – Highly-automated state-of-the-art structure refinement

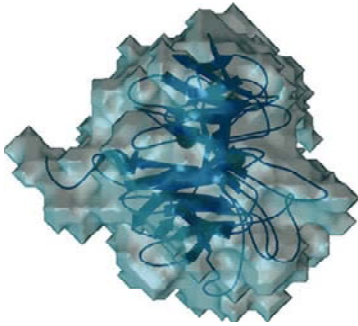
HySS – Highly-automated location of anomalous scatterers

eLBOW – Parameter and topology file builder for unknown ligands

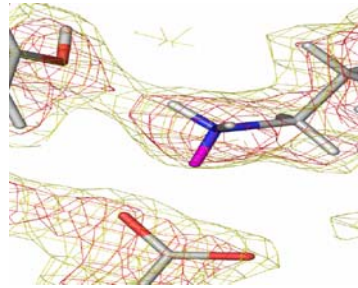
Xtrriage – Comprehensive analysis of experimental data

Below we mostly focus on phenix.refine. For information about other components please refer to separate documentation: <http://phenix-online.org/>

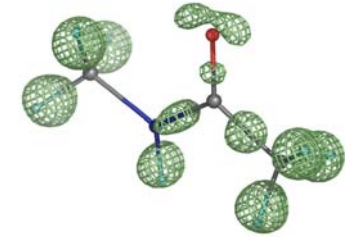
Low



Middle and High



Subatomic



- Group ADP refinement
- Rigid body refinement
- Torsion Angle dynamics

- Restrained refinement (xyz, ADP: isotropic, anisotropic, mixed)
- Automatic water picking

- Bond density model
- Unrestrained refinement
- FFT or direct
- Explicit hydrogens

- Automatic NCS restraints
- Simulated Annealing
- Occupancies (individual, group, constrains for alternative conformation)
- TLS refinement
- Use hydrogens at any resolution
- Refinement with twinned data
- X-ray, Neutron, joint X-ray + Neutron refinement

PDB model,
Any data format
(CNS, Shelx, MTZ, ...)



Input data and model processing

Refinement strategy selection

Bulk-solvent, Anisotropic scaling, Twinning
parameters refinement

Ordered solvent (add / remove)

Target weights calculation

Coordinate refinement
(rigid body, individual) (minimization or Simulated
Annealing)

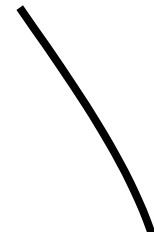
ADP refinement
(TLS, group, individual iso / aniso)

Occupancy refinement (individual, group)

Output: Refined model, various maps, structure
factors, complete statistics, ready for deposition PDB
file



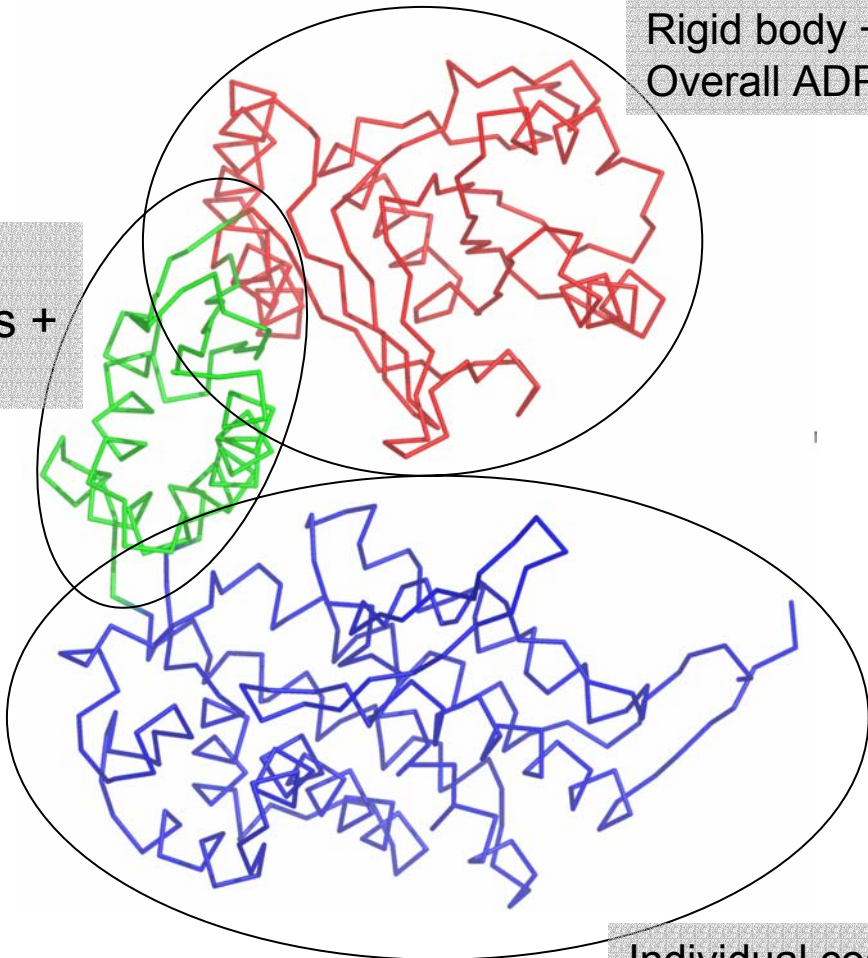
Files for
COOT, O,
PyMol



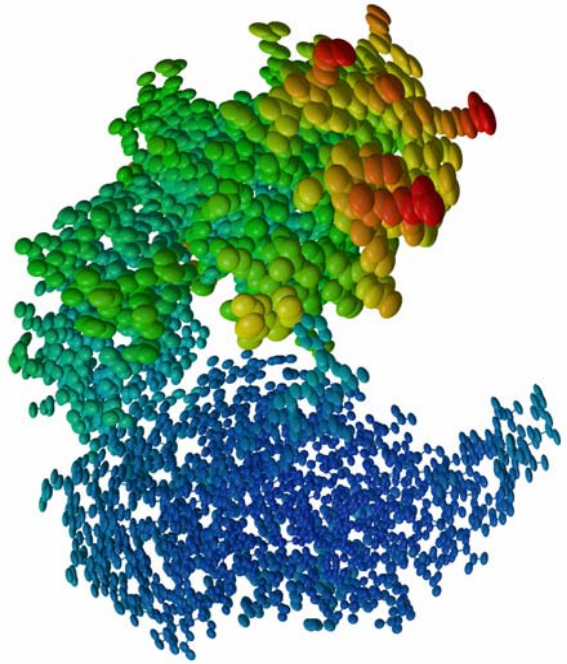
Repeated
several times

Individual coordinates + TLS

Rigid body + Overall ADP



Individual coordinates + TLS + restrained isotropic ADP



- + Automatic water picking
- + Simulated Annealing
- + Add and use hydrogens

Designed to be very easy to use:

Refinement of individual coordinates and B-factors:

```
% phenix.refine model.pdb data.hkl
```

Same as above plus Simulated Annealing (SA) and water picking:

```
% phenix.refine model.pdb data.hkl simulated_annealing=true ordered_solvent=true
```

Rigid-body + SA + grouped B-factors refinement:

```
% phenix.refine model.pdb data.hkl strategy=rigid_body+group_adp simulated_annealing=true
```

TLS + SA + individual coordinates and B-factors refinement:

```
% phenix.refine model.pdb data.hkl strategy=tls+individual_sites+individual_adp \
simulated_annealing=true
```

Refinement of individual coordinates and B-factors using neutron data:

```
% phenix.refine model.pdb data.hkl scattering_dictionary=neutron
```

To see all parameters (more than 200):

```
% phenix.refine model.pdb data.hkl --show_defaults=all
```

See the documentation for more examples: <http://www.phenix-online.org/>

Running phenix.refine with a parameter file:

```
% phenix.refine model.pdb data.hkl parameters_file
```

where the `parameter_file` contains following lines:

```
refinement.main {  
  high_resolution = 2.0  
  low_resolution = 15.0  
  simulated_annealing = True  
  ordered_solvent = True  
  number_of_macro_cycles = 5  
}  
refinement.refine.adp {  
  tls = chain A  
  tls = chain B  
}
```

This is equivalent to typing from the command line:

```
% phenix.refine model.pdb data.hkl xray_data.high_resolution=2 xray_data.low_resolution=15  
simulated_annealing=true ordered_solvent=True adp.tls="chain A" adp.tls="chain B"  
main.number_of_macro_cycles=5
```

Shortcuts are allowed, as long as they are unambiguous:

```
% phenix.refine model.pdb data.hkl xray_data.high_res=2 xray_data.low_res=15  
simulated_annealing=true ordered_s=True adp.tls="chain A" adp.tls="chain B"  
main.number_of_m=5
```

When running:

```
% phenix.refine model.pdb data.hkl
```

each item in `model.pdb` is matched against the CCP4 Monomer Library to extract the topology and parameters and to automatically build corresponding restraints.

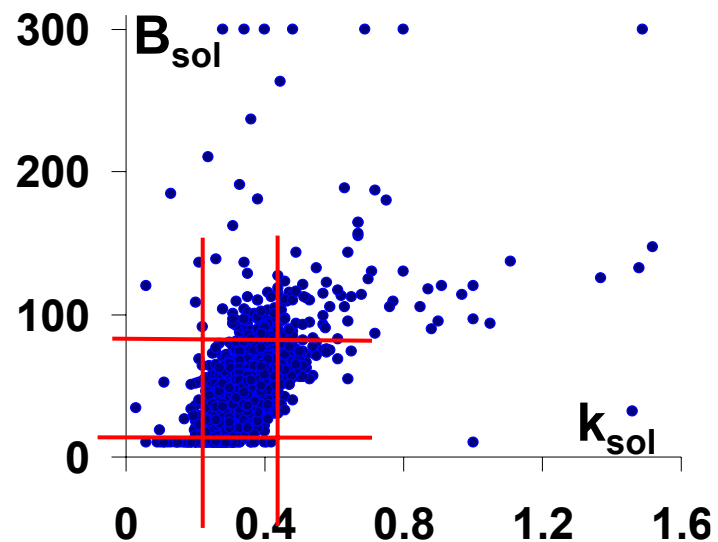
If `model.pdb` contains an item not available in CCP4 Monomer Library, e.g. a novel ligand, use `eLBOW` to generate the required topology and parameter definitions for refinement:

```
% phenix.elbow model.pdb --residue=LIG
```

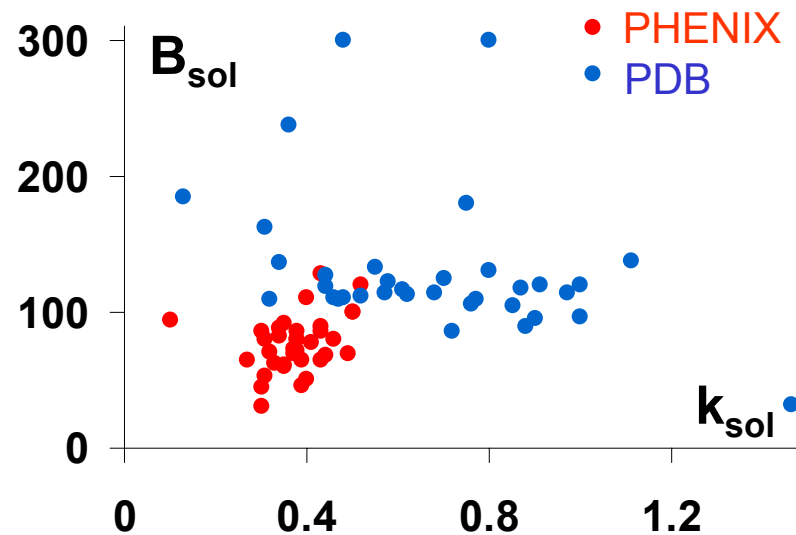
This will produce the file `LIG.cif` which can be used for refinement:

```
% phenix.refine model.pdb data.hkl LIG.cif
```

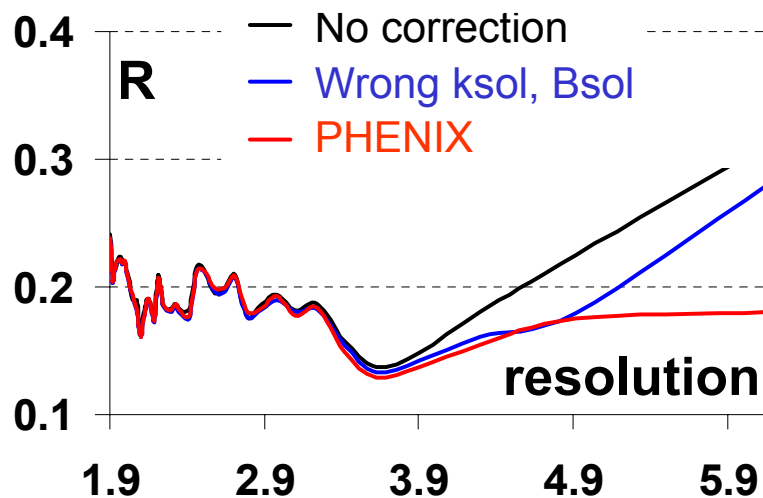
PDB survey



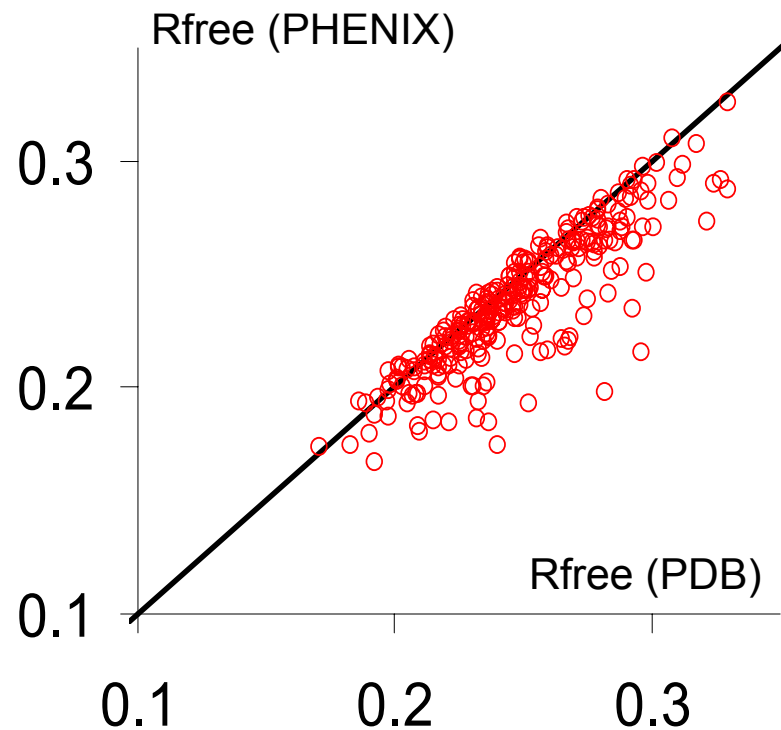
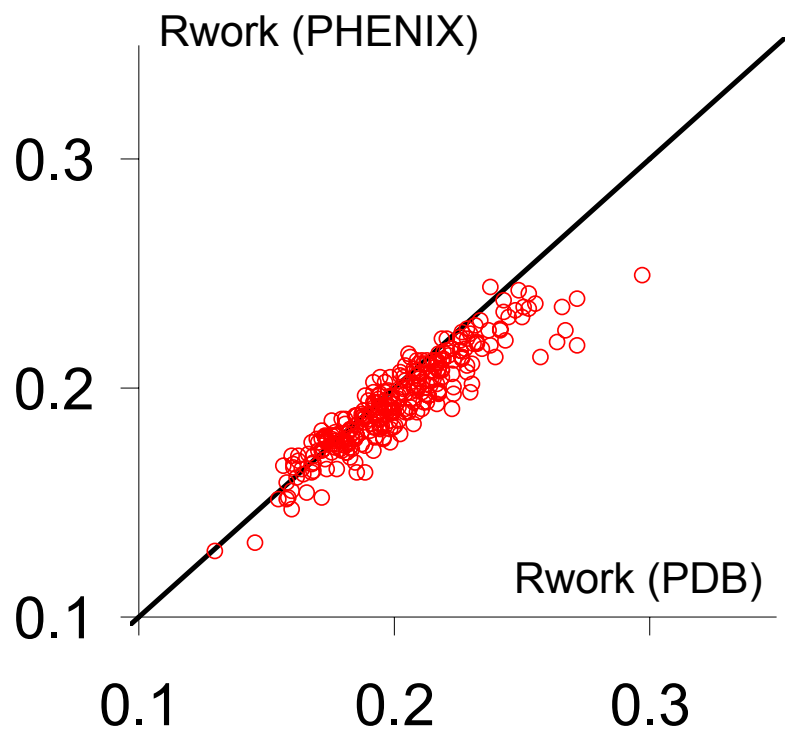
Fixing outliers with PHENIX



Effect on R-factors



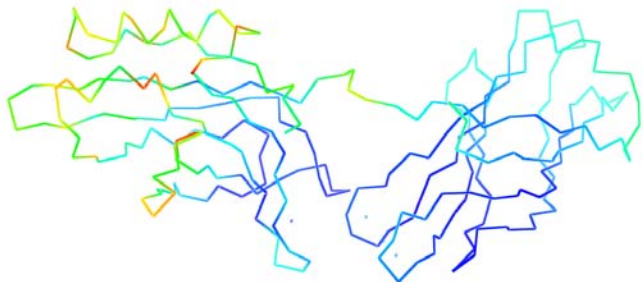
- Re-refinement of 355 PDB models (with TLS parameters)
- In many cases the phenix.refine results are significantly better
- Never crashed or got “unstable”



Restraints target for individual isotropic ADP refinement

$$E_{\text{ADP}} = \sum_{i=1}^{N_{\text{atoms}}} \left[\sum_{j=1}^{M_{\text{atoms}}} \frac{1}{r_{ij}^{\text{distance_power}}} \frac{(U_i - U_j)^2}{\left(\frac{U_i + U_j}{2}\right)^{\text{average_power}}} \right]_{\text{sphereR}}$$

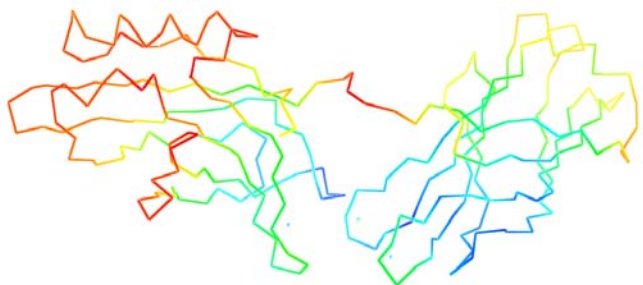
Synaptotagmin refinement at 3.2 Å



CNS
R-free = 34%
R = 29%



PHENIX – Isotropic restrained ADP
R-free = 27.7%
R = 24.6%



PHENIX – TLS + Isotropic ADP
R-free = 24.4%
R = 20.7%

- Currently LS X-ray refinement target:

$$E = \sum_j w_j (F_{\text{obs}} - G_{\text{model}})^2$$

$$G_{\text{model}} = \sqrt{(1 - \alpha)kF_{\text{model,h}}^2 + \alpha kF_{\text{model,Rh}}^2}$$

$$\mathbf{F}_{\text{model}} = \mathbf{F}_{\text{atoms}} + f(k_{\text{sol}}, B_{\text{sol}})\mathbf{F}_{\text{sol}}$$

- Running twin refinement is easy and includes two steps:

- run phenix.xtriage to get twin operator (twin law):

```
% phenix.xtriage data.mtz
```

- run phenix.refine:

```
% phenix.refine model.pdb data.mtz twin_law="-h-k,k,-l"
```

- Taking twinning into account makes difference:

PDB code: 1l2h

	R/R-free (%)
Deposited (SHELXL):	15.4 / 19.3
PHENIX (no twinning):	24.9 / 27.4
PHENIX (twin refinement):	15.3 / 19.2

PDB code: 1p7g

	R/R-free (%)
Deposited (CNS):	16.1 / 21.7
PHENIX (twin refinement):	16.2 / 21.6
PHENIX (twin + TLS refinement):	14.6 / 19.6

Macromolecular Neutron Crystallography Consortium (MNC)



Los Alamos National Laboratory
Paul Langan, Marat Mustyakimov, Benno Schoenborn



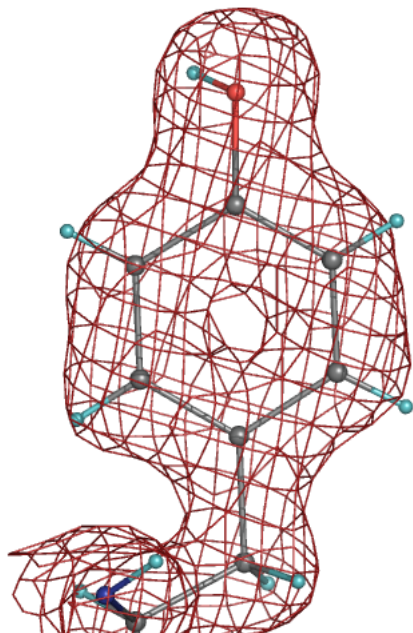
Lawrence Berkeley National Lab (LBNL)
Paul Adams, Pavel Afonine

<http://mnc.lanl.gov/>

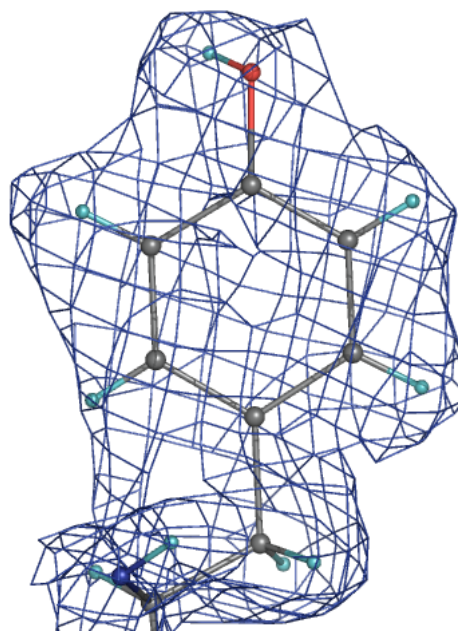
- Different techniques – different information

2mFo-DFc maps (Aldose Reductase)

X-ray (1.8 Å)



Neutron (2.2 Å)

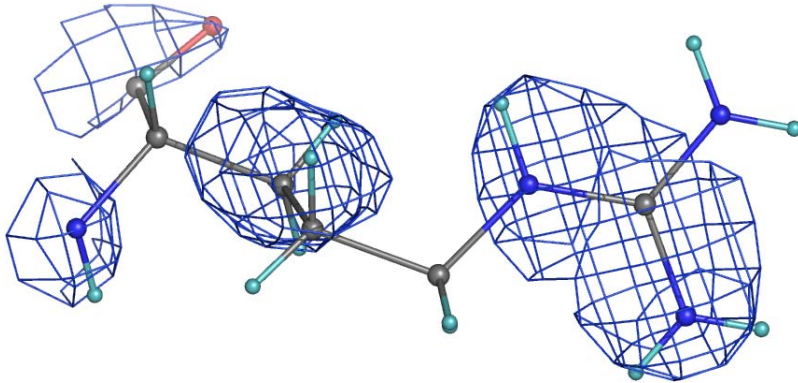


- Target used for joint X-ray + neutron refinement:

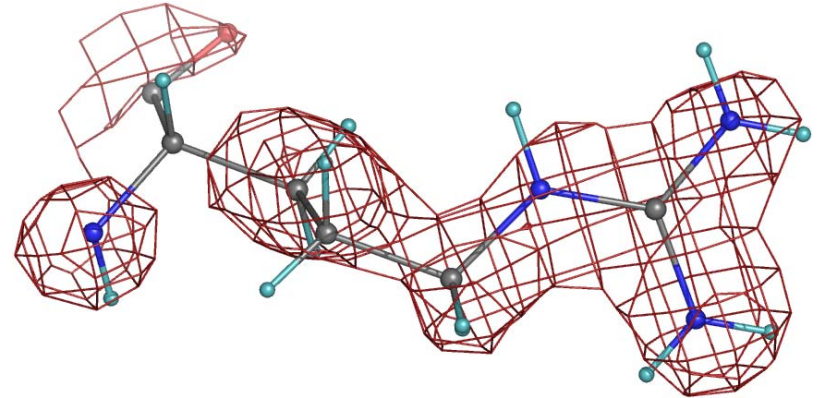
$$\text{Target}_{\text{JOINT}} = E_{\text{XRAY}} * w_{\text{XC}} + E_{\text{NEUTRON}} * w_{\text{NC}} * w_{\text{XN}} + E_{\text{GEOM}}$$

2mFo-DFc, neutron data, 2σ , 2.2 Å resolution

Refinement (neutron data only)



Refinement (X-ray and neutron data)



- Neutron maps are improved after joint refinement comparing to refinement with neutron data only

Running joint X-ray + neutron refinement

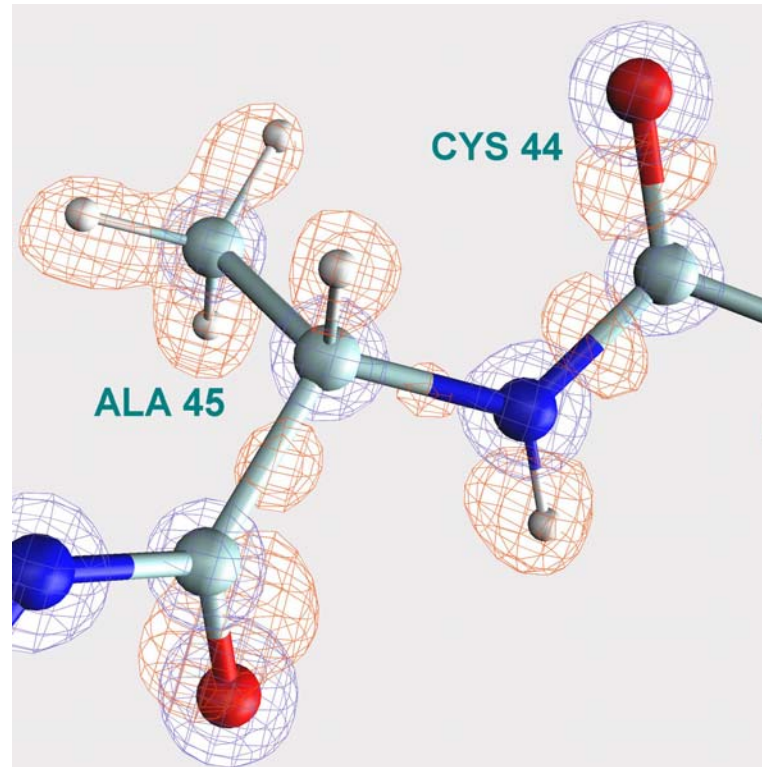
```
% phenix.refine model.pdb data_xray.hkl neutron_data.file_name=data_neutron.hkl  
input.xray_data.labels=FOBSx input.neutron_data.labels=FOBSn
```

- phenix.refine offers various options for handling H atoms:
 - Refine as riding model at any resolution
 - Refine as individual atoms (at ultrahigh resolution or using neutron data)
 - Account for scattering contribution or just use to improve the geometry
- Expected benefits from using the H atoms in refinement:
 - Improve R-factors
 - Improve model geometry
 - Model residual density at high resolution or in neutron maps
- Example from automatic re-refinement of 1000 PDB models with and without H

pdb	resolution	Rfree(no H) - Rfree(with H)
1dv7	1.3	2.47
1byp	1.8	1.51
1dkp	2.3	1.36
1rgv	2.9	2.01

- Subatomic resolution (higher than $\sim 0.9 \text{ \AA}$): bond densities and H atoms

Aldose Reductase (0.66 \AA resolution)

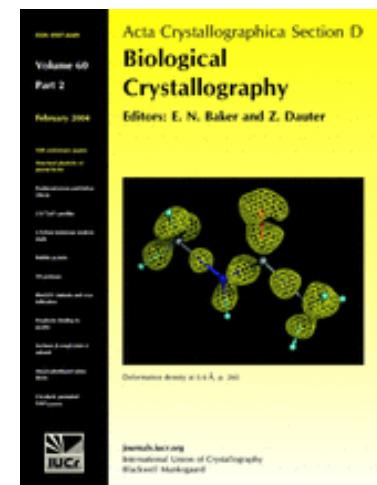


Fo-Fc (red)

0.28 e/\AA^3

2Fo-Fc (blue)

3.85 e/\AA^3



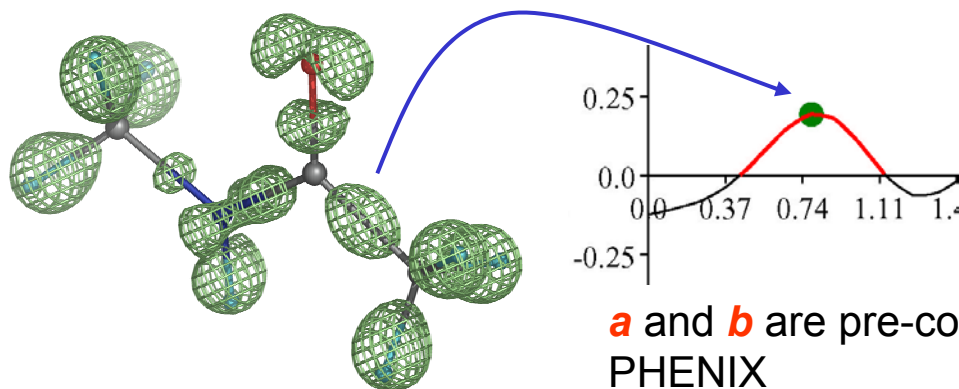
- Basics of IAS model:

Afonine *et al*, Acta Cryst. D60, 2004

- First practical examples of implementation and use in PHENIX:

Afonine *et al*, Acta Cryst. D63, 1194-1197

IAS modeling in PHENIX



Simple Gaussian is good enough:

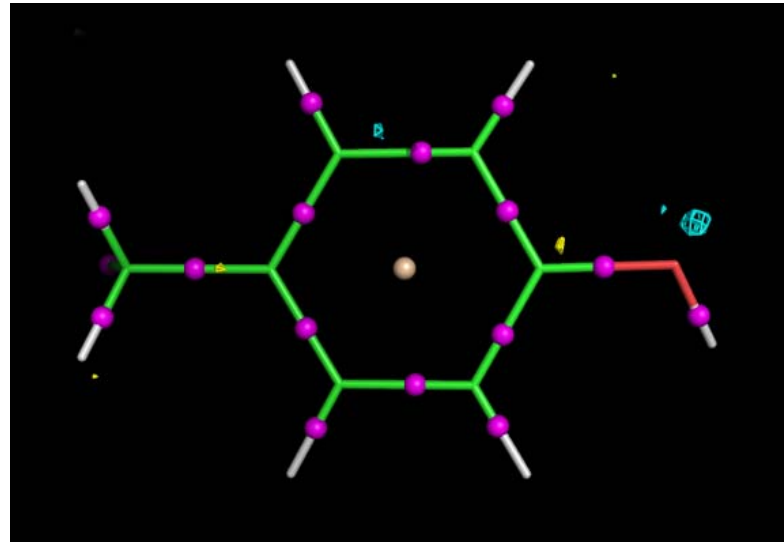
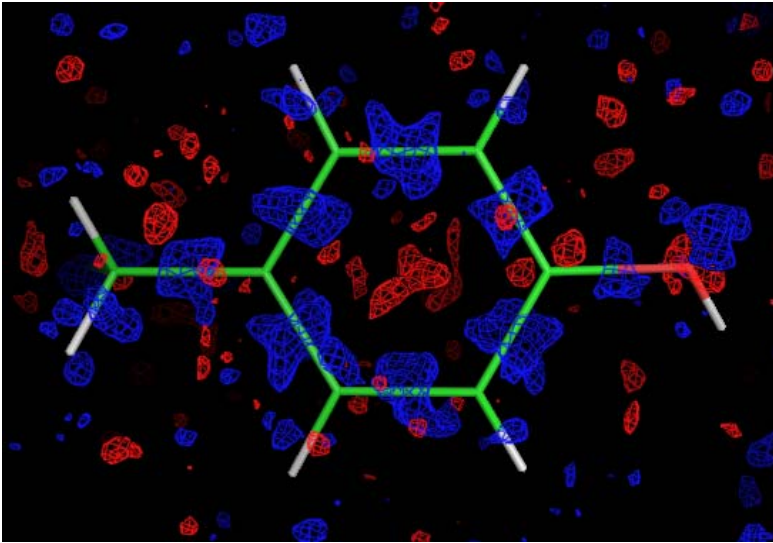
$$f_{bond_scatterer}(s) = a \exp\left(-\frac{b}{4} s^2\right)$$

a and **b** are pre-computed for most of bonds and implemented in PHENIX

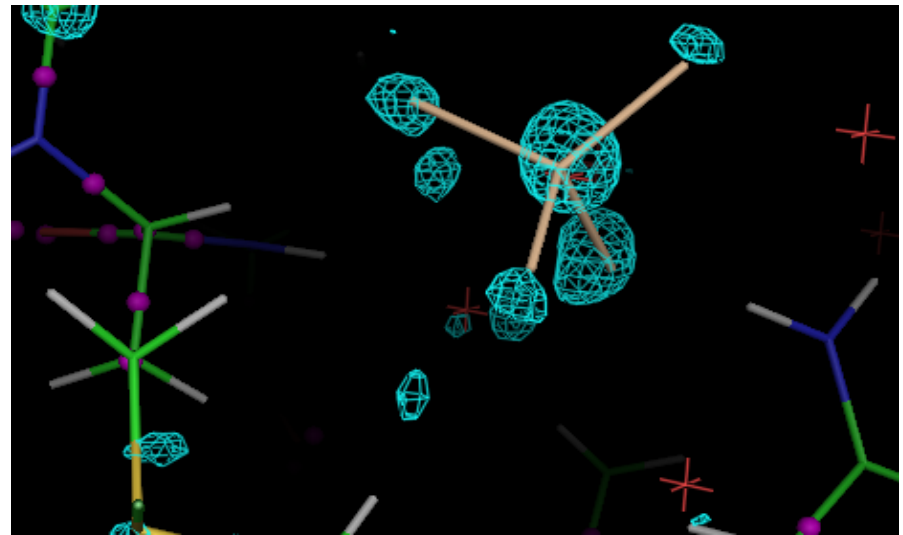
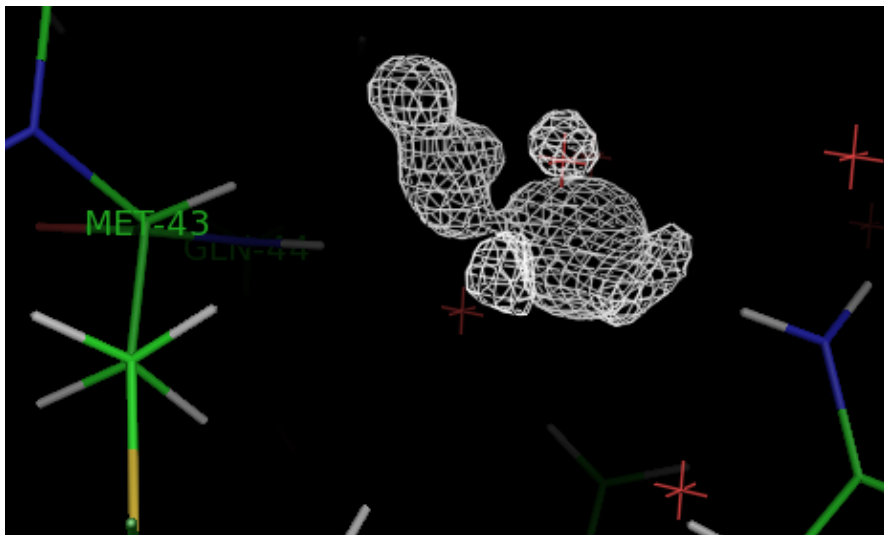
- Compared to Multipolar model that is commonly used at ultra-high resolutions, the new IAS model features:

- faster computations,
- no risk of overfitting,
- similar results as Multipolar model (R-factors, ADP, maps)

- Improve maps: reduce noise. Before (left) and after (right) adding of IAS.



- Find new features: originally wrong water (left) replaced with SO₄ ion (right) clearly suggested by improved map after adding IAS



- Real space refinement: fast ligand fitting, reduce amount of manual work
- Normal mode refinement: model more flexibility with general tools
- Torsion angle refinement: efficient low resolution refinement
- Automatic rigid groups search: fully automated TLS refinement
- GUI: ease navigation through refinement process
- Automatic H/D location at maps: enhance neutron and subatomic resolution refinement
- More automation

Availability

<http://www.phenix-online.org/>

Funding

- NIH / NIGMS [*P01GM063210, R01GM071939, P01GM064692*]
- LBNL [*DE-AC03-76SF00098*]
- PHENIX industrial consortium